

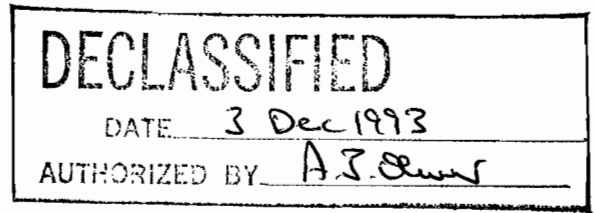
**MICROBIAL PLUGGING OF
URANIUM MINE TAILINGS TO
PREVENT ACID MINE DRAINAGE
FINAL REPORT**

MEND Project 2.44.1

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**MICROBIAL PLUGGING OF URANIUM MINE
TAILINGS TO PREVENT ACID MINE
DRAINAGE - FINAL REPORT**

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Environmental Laboratory

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MICROBIAL PLUGGING OF URANIUM MINE TAILINGS TO PREVENT ACID MINE DRAINAGE - FINAL REPORT

by

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ABSTRACT

Four test conditions and a set of controls were used to examine the best conditions for microbial cap formation. The following conditions were executed in triplicate (1) inoculation of three columns with Ultramicrobacteria (UMB's) and addition of liquid nutrient (BHI + sucrose); (2) inoculation with UMB's, liquid and solid nutrients (Stillage concentrate), (3) addition of the same as the previous set plus calcium and (4) only liquid nutrient was added. The control maintained a constant pH around 3, an Eh higher than 415 mV, an oxygen level always higher than 5.1 mg/L. However, the Fe content in general decreased over time which was unexpected. The series with UMB's, liquid and solid nutrients maintained a lower Eh than the other series for the longest period of time followed by the series of columns with liquid nutrient only. All conditions tested were generally better than the controls in increasing the pH and also in reducing the Eh of the effluent but nothing can be concluded with respect to the dissolved oxygen and iron contents due to continuous variations in the results. There was a significant decrease in the redox potential of the permeates, however none of the conditions tested completely reduced the permeability of the tailings over a long period of time. The scavenging of the oxygen and the increase in pH by the heterotrophs help reduce AMD generation by preventing growth of *Thiobacillus ferrooxidans*. The use of the UMB, *Klebsiella oxytoca* a heterotrophic strain, is not essential since the results of uninoculated columns with liquid nutrient only were equivalent to the columns with liquid nutrients and UMB's. It is likely that the nutrients increased the metabolic activity of the indigenous heterotrophs already present in the tailings.

Keywords: Acid Mine Drainage, Ultramicrobacteria, Microbial Plugging, Microbial Capping

PRÉVENIR LA FORMATION DE DRAINAGE MINIER ACIDE - RAPPORT FINAL

par

L. Lortie, M. Skaff et W.D. Gould

RÉSUMÉ

Quatre conditions expérimentales avec témoins ont été étudiées pour déterminer laquelle permettrait de mieux former une couche microbienne. Les analyses ont été effectuées en triplicata (1) trois colonnes ont été inoculées avec des ultramicrobactéries (UMB) et additionnées de nutriments liquides (BHI + sucrose); (2) trois ont été inoculées avec des UMB, et additionnées de nutriments liquides et solides (Concentré de brasserie); (3) trois ont été préparées comme (2) avec en plus du calcium et (4) trois ont été additionnées de nutriments liquides sans être inoculées. Le pH des témoins s'est maintenu à environ 3, le Eh à plus de 415 mV et le niveau d'oxygène à plus de 5.1 mg/L. Cependant, le contenu en Fe a en général diminué avec le temps, ce qui n'était pas prévu. Les colonnes inoculées et additionnées de nutriments liquides et solides ont conservé un Eh plus bas que celui des autres colonnes pour une plus longue durée de temps, suivi par les colonnes avec seulement des nutriments liquides. Toutes les conditions étudiées permettaient en général de mieux augmenter le pH et réduire le Eh de l'effluent que ne le faisaient les témoins, mais on ne peut pas tirer de conclusion pour l'oxygène dissout et le contenu en fer car les résultats variaient continuellement. Il y avait une baisse significative du potentiel d'oxydo-réduction des perméats, cependant aucune des conditions étudiées n'a complètement permis de réduire la perméabilité des résidus sur une longue période de temps. L'utilisation de l'oxygène et l'augmentation du pH par les hétérotrophes aident à réduire la production de DMA en empêchant la croissance de *Thiobacillus ferrooxidans*. L'utilisation d'UMB, *Klebsiella oxytoca* une souche hétérotrophe, n'est pas essentielle puisque les résultats des colonnes avec seulement des nutriments liquides, étaient semblables aux résultats obtenus avec les colonnes inoculées. Les nutriments augmentent probablement l'activité métabolique des bactéries hétérotrophes qui sont déjà présentes dans les résidus.

Mots-clés: Drainage minier acide, Ultramicrobactéries, Recouvrement bactérien

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OBJECTIVE

Previous results have shown that a bacterial cap can be formed consisting of more than 10^5 bacteria per gram of tailings and is capable of scavenging oxygen from the interstitial water. However, since it did not reduce the permeability of the coarse tailings, the cap was not considered completely satisfactory. The objective of the present study was to determine if a microbial cap capable of reducing both the penetration of oxygen and water can be formed. A successful cap would reduce both the water permeability of the tailings as well as the oxygen content of the permeate.

BACKGROUND

Acid mine drainage results from the production of H_2SO_4 in sulphide containing tailings derived from ores bearing pyrite and pyrrhotite minerals via chemical and biological processes that require the presence of oxygen. In natural soils, pyrite is not oxidized because of the vigorous growth of bacteria in the nutrient-rich surface horizons. The heterotrophic bacterial activity in natural soils scavenges oxygen that would otherwise be available for the oxidation of sulphide minerals. A strategy is proposed to develop an oxygen excluding surface layer (bacterial cap) on tailings by top dressing tailings with ultramicrobacteria (UMB's) and nutrients. Already, ultramicrobacteria have shown success in the plugging of permeable formations in oil reservoirs in order to prevent a breakthrough during water injection in secondary oil recovery. In the present study, the bacterial growth of UMB's would consume oxygen and the biomass produced would block the pore spaces. Oxygen diffusion would be decreased due to the reduced permeability so that deeper layers of soil become anaerobic, preventing growth of *Thiobacillus ferrooxidans*, which would oxidize metal sulphides to sulphuric acid and soluble metals. This would greatly reduce or prevent the oxidation of metal sulphides within the tailings and thereby prevent the formation of acid mine drainage (AMD). Once the cap has been developed, it is expected that cap integrity would be maintained for a period of six months to a year. Incremental nutrient additions would probably be necessary to maintain the microbial cap. If a vegetative cover was

established on the tailings, sufficient nutrients from the vegetation root exudates would be produced to maintain a metabolically active cap.

MATERIALS AND METHODS

Procedures

The experimental design and the procedures used in these studies were discussed with Mr. Ken Wheeland of Noranda and Dr. Ron McCready of C.A.R.E. before the experiments were initiated.

Tailings

Coarse oxidized uranium mill tailings from Denison Mines in Elliot Lake were tested. Their particle size distribution, general mineralogical and chemical properties were given by C.A.R.E. in the previous update report of September 1992 (Appendix - E). New tailings samples were also analyzed for calcium, iron, sulphate and sulphide contents. The results showed variations in the calcium content depending where the sample was taken. For example a sample taken at the top of the bucket had an average of 0.27 % Ca by weight, if taken in the middle 0.57% and at the bottom 0.19%. The average of these results was found to be 0.34% which is very close to the C.A.R.E result of 0.28%. The percentage by weight of Fe, S and SO₄ were 1.0, 1.3 and 2.2%, respectively.

Set-up of Columns

Five series of different conditions performed in triplicate including a series of controls were tested to verify the formation of the best cap. A total of fifteen columns (6.2 cm I.D. x 31 cm height) were filled with 1.5 kg wet weight of unoxidized tailings. The bottom of each column was packed with quartz (+ ¼ - 3/8") and glass wool to retain the tailings in the columns. Each column was prepared independently. A slurry consisting of 500 mL tap

water and 1.5 kg tailings was mixed and poured into each column. Another 500 mL was used to rinse residual tailings in the bucket. Columns which did not visually appear to be homogeneous were repacked. When all the columns were packed, water was added to the top of the tailings and allowed to settle overnight. The water was completely drained to remove any oxidation products and thereafter water was constantly kept on the tailings until the beginning of the experiments.

The day before the inoculation of the columns, a volume of 200 mL of tap water was added to each column and allowed to drain. Samples were taken at specific times in order to determine their initial permeability prior to the addition of bacteria and nutrients.

Inoculation and Nutrient Feeding

A volume of 100 mL of bacterial solution of UMB's (10^8 starved bacteria/mL of *Klebsiella oxytoca*) prepared by C.A.R.E was added to columns 4 to 12 while 100 mL of water was added to column series (1, 2, 3) and (13, 14, 13) on May 27th. The columns were all drained until the inoculum or water level was below the surface of the tailings then nutrients were added on May 28th. A volume of 100 mL of liquid nutrients (Brain Heart Infusion broth (BHI) 1/10 + sucrose (1%)) was added to column series 4, 5, 6 and 13, 14, 15. To columns 7, 8, and 9, 100 mL of a preparation of liquid nutrients and solid nutrient (Stillage concentrate 0.2%, pH 3.5 with BHI final pH is 7.3). One hundred mL of the preceding nutrients with CaSO_4 (17%) in addition was poured into columns 10, 11 and 12. A chemical analysis of the stillage concentrate used as the solid nutrient was performed as mentioned in the proposal. The acetic acid content was 2.3 g/L, lactic acid content 7.6 g/L, propionic acid 3 g/L, and organic carbon 42 g/L. The percentage of calcium, nitrogen and sodium were respectively 0.04, 0.21 and 0.03%.

The nutrients were all drained until they reached the surface level of the tailings before shutting off the clamps. Once all clamps were closed, 5 mL of water was added to the top of the tailings in each column. Thus, the surface of the tailings was consistently kept

moist for the duration of the experiments. Another nutrient addition was carried out one week after the initial addition on June 4th, this day corresponded to the first day of measurements. Oxygen permeability was monitored by measuring Eh and dissolved oxygen. Water quality was monitored by pH and total iron. At this time no more calcium was added to columns 10, 11, and 12. Nutrients were also added on July 29th, August 26th and September 16th.

Precipitation Events

A volume of 100 mL of tap water was added to each of the columns, June 17th, three weeks after the initial inoculation to simulate precipitation events. Once all columns were set, a timer was started to measure the flow rate. The volume of permeate recovered during the time elapsed for the water level to reach the surface of the tailings was used to calculate the flow rate in mL/min. This procedure was carried out on a weekly basis. The permeates were collected in narrow-mouth bottles to minimize infiltration of atmospheric O₂. Once the water was drained, the dissolved oxygen was immediately measured with a portable oxygen probe (Orion Research Model # 97-08) followed by the measurement of Eh and pH. The permeates were then acidified with H₂SO₄ to stabilize the iron. The total iron was determined by Atomic Absorption Spectrophotometry.

RESULTS AND DISCUSSION

The project began on May 28th with the inoculation and the first addition of nutrients to the columns (Table 1). Results were recorded from June 4th until September 16th, they are listed in Tables 2 to 6. The minutes from the meeting held on September 30th are provided in Appendix - A. The economics of the process written by Dr. Ron McCready is in Appendix - B. Table 7 in Appendix - C includes the averages and standard deviations used for Figures 1 to 16. In order to improve clarity and ease of interpretation the error bars are only included in the figures when the experimental values are significantly different from the controls. Graphs comparing different treatments to the controls for each parameter tested are

given in Appendix - D. Appendix - E is a summary of results for each of the columns versus time.

Two of the treatments; one with UMB's plus liquid and solid nutrients (Figure 2) and the other one with liquid nutrient alone (Figure 4) both caused a consistent increase in effluent pH. An increase in effluent pH was observed between days 20 to 55 for UMB's plus liquid (Figure 1) and between days 20 to 40 followed by a decrease and increase again between days 62 to 83 for UMB's plus liquid and solid nutrients and calcium (Figure 3). Except for the addition of the UMB's plus nutrients and calcium (Figure 7) a consistent decrease in Eh was observed for all of the treatments (Figures 5, 6 and 8). No significant difference in permeability (measured flow rate) was observed between the controls and the various treatments (Figures 9, 10, 11 and 12). Although no consistent decrease in oxygen was observed for the various treatments, all of them did show significant lower dissolved oxygen values than the controls at the end of the experiment (Figures 13, 14, 15 and 16). The soluble iron in the effluent from the controls and all of the treatments decreased during the course of the experiment except for column 4, 5 and 6 (Figures 17, 18, 19 and 20). No differences were observed between the control columns and the various treatments with respect to the total iron contents of the effluent.

Most of the data shows high variability for most of the experimental time period. In future work it would be important to closely control the level of the water table and the timing of nutrient addition. Only the pH and Eh were significantly affected by the treatments of the parameters measured. Some decrease in dissolved oxygen was observed for the nutrient treatments at the end of the experiment. Of the two major criteria for successful cap formation (1) decreased permeability and (2) oxygen scavenging by the bacteria only the latter criterion was attained with some degree of success. The addition of liquid nutrients alone was equally as successful as the treatment in which UMB's were also added. It appears that the addition of UMB's is not necessary as the indigenous bacterial population will carry out the same function. The treatment costs will be significantly lower if a bacterial inoculant does not need to be added to the tailings.

CONCLUSIONS

It was not demonstrated that a microbial cap could significantly reduce the permeability of the tailings. Removal of oxygen from the permeates was noticeable as Eh values as low as -181 mV and low dissolved oxygen values were obtained. The results obtained from the inoculated columns with added nutrients were equivalent to the columns with nutrients plus UMB's. The addition of UMB's to the tailings is not necessary because indigenous bacteria are also able to colonize the surface of the tailings. It would be less expensive if the inoculation step could be avoided. Although a reduction in permeability was not observed in our experiments, it is possible that in fine-textured tailings a significant reduction in water permeability would be observed.

RECOMMENDATIONS

1. Microbial plugging of fine tailings should be evaluated further with the following experimental modifications:
 - (a) Use of humidity cells;
 - (b) Experimentation with different acid generating tailings;
 - (c) Longer term experiments;
 - (d) More frequent nutrient addition; and
 - (e) Analyze sulphate content of leachates.

2. Because microbial plugging of tailings is presently being investigated further by private industry we recommend that no further studies be undertaken by CANMET at the present time.

Table 1. Project schedule

Dates	Inoculation and feeding days	# Days after first sampling	# Weeks since inoculation
May 28	Inoculation and first feeding		0
June 4	Feeding and beginning of sampling	0	1
June 17		13	3
June 24		20	4
July 2		28	5
July 8		34	6
July 15		41	7
July 22		48	8
July 29	Feeding	55	9
August 5		62	10
August 12		69	11
August 19		76	12
August 26	Feeding	83	13
September 2		90	14
September 9		97	15
September 16	Feeding	104	16

Table 2. Data summary for columns 1, 2, 3

CONTROLS															
DAYS	RATE (mL / min.)			OXYGEN (ppm)			Eh (mV)			pH			Fe (ppm)		
	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
0	1.1	0.6	1.1	5.17	6.63	6.13	452	456	467	2.73	2.58	2.77	182.0	395.0	72.0
13	0.6	0.6	0.5	7.67	7.67	6.14	438	438	441	2.77	2.77	2.67	176.0	261.0	187.0
20	1.3	1.6	1.5	7.76	5.80	7.35	430	451	453	2.79	2.86	3.54	124.0	175.0	129.0
28	1.0	1.0	1.6	7.69	5.54	6.11	425	435	441	2.82	2.63	2.81	117.0	170.0	116.0
34	1.2	1.3	1.3	6.45	5.63	6.38	437	459	469	2.77	2.52	2.85	126.0	233.0	97.0
41	1.6	1.7	1.3	5.53	6.96	6.23	445	417	443	2.49	2.69	2.91	248.0	146.0	93.0
48	1.4	1.4	1.3	6.18	7.40	7.08	434	429	432	2.40	2.59	2.73	131.0	240.0	93.0
55	1.7	1.8	1.5	5.70	6.55	5.78	437	434	431	2.65	2.72	2.85	177.0	92.0	88.0
62	1.7	1.0	1.4	6.40	7.67	6.38	444	446	455	2.70	2.80	2.68	84.4	60.5	106.0
69	2.2	0.8	1.5	6.14	8.40	6.05	486	503	505	2.79	2.72	2.69	52.9	56.6	94.0
76	2.0	0.8	1.3	6.28	6.26	5.94	437	445	451	2.94	2.91	2.72	35.8	43.6	115.0
83	2.0	1.1	1.7	5.35	6.76	5.76	436	443	451	2.99	3.09	2.74	26.2	17.2	92.2
90	1.8	0.7	1.2	7.24	6.64	6.35	430	441	459	3.00	3.18	2.77	30.8	16.3	93.5
97	1.6	0.6	1.2	7.14	8.19	6.75	424	456	466	3.04	3.28	2.85	28.6	7.7	69.5
104	2.0	0.4	1.3	6.56	7.92	6.75	441	533	570	3.02	3.15	2.83	26.2	8.9	55.0

Table 3. Data summary of columns 4, 5, 6

UMB + LIQUID NUTRIENTS															
DAYS	RATE (mL / min.)			OXYGEN (ppm)			Eh (mV)			pH			Fe (ppm)		
	4	5	6	4	5	6	4	5	6	4	5	6	4	5	6
0	0.0	0.1	0.1	7.80	7.80	7.80	475	511	451	2.41	2.57	2.88	393.0	259.0	189.0
13	0.5	0.1	0.1	6.27	NA	NA	338	NA	NA	2.90	NA	NA	154.0	62.0	57.0
20	0.2	0.0	0.6	5.67	6.40	6.73	355	403	280	3.52	2.85	3.45	56.0	42.0	34.0
28	0.1	0.0	0.4	6.63	6.85	7.15	127	180	103	4.57	4.33	4.71	177.0	44.0	35.0
34	0.2	0.2	0.6	4.49	5.13	6.57	44	113	40	5.11	5.15	5.35	214.0	104.0	95.0
41	0.0	0.8	0.6	4.38	2.56	4.43	-46	-75	69	6.05	6.24	4.84	87.0	35.0	278.0
48	0.0	0.8	1.4	6.40	4.02	3.64	-35	-54	379	7.11	6.15	2.35	27.5	68.0	505.0
55	0.4	1.2	1.3	4.04	4.68	1.26	-49	391	4	6.32	5.55	2.43	36.0	44.0	720.0
62	1.0	0.9	0.9	5.19	6.03	0.55	-26	242	434	6.22	3.22	2.60	51.4	370.0	408.0
69	0.5	0.6	2.0	5.88	4.63	4.48	-53	388	387	5.83	2.80	2.78	214.0	490.0	262.0
76	0.8	0.5	0.9	5.32	1.32	4.23	-82	416	393	5.27	2.65	2.90	312.0	310.0	109.0
83	0.7	1.1	1.4	5.13	4.64	6.00	-72	368	307	5.64	2.57	3.17	33.3	122.0	55.6
90	1.2	0.5	2.0	4.02	4.33	5.83	-50	393	283	6.19	2.42	3.30	16.5	298.0	110.0
97	1.1	0.3	1.6	3.28	5.18	4.13	-121	420	20	5.68	2.56	4.87	28.0	282.0	197.0
104	1.2	0.3	1.9	3.70	1.48	4.11	-46	433	15	5.06	2.76	5.06	99.0	202.0	167.0

Table 4. Data summary for columns 7, 8, 9

UMB - LIQUID & SOLID NUTRIENTS															
DAYS	RATE (mL / min.)			OXYGEN (ppm)			Eh (mV)			pH			Fe (ppm)		
	7	8	9	7	8	9	7	8	9	7	8	9	7	8	9
0	0.4	1.3	0.0	6.09	5.70	7.03	451	413	569	2.69	2.80	2.57	269.0	282.0	187.0
13	1.9	1.5	0.0	6.06	6.78	NA	327	297	NA	2.85	3.03	NA	98.0	102.0	NA
20	0.8	0.8	0.0	7.05	7.82	NA	238	195	NA	3.59	4.01	2.47	37.0	49.0	157.0
28	0.4	0.5	0.0	7.73	7.89	5.45	18	44	320	4.65	4.71	2.96	103.0	101.0	0.7
34	0.6	0.6	0.0	4.97	6.89	0.73	2	56	-97	5.31	5.10	6.69	113.0	166.0	4.3
41	0.7	0.8	0.0	4.95	4.45	5.73	-60	-55	117	6.15	5.76	7.54	54.0	144.0	5.1
48	1.4	0.9	0.0	4.62	5.11	0.39	-44	-34	-126	6.16	6.02	1.53	36.5	79.0	14.5
55	1.9	0.7	0.0	4.49	6.12	1.38	-35	8	61	4.95	5.94	6.92	161.0	81.0	8.5
62	2.0	0.6	0.7	6.53	7.23	6.25	42	-39	-38	4.12	5.91	6.45	230.0	65.1	14.3
69	2.2	0.9	0.8	4.86	4.50	5.22	-135	-136	-59	4.86	5.57	6.31	109.0	47.0	51.0
76	1.3	0.8	0.9	2.41	4.81	4.68	-142	-119	-66	4.96	5.01	5.92	66.0	65.0	94.0
83	1.2	0.7	1.1	2.87	3.06	6.56	-158	-147	-41	5.49	5.51	7.28	3.3	20.0	6.3
90	2.2	1.1	0.4	4.36	2.10	7.22	-160	-165	-52	5.66	5.68	6.89	3.3	8.3	13.5
97	0.9	0.9	0.5	5.84	5.58	6.67	-150	-126	10	5.84	5.58	5.76	2.1	6.8	97.5
104	1.3	1.0	0.6	4.13	3.96	2.38	-72	-74	-7	5.05	4.91	5.27	29.8	33.7	220.0

Table 5. Data summary for columns 10, 11, 12

UMB + LIQUID & SOLID NUTRIENTS + CALCIUM															
DAYS	RATE (mL / min.)			OXYGEN (ppm)			Eh (mV)			pH			Fe (ppm)		
	10	11	12	10	11	12	10	11	12	10	11	12	10	11	12
0	0.2	1.2	0.1	7.80	5.52	8.83	467	454	548	2.80	2.82	2.76	146.0	103.0	57.0
13	0.4	1.6	1.7	7.73	5.22	7.29	446	505	362	2.71	2.56	2.91	86.0	96.0	18.0
20	0.3	1.7	0.1	8.37	3.97	8.36	260	284	393	3.02	3.22	3.31	19.0	44.0	6.0
28	1.0	1.0	0.3	7.53	7.31	6.80	186	126	110	4.07	4.51	4.72	21.0	84.0	11.0
34	0.5	0.6	0.3	7.37	6.03	6.65	210	51	67	4.85	5.35	5.28	52.0	101.0	13.0
41	2.2	1.6	0.0	4.83	3.78	3.31	-56	-38	-68	5.42	5.70	7.55	112.0	188.0	6.6
48	1.4	0.8	0.7	4.41	5.33	7.24	-21	14	-8	5.58	5.29	5.85	189.5	270.0	56.5
55	1.3	1.1	1.0	6.25	6.32	3.90	75	270	442	4.59	3.05	2.93	229.0	101.0	95.0
62	2.1	0.4	0.1	5.04	4.93	6.62	382	404	547	3.28	3.07	2.73	135.0	184.0	43.7
69	1.6	2.3	0.1	5.44	5.07	8.05	-8	29	520	4.06	3.31	2.82	116.0	115.0	40.7
76	1.5	1.9	0.4	3.84	4.24	6.98	-86	-83	253	4.94	4.62	3.96	346.0	172.0	33.2
83	1.8	1.7	1.6	3.62	2.91	4.10	-22	-75	3	5.49	5.29	5.33	65.1	182.0	103.0
90	1.6	1.5	1.1	1.73	1.85	5.43	-181	-92	-2	6.26	5.92	5.53	5.6	24.0	175.0
97	1.4	1.2	0.8	2.15	3.87	6.47	-160	-125	30	6.27	5.53	4.99	3.4	14.3	224.0
104	1.6	1.2	2.0	1.08	4.03	3.75	-138	-32	23	5.42	4.91	4.89	30.0	81.5	260.0

Table 6. Data summary for columns 13, 14, 15

LIQUID NUTRIENTS															
DAYS	RATE (mL / min.)			OXYGEN (ppm)			Eh (mV)			pH			Fe (ppm)		
	13	14	15	13	14	15	13	14	15	13	14	15	13	14	15
0	0.3	1.3	0.8	6.09	6.19	5.91	473	155	475	2.87	2.74	2.87	122.0	390.0	77.0
13	1.7	1.7	0.7	6.67	6.83	5.88	404	403	402	3.06	3.01	3.06	150.0	332.0	142.0
20	0.7	1.6	0.7	7.12	7.35	7.12	386	375	386	3.34	3.33	3.34	148.0	267.0	65.0
28	1.9	1.6	1.0	5.80	6.70	7.36	361	244	228	3.47	4.37	4.13	50.0	166.0	62.0
34	1.7	1.3	1.2	5.83	6.85	6.84	325	163	79	3.58	5.31	4.59	43.0	180.0	89.0
41	1.7	1.5	1.7	5.94	6.62	5.62	90	-15	60	3.44	6.36	4.25	60.0	49.0	169.0
48	0.2	1.3	1.4	7.02	5.31	5.59	79	-69	414	3.38	6.55	2.44	99.0	65.0	310.0
55	0.1	1.5	1.5	7.90	4.96	3.91	339	-67	420	3.88	6.38	2.52	138.0	62.0	104.0
62	0.5	2.2	1.8	6.83	6.39	4.04	24	-33	447	5.40	6.52	3.58	162.0	31.7	313.0
69	0.7	2.2	1.6	6.62	5.32	5.97	-68	-116	391	5.49	6.43	2.85	140.0	39.4	209.0
76	0.6	1.9	1.5	3.66	5.60	5.01	-127	-111	392	5.13	5.91	2.94	97.0	108.0	61.8
83	0.7	2.1	1.8	2.82	2.38	4.47	-138	-52	11	5.69	5.63	3.74	17.4	50.0	32.1
90	0.8	1.6	2.0	3.98	2.58	3.26	-117	-178	-103	6.08	6.12	4.98	5.1	3.9	59.0
97	0.6	1.4	1.5	2.05	3.48	2.65	-161	-168	-119	5.59	6.30	5.75	8.4	3.8	57.5
104	0.9	1.8	1.7	0.32	4.37	0.78	-99	-107	-154	4.83	5.33	5.44	55.8	29.4	61.5

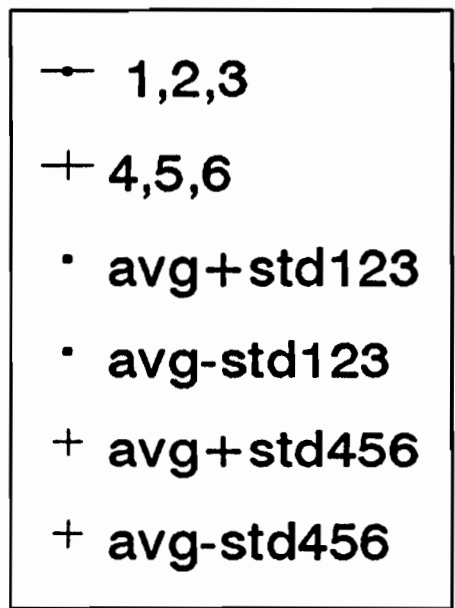
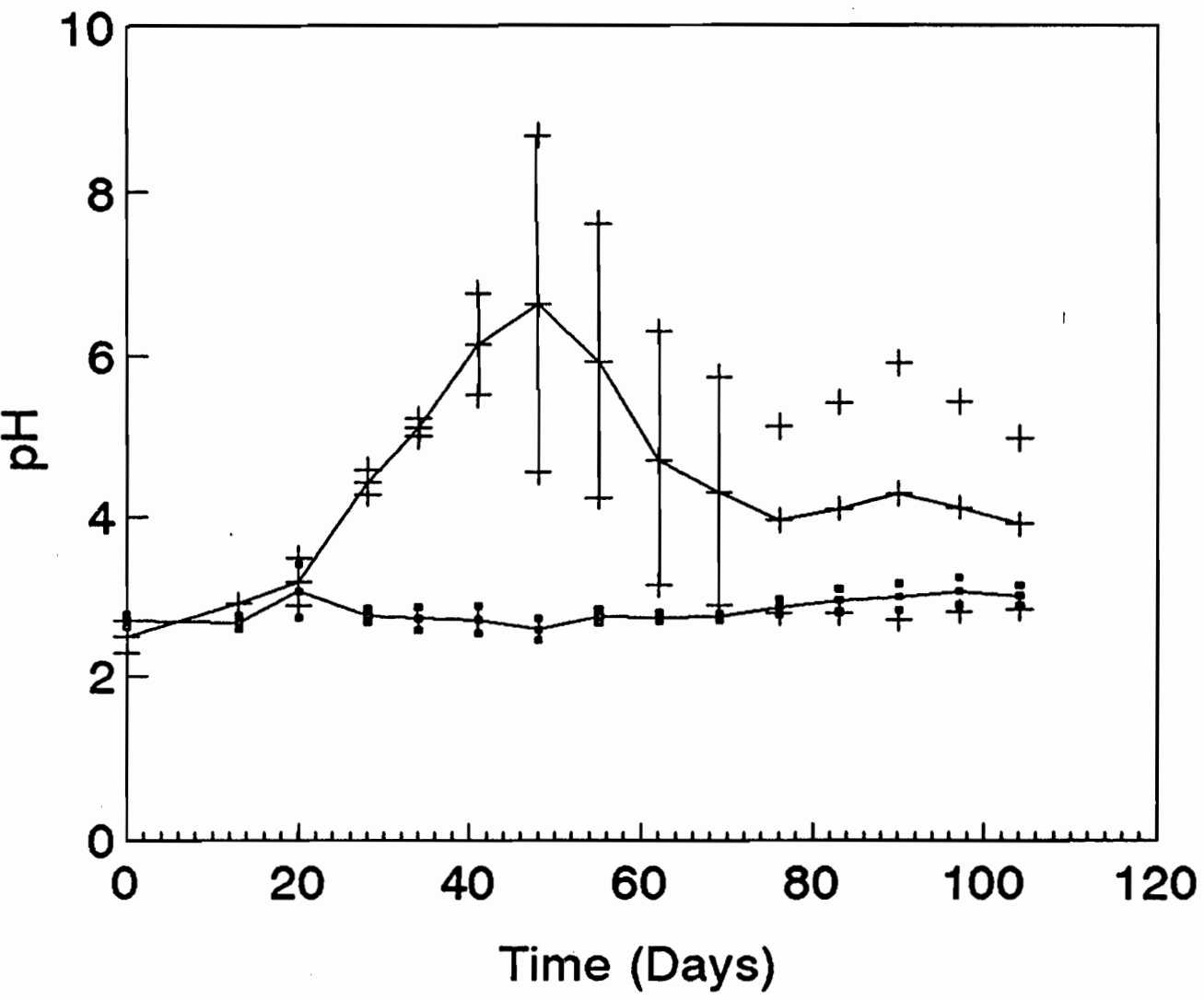


Figure 1. Control pH average compared to columns 4, 5, 6

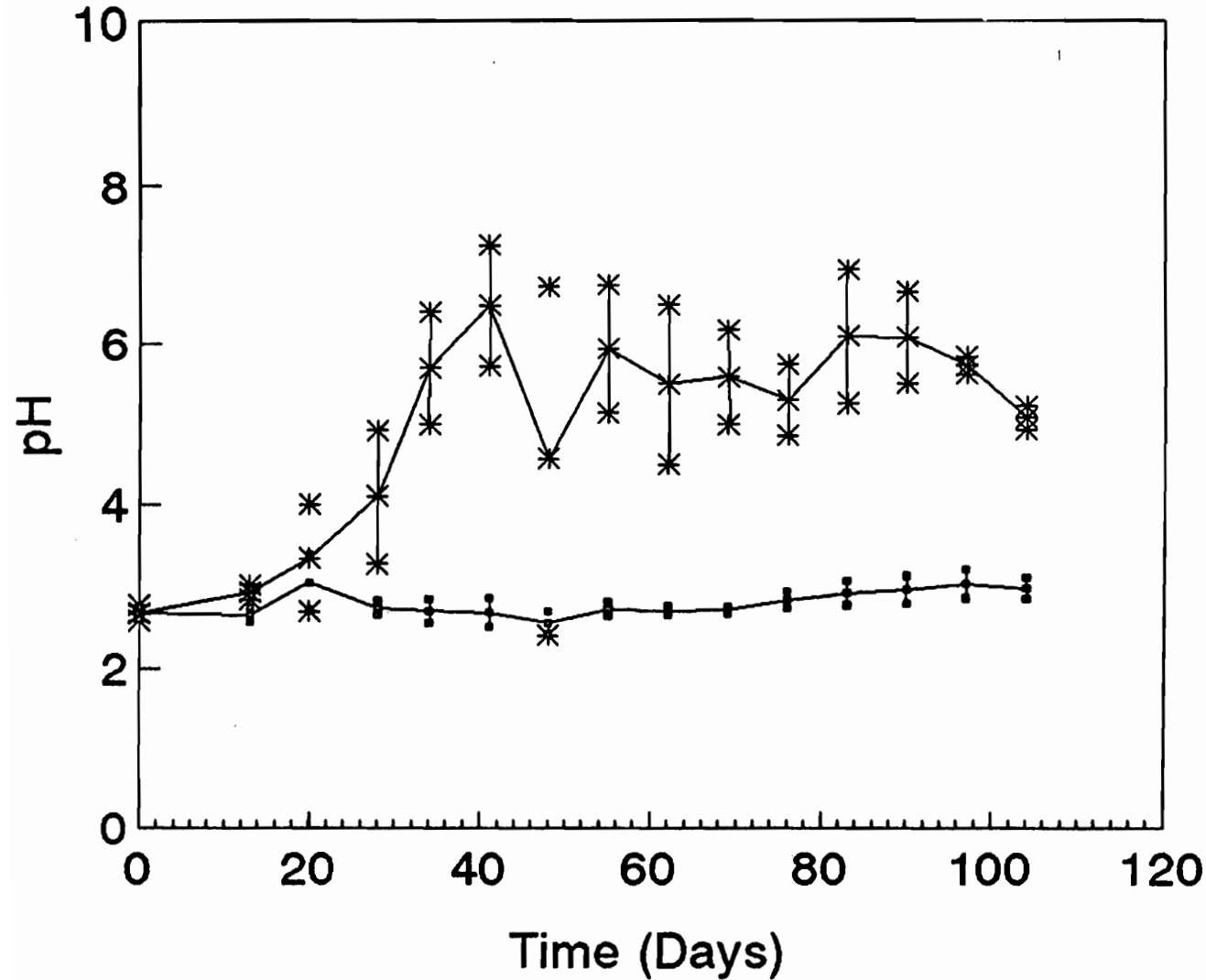
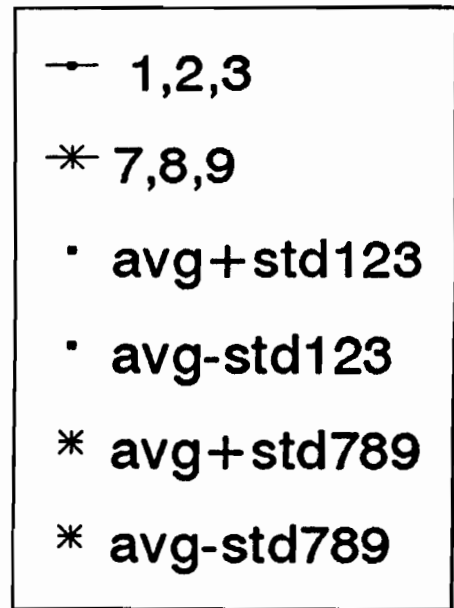


Figure 2. Control. pH average compared to columns 7, 8, 9



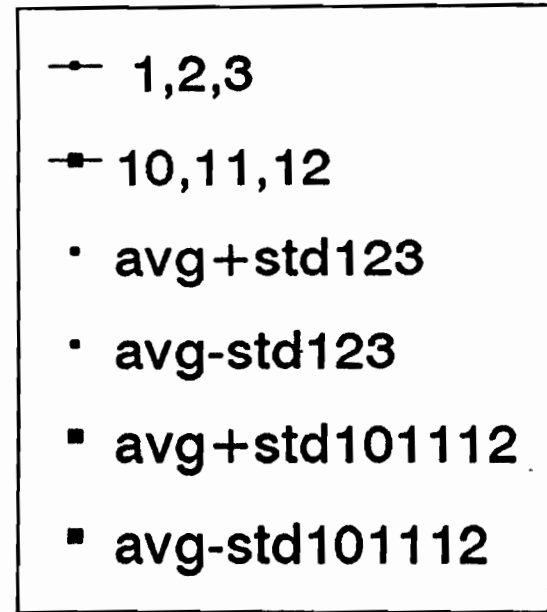
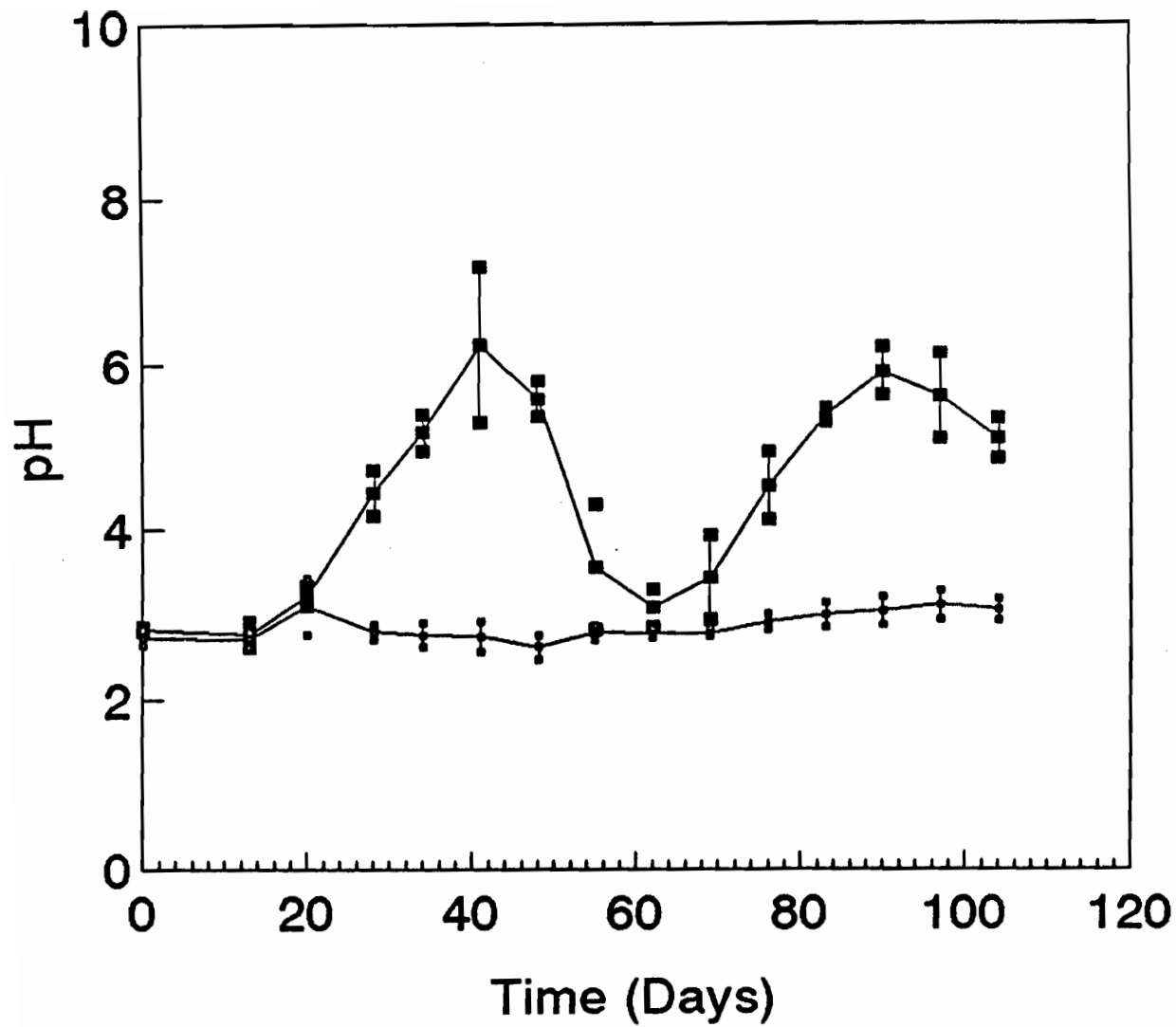


Figure 3. Control pH compared to columns 10, 11, 12

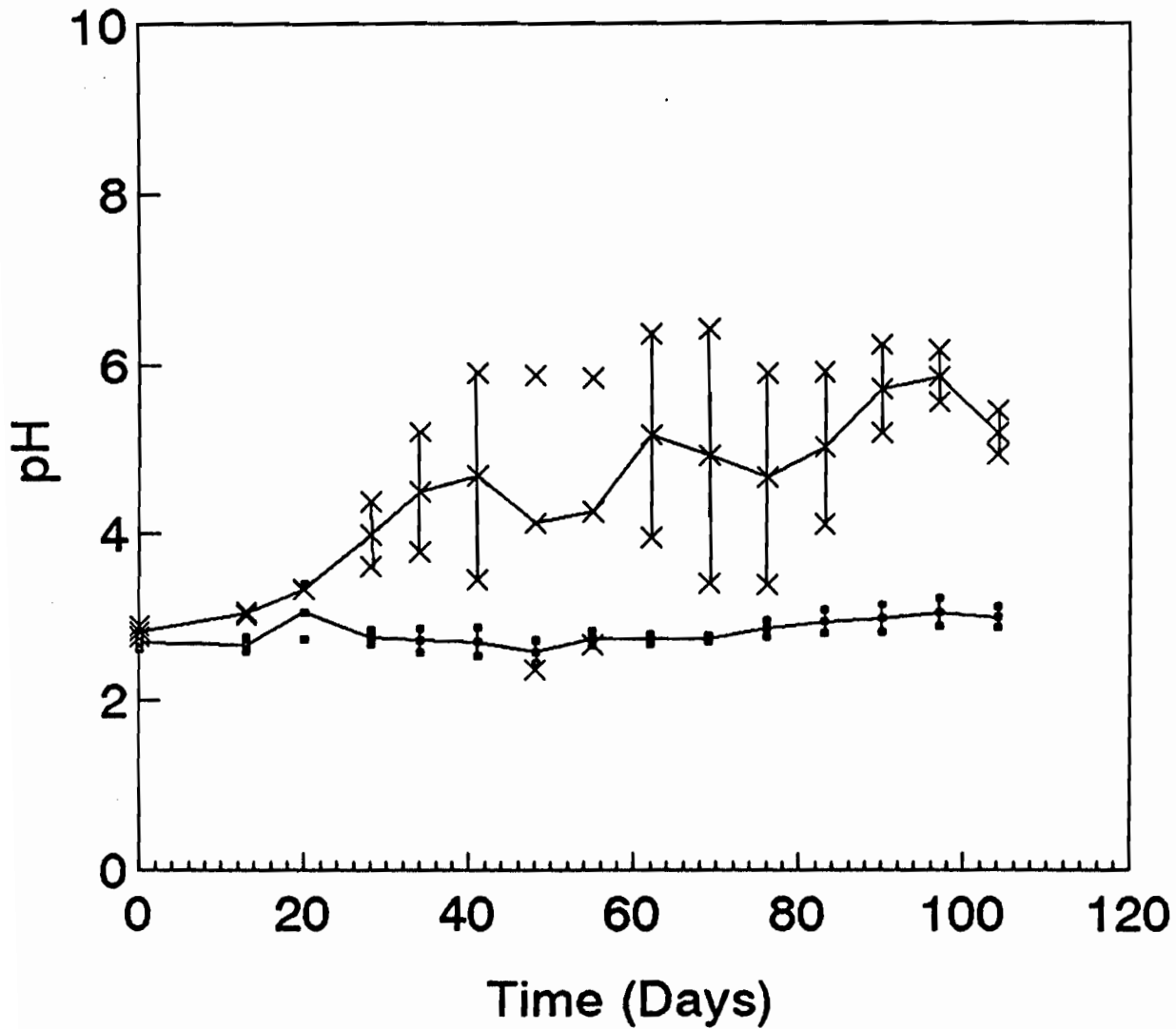


Figure 4. Control pH compared to columns 13, 14, 15

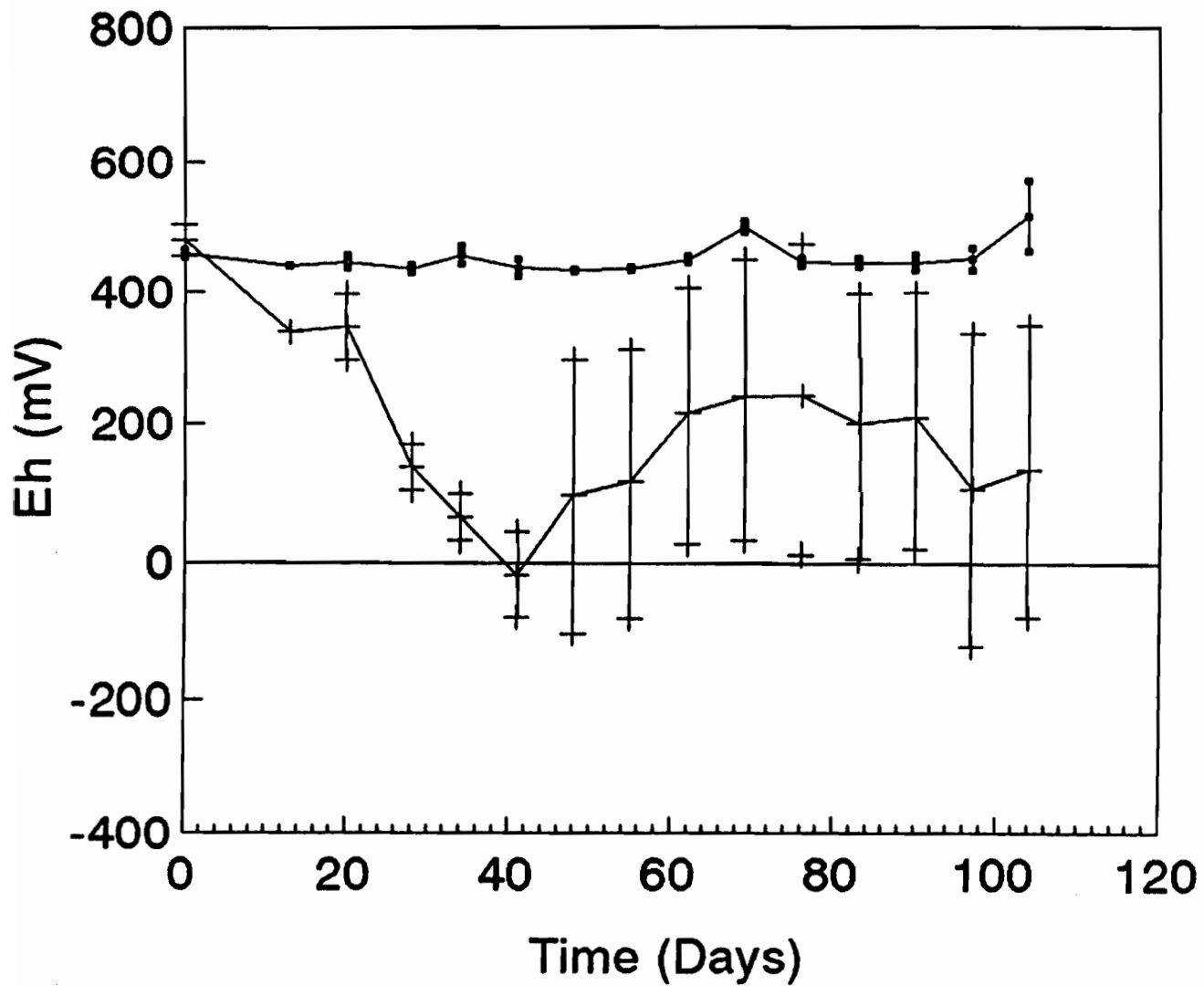


Figure 5. Control Eh average compared to columns 4, 5, 6

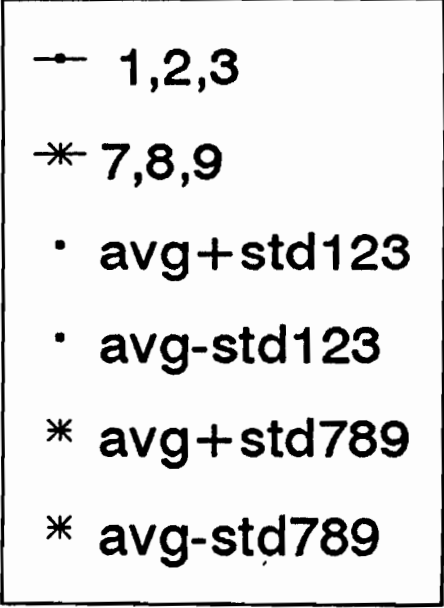
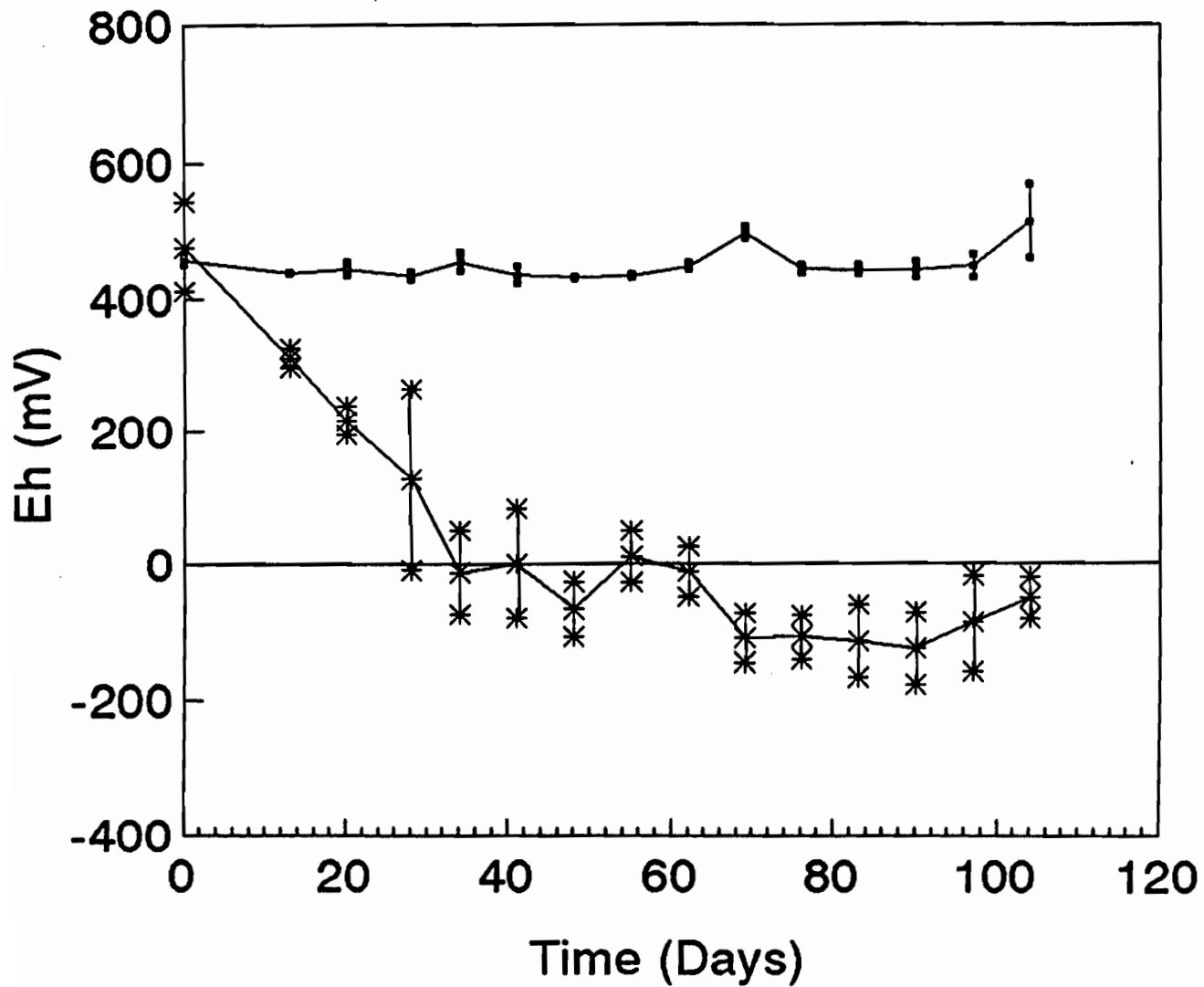


Figure 6. Control Eh average compared to columns 7, 8, 9

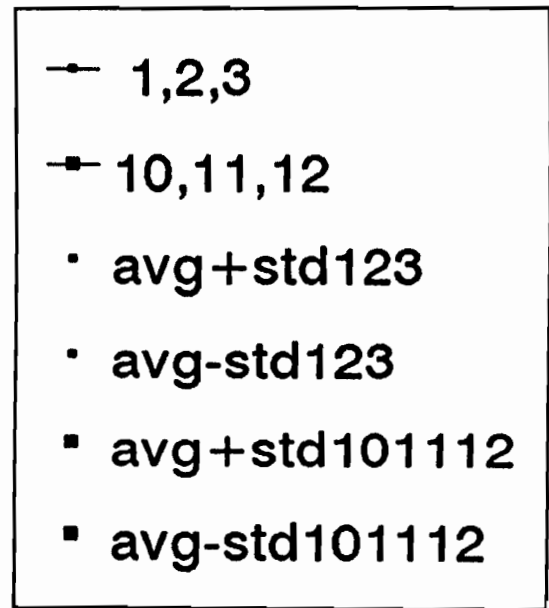
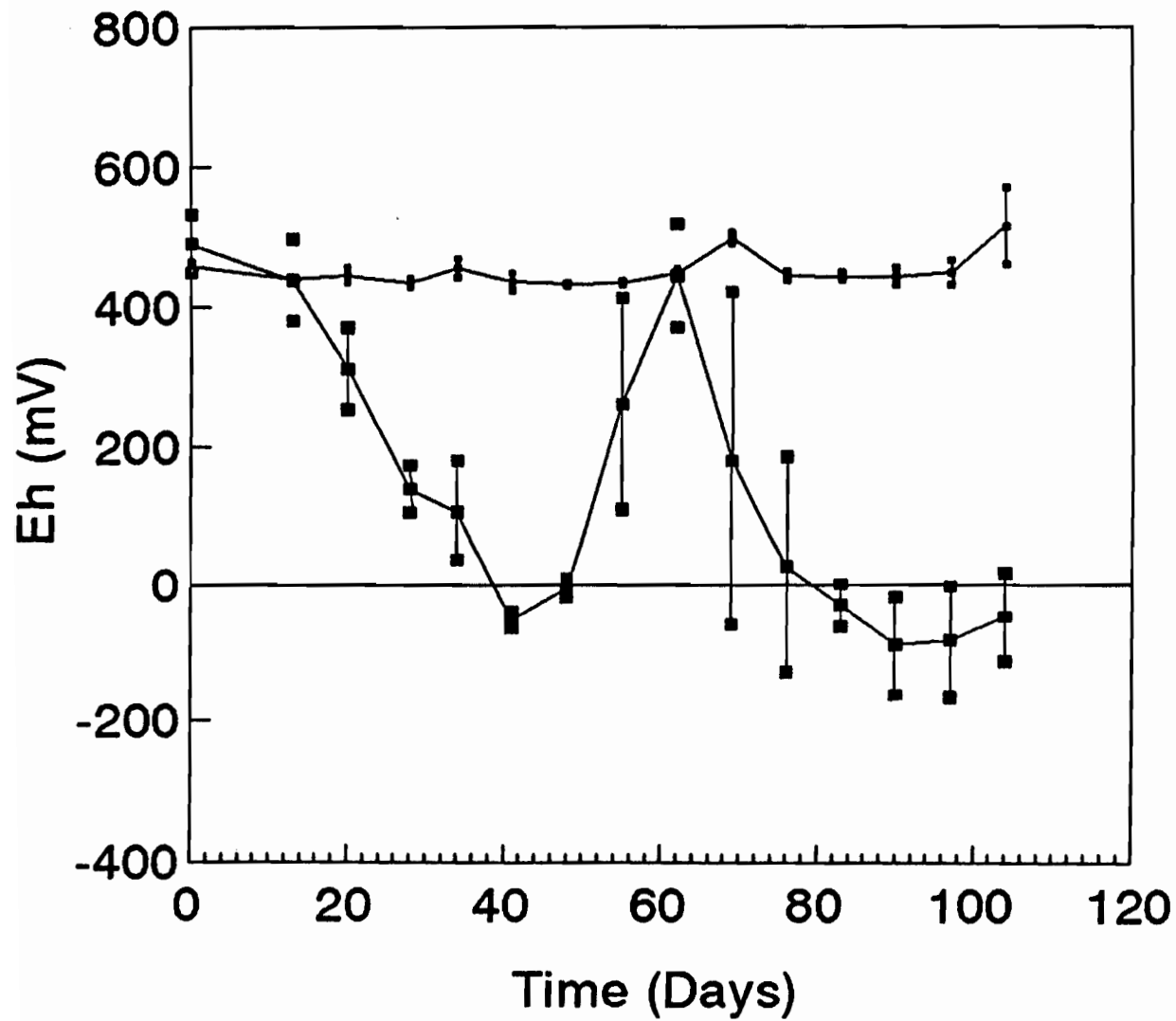


Figure 7. Control Eh average compared to columns 10, 11, 12

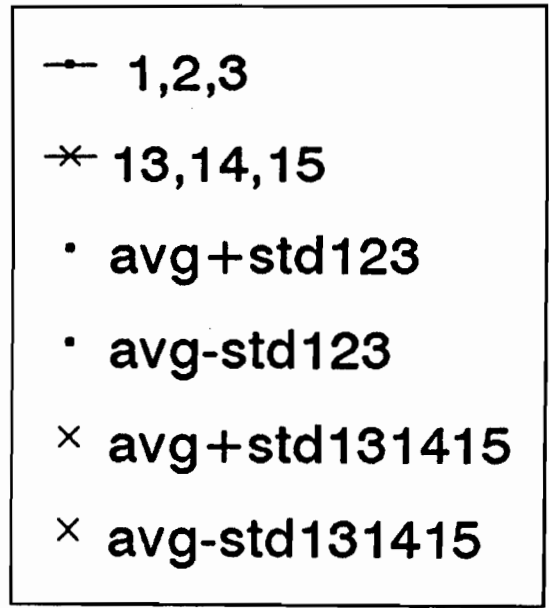
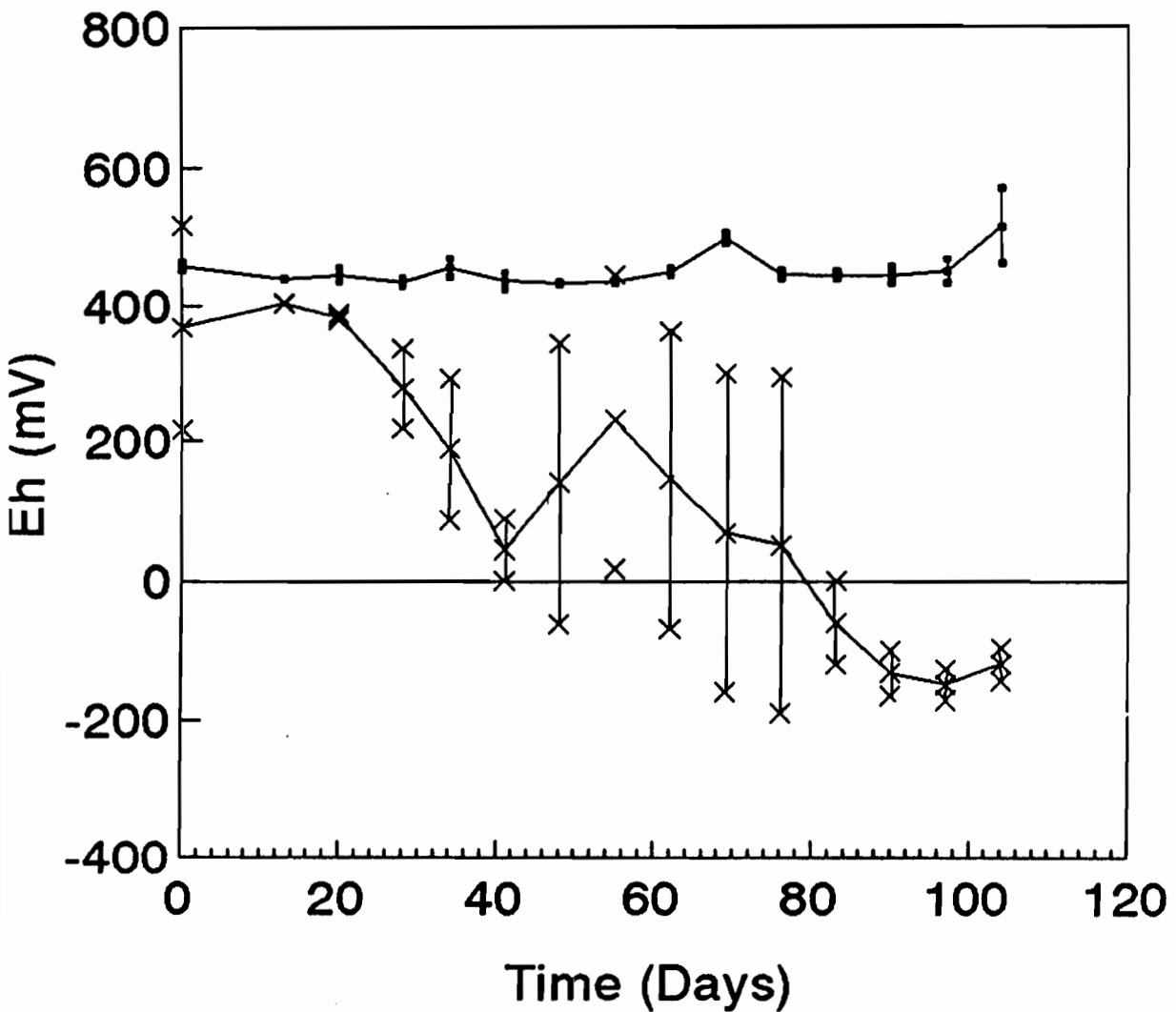


Figure 8. Control Eh average compared to columns 13, 14, 15

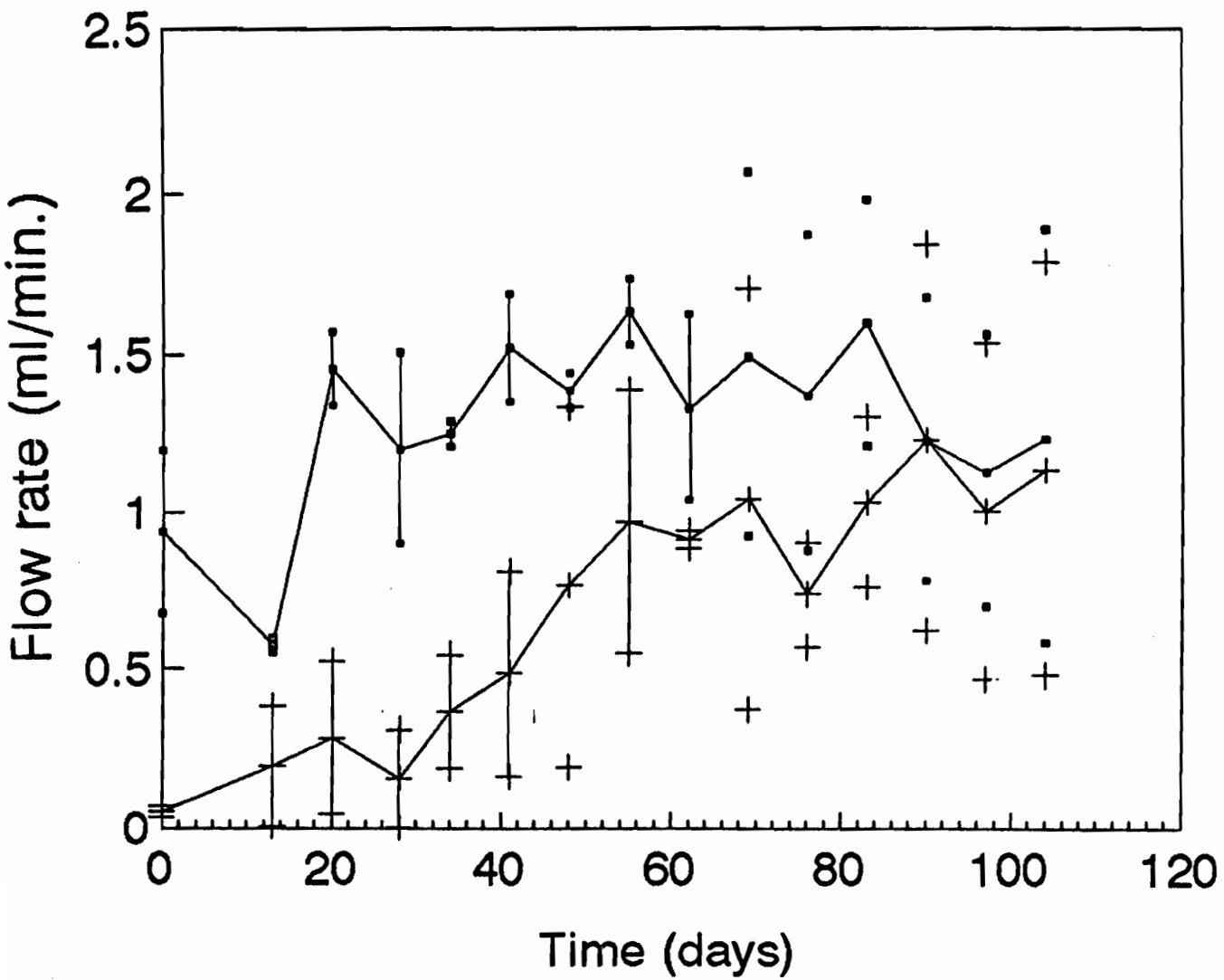
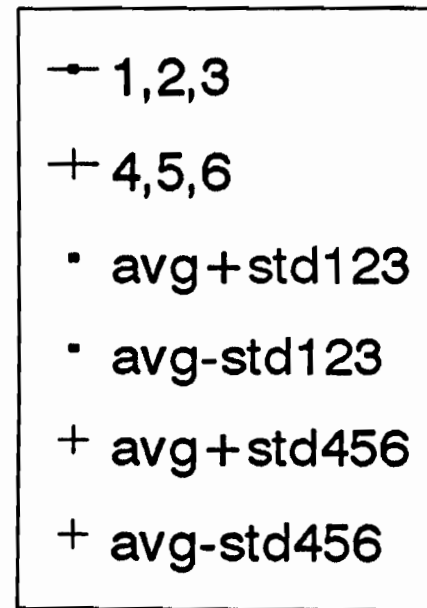


Figure 9. Control flow rate average compared to columns 4, 5, 6



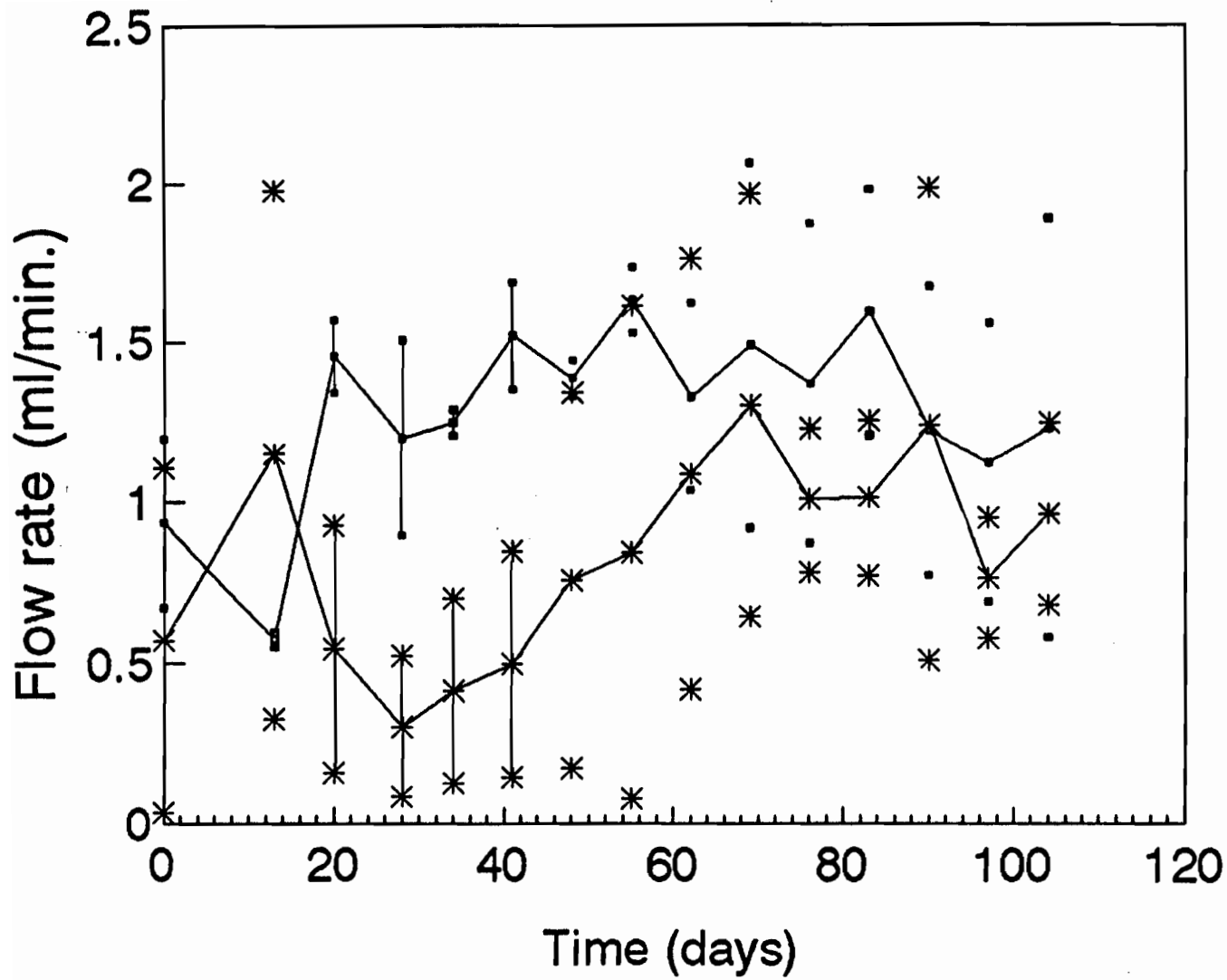
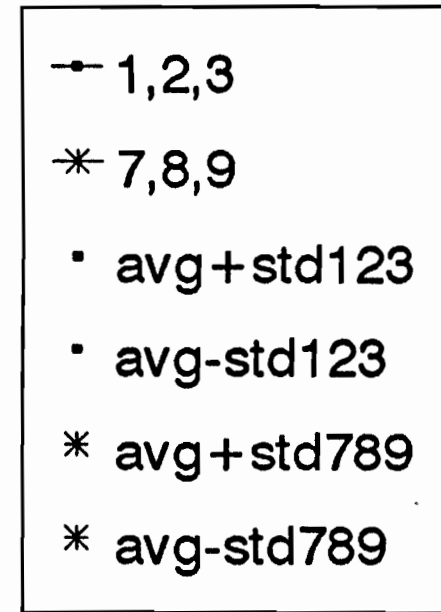


Figure 10. Control flow rate average compared to columns 7, 8, 9



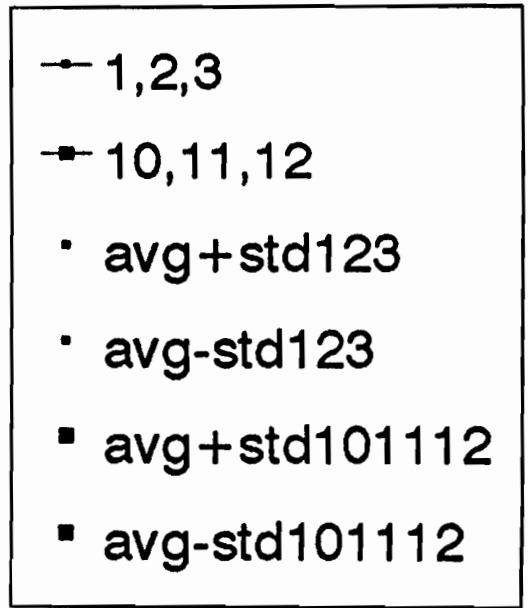
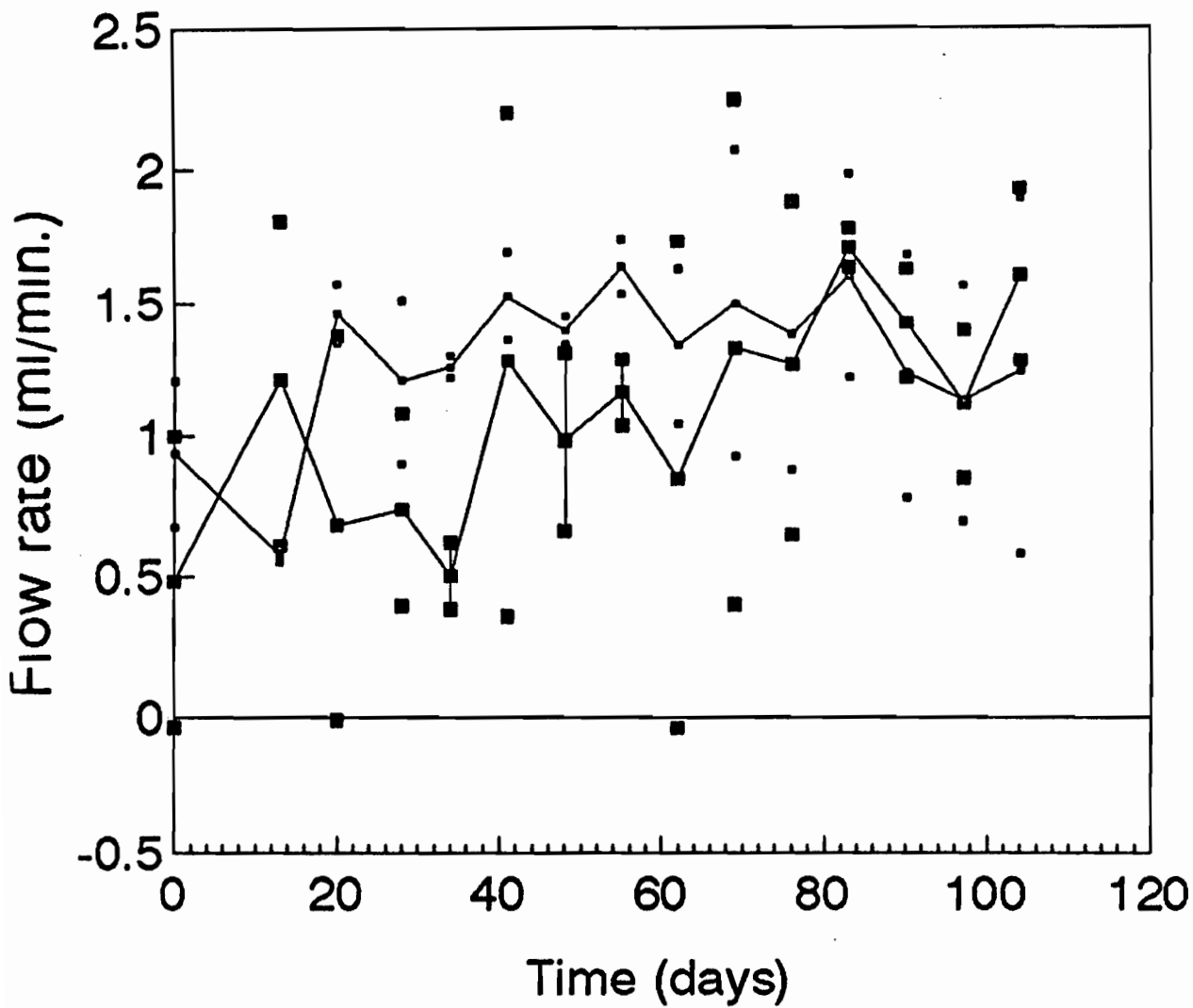


Figure 11. Control flow rate average compared to columns 10, 11, 12

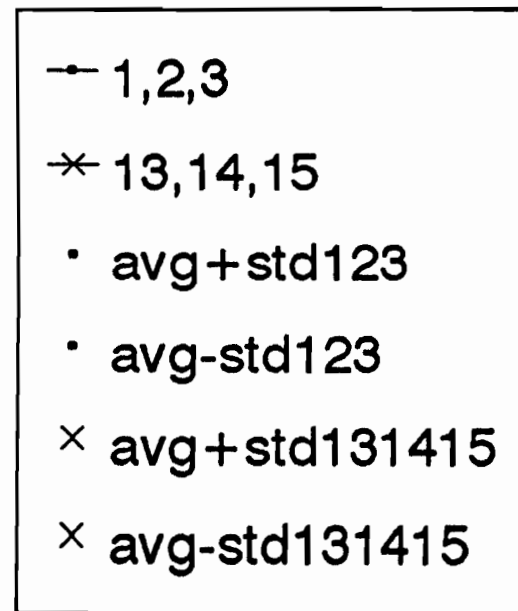
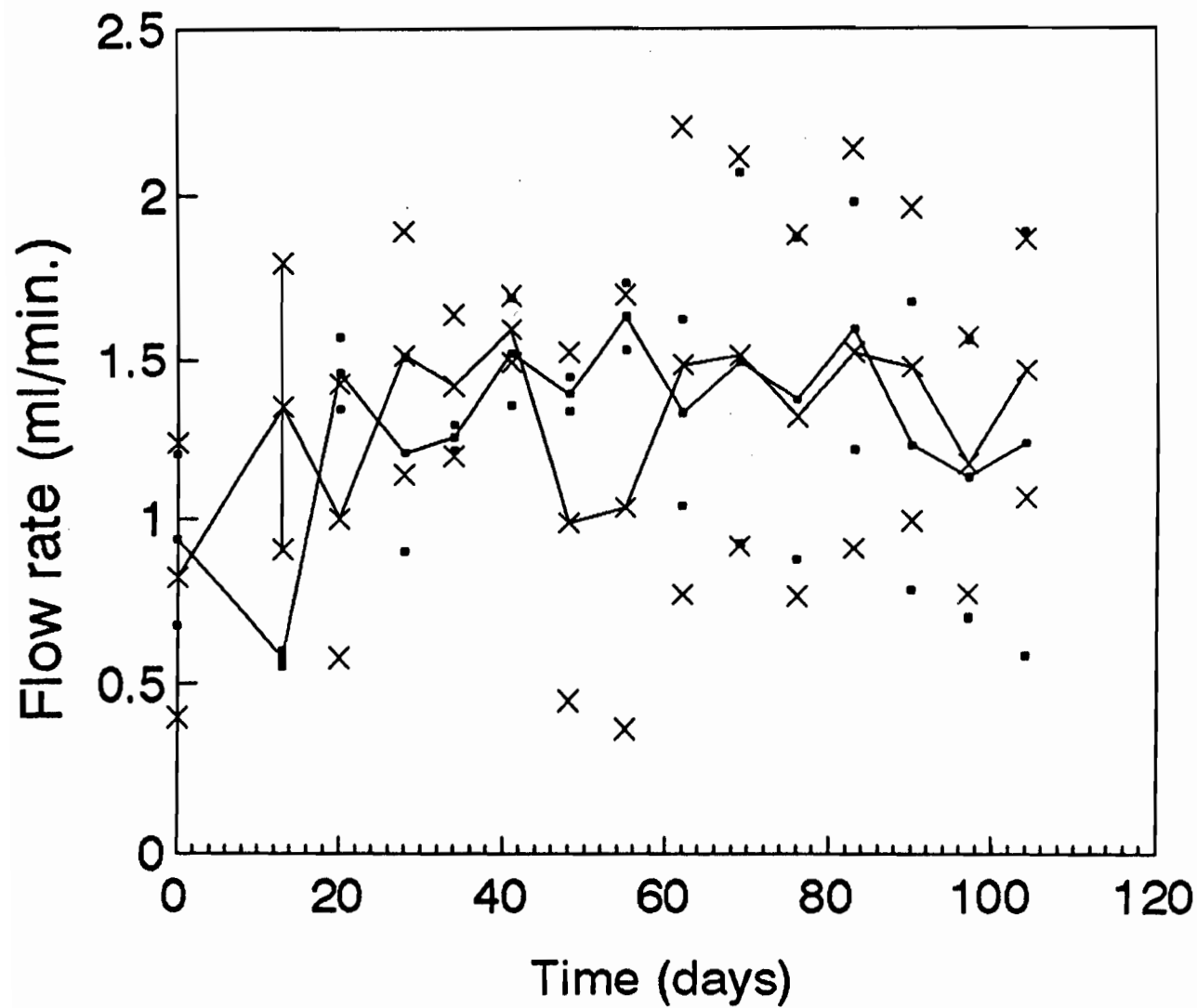


Figure 12. Control: flow rate average compared to columns 13, 14, 15

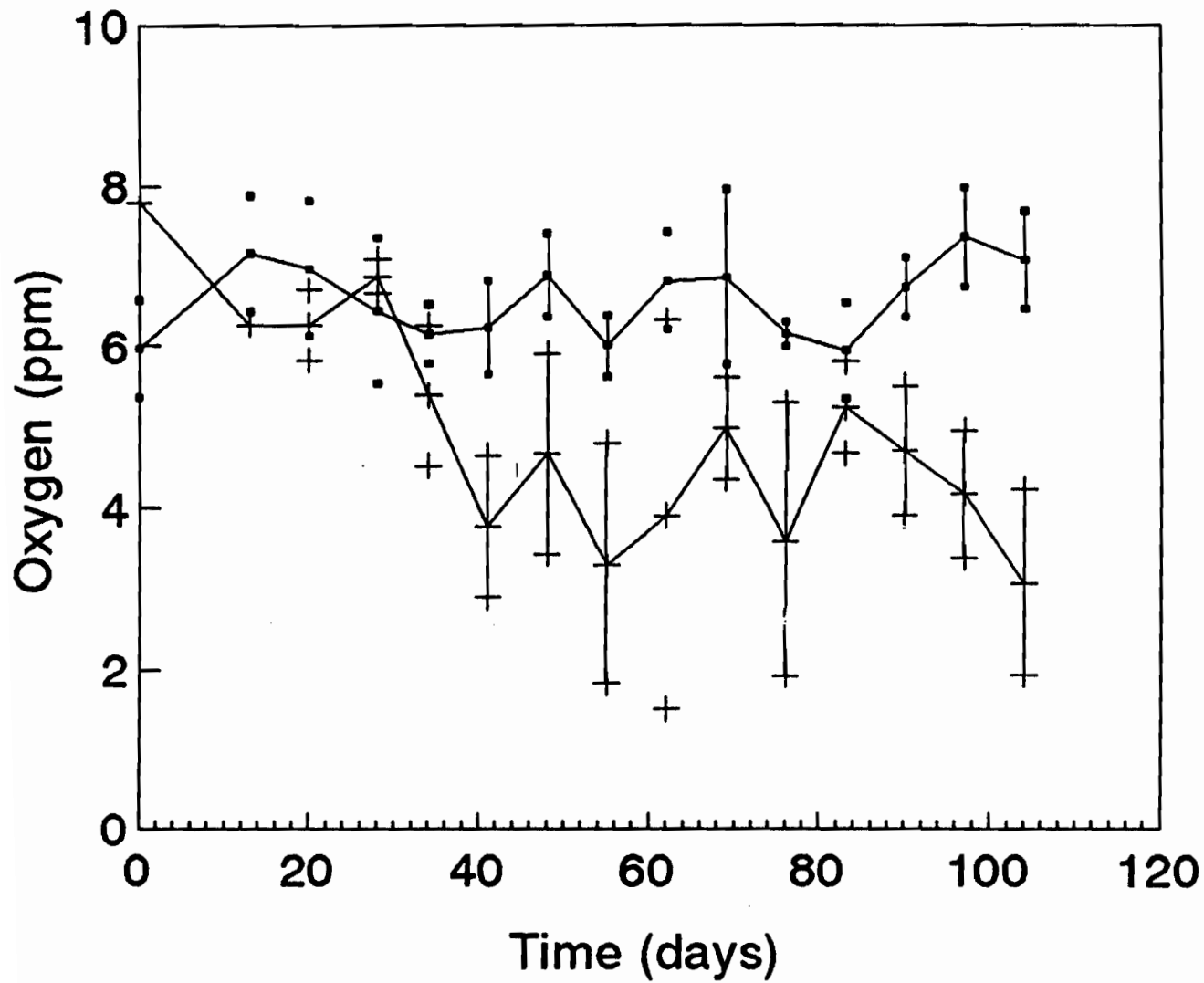
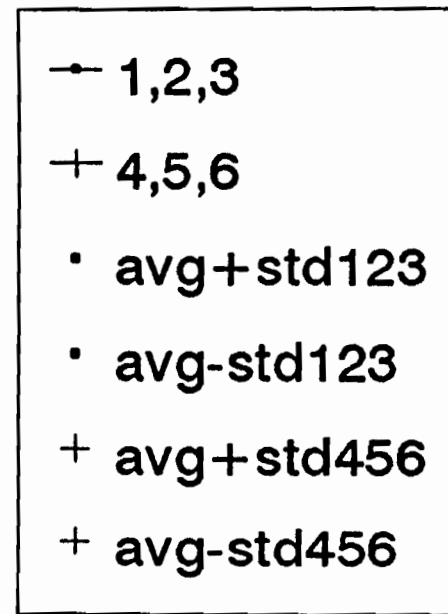


Figure 13. Control dissolved oxygen average compared to columns 4, 5, 6



25

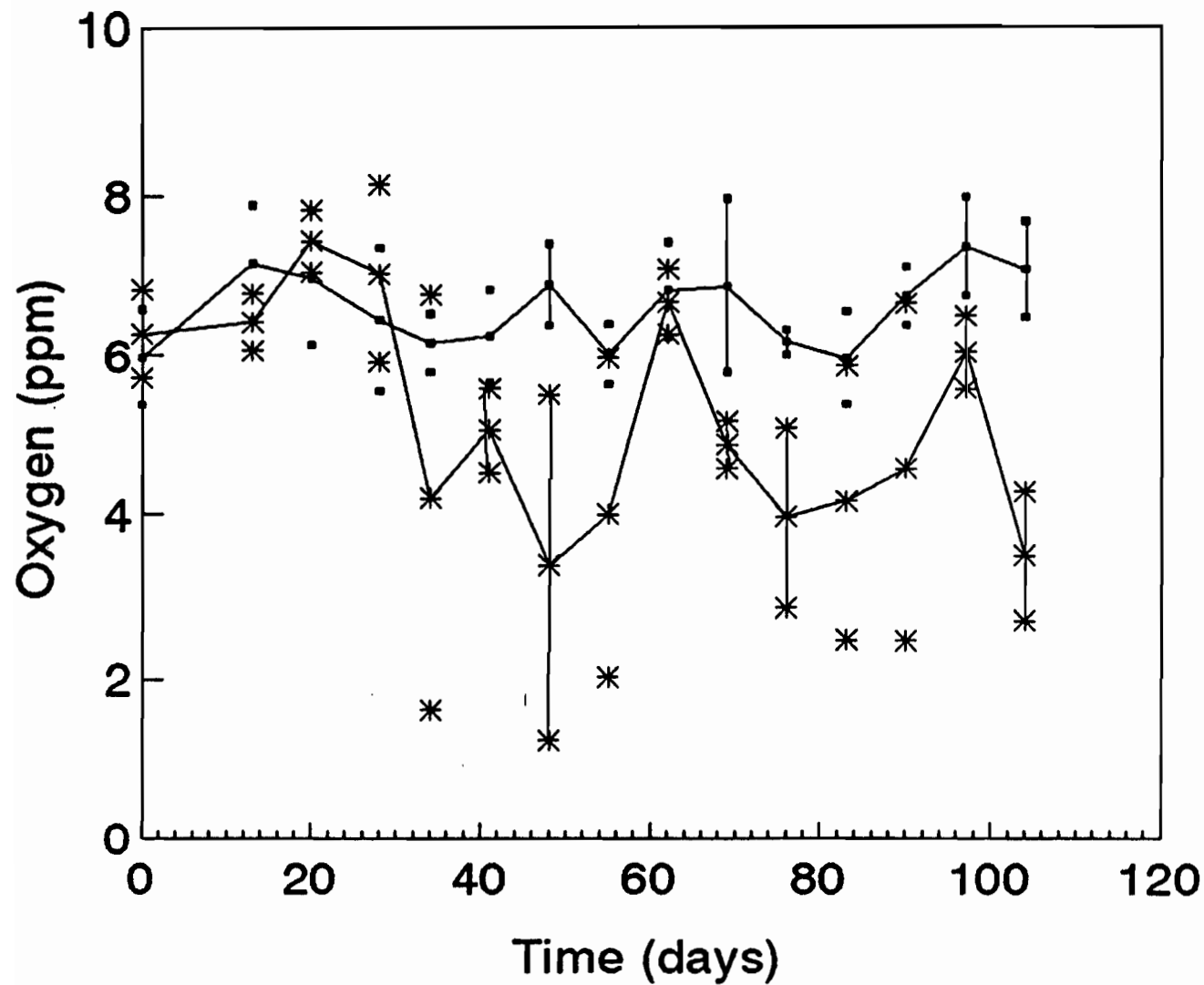
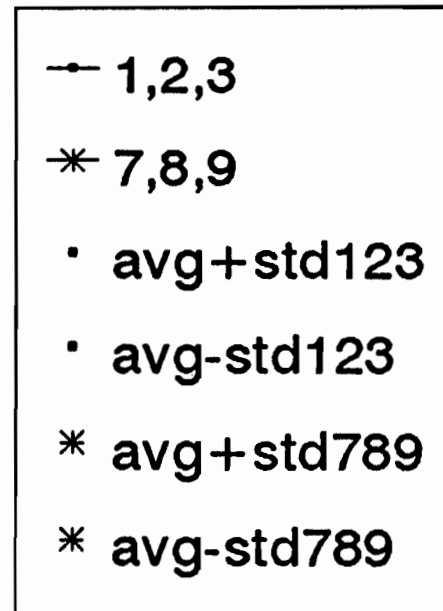


Figure 14. Control dissolved oxygen average compared to columns 7, 8, 9



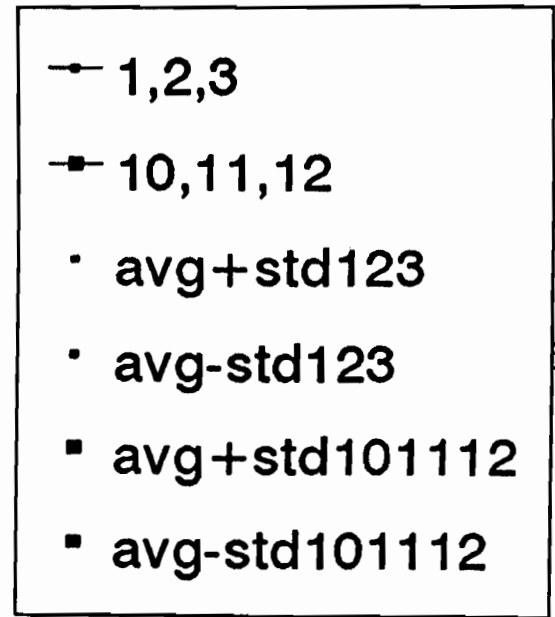
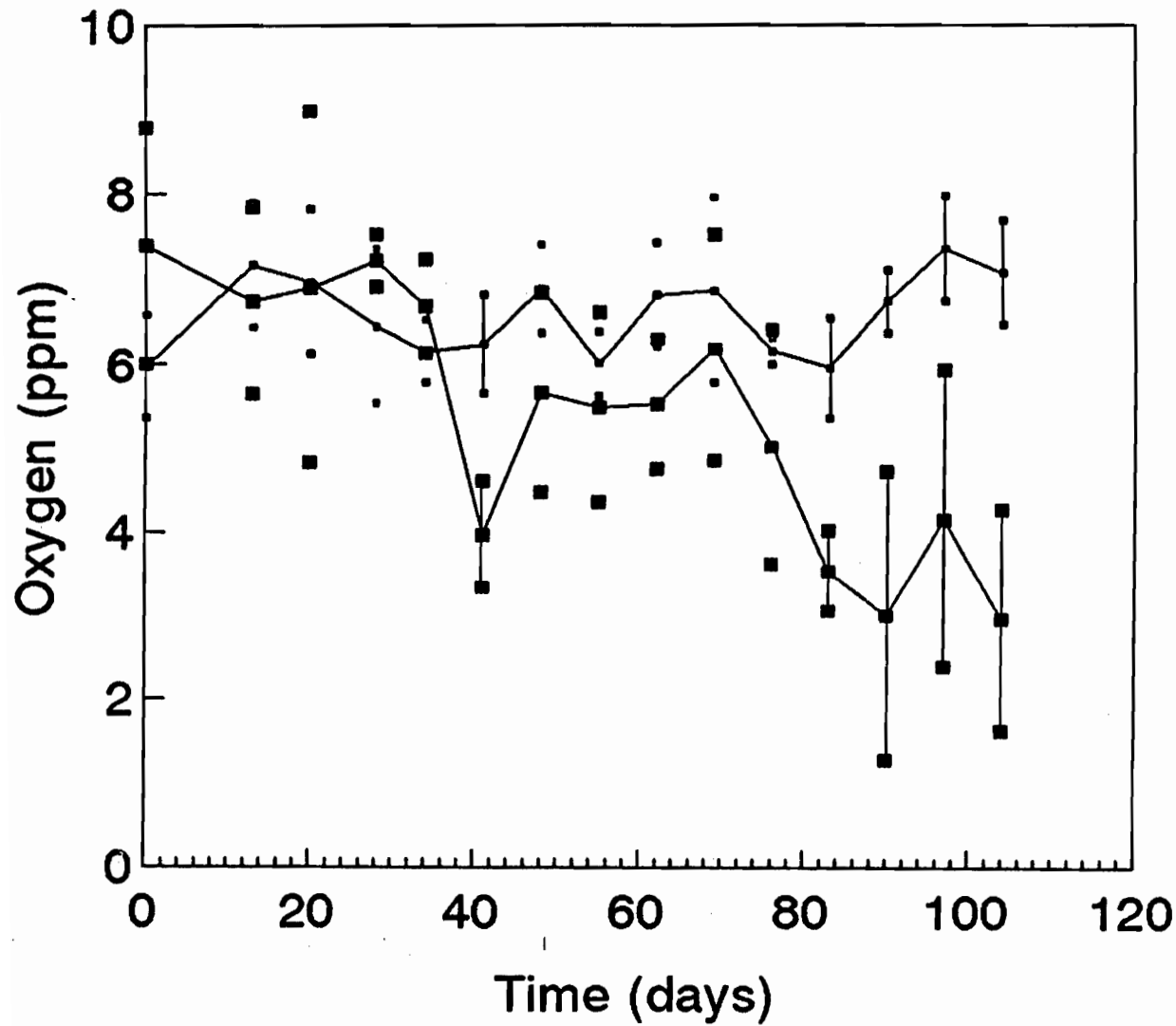


Figure 15. Control dissolved oxygen average compared to columns 10, 11, 12

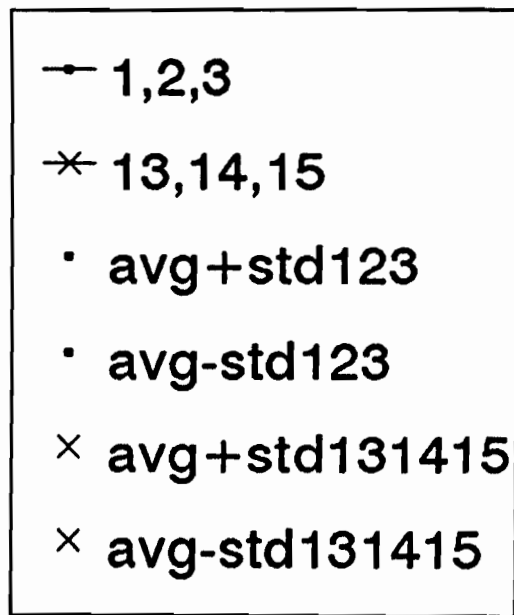
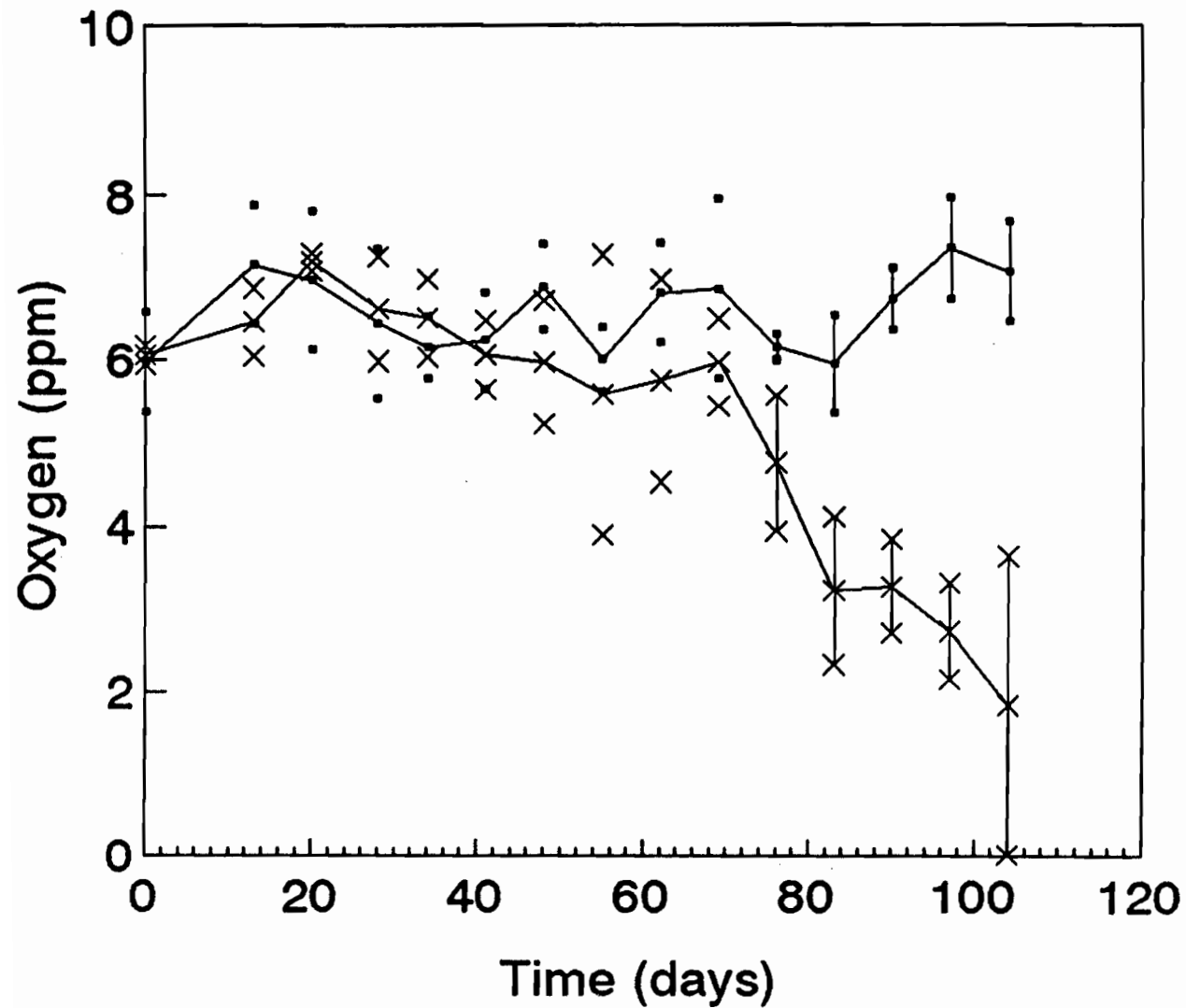


Figure 16. Control dissolved oxygen average compared to columns 13, 14, 15

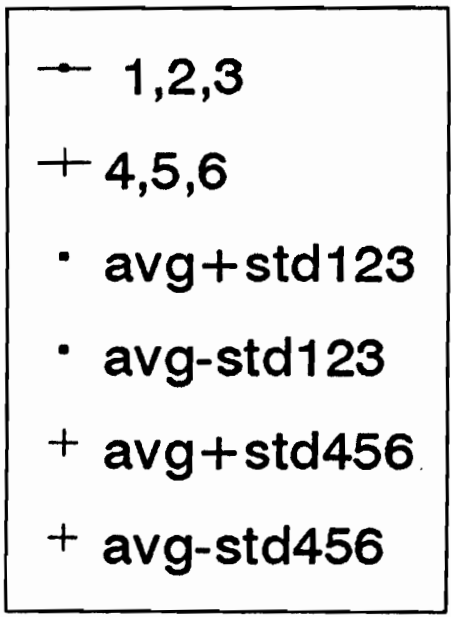
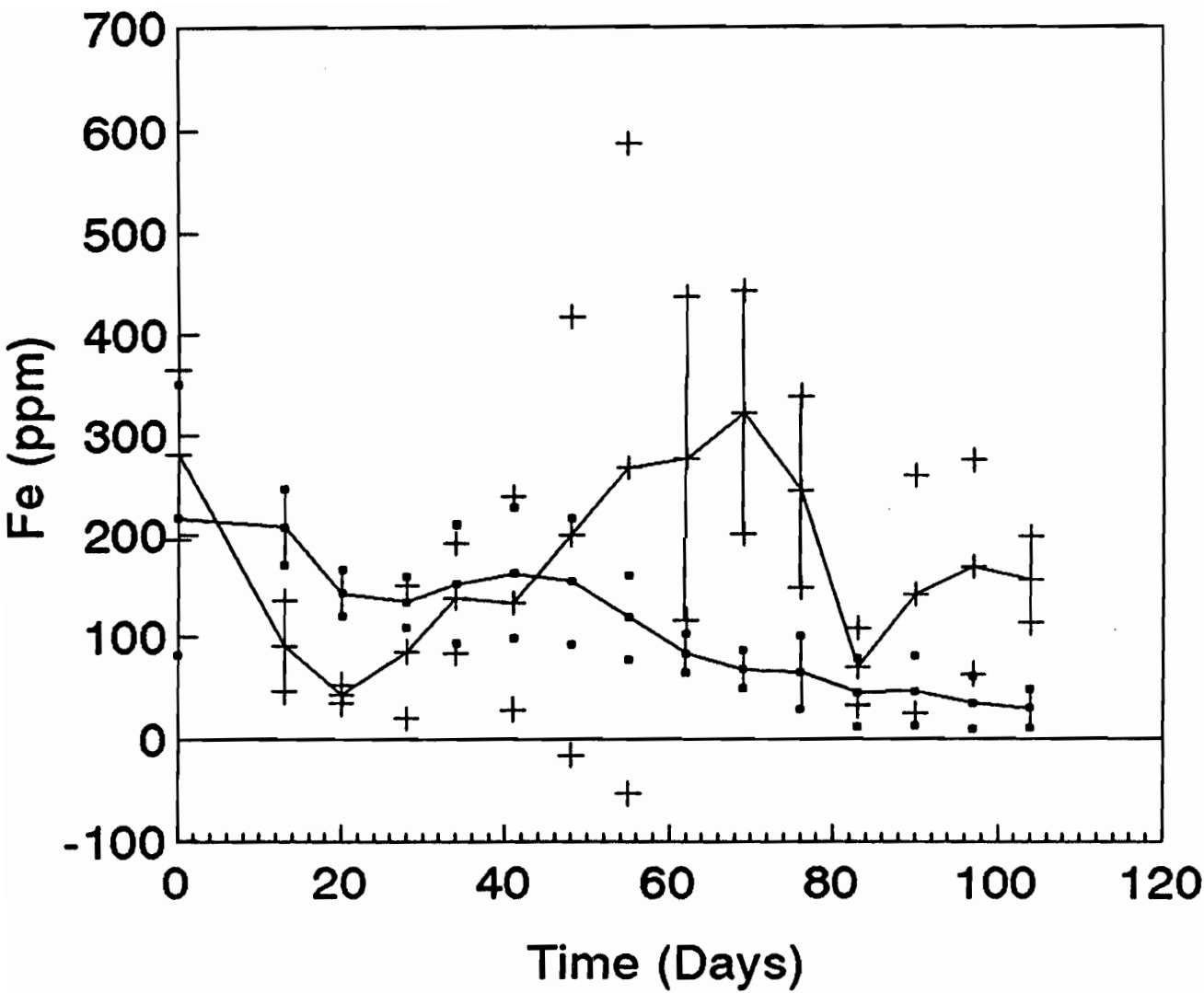


Figure 17. Control total iron average compared to columns 4, 5, 6

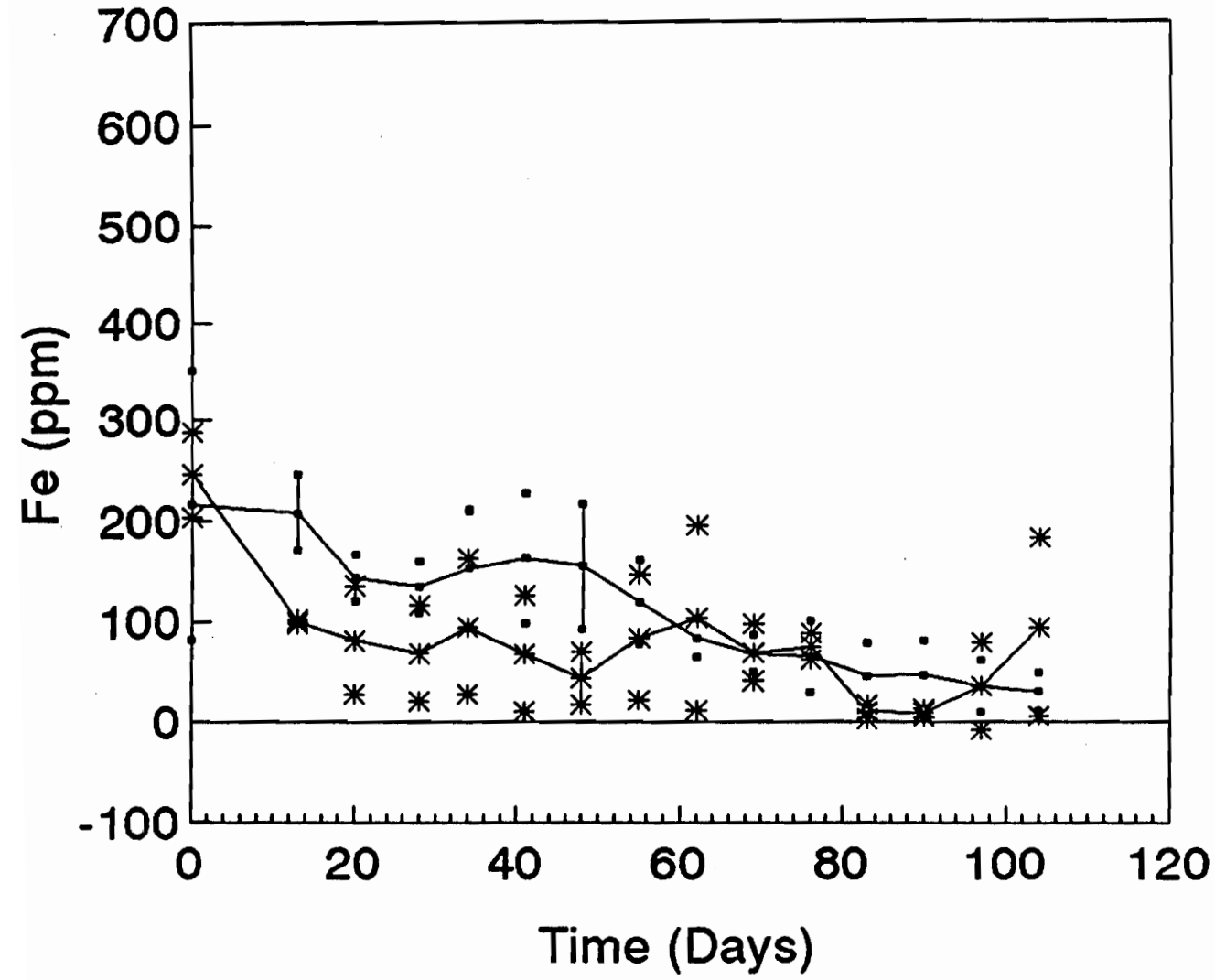
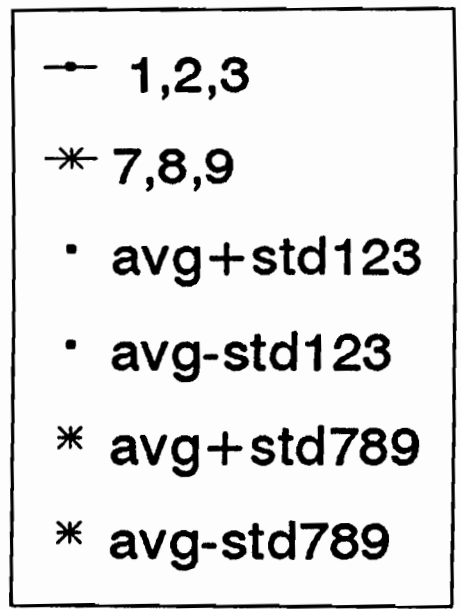


Figure 18. Control total iron average compared to columns 7, 8, 9



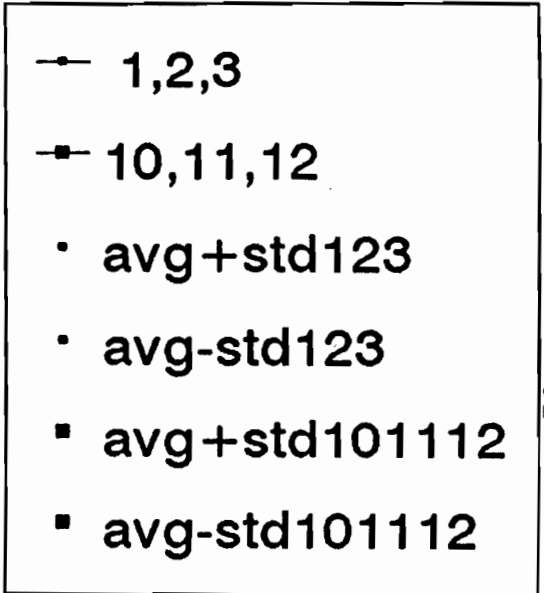
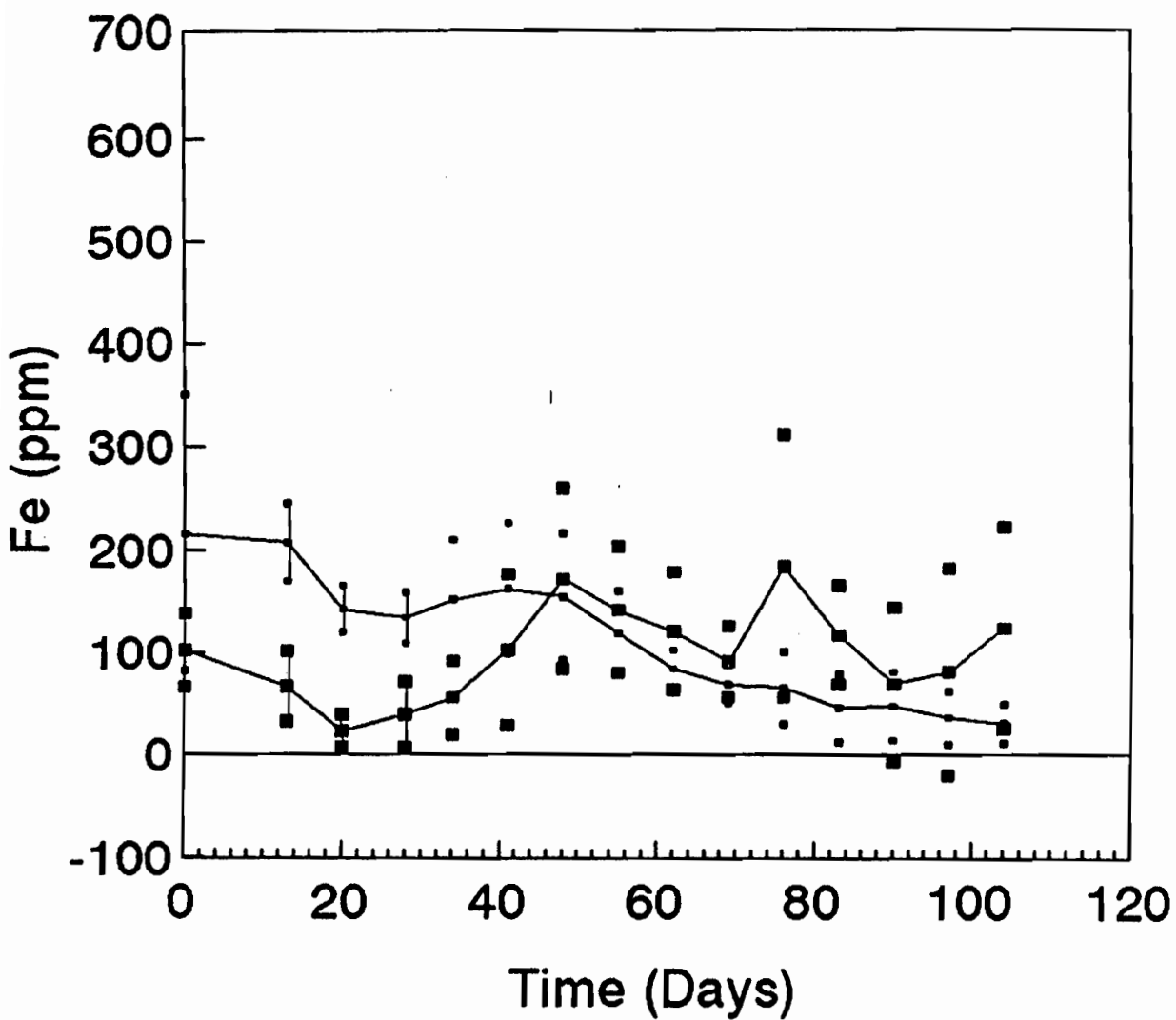


Figure 19. Control total iron average compared to columns 10, 11, 12

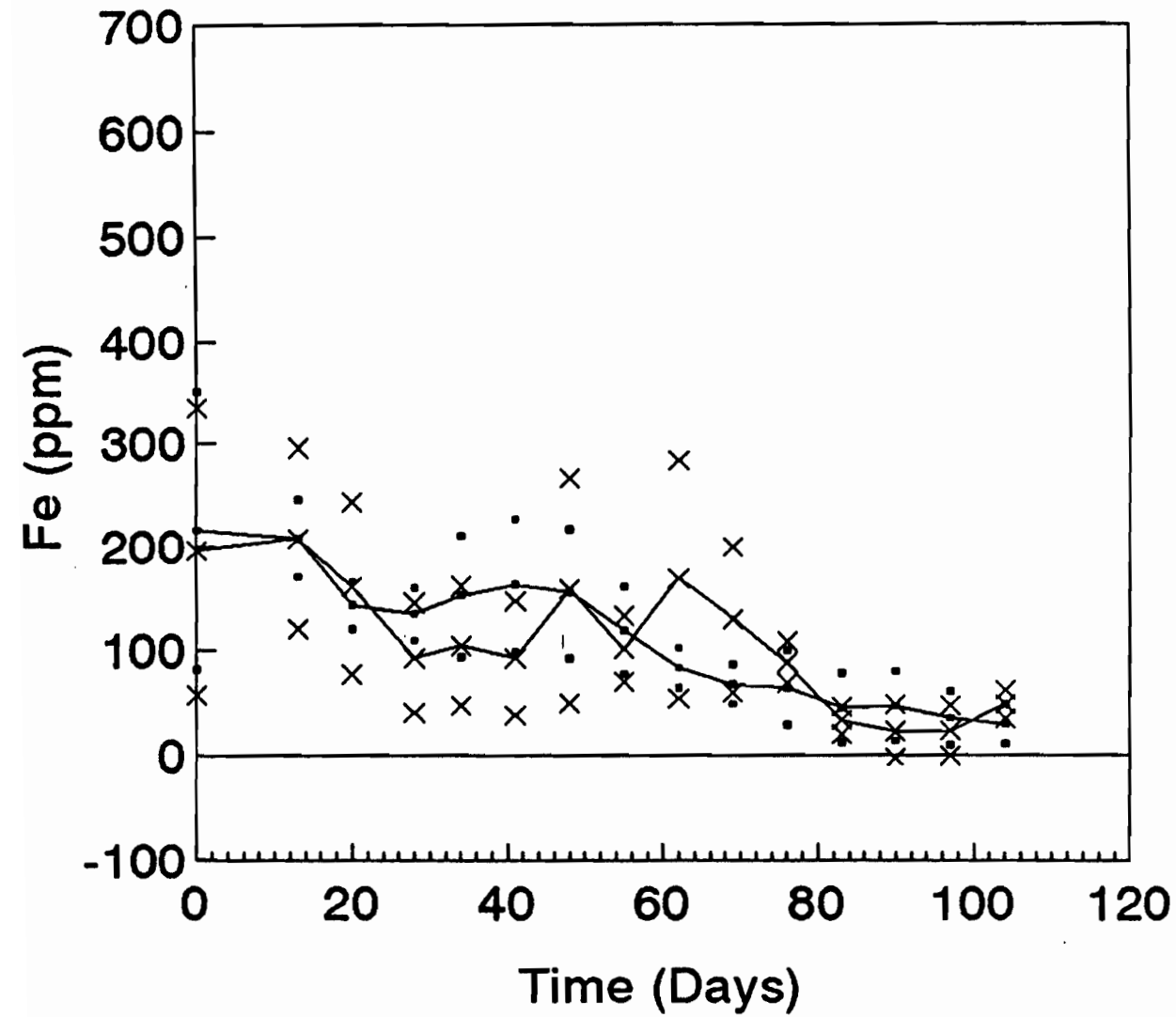


Figure 20. Control total iron average compared to columns 13, 14, 15

PROTECTED BUSINESS INFORMATION

APPENDIX - A

**Minutes of the
Technical Review Meeting
on
MEND PROJECT 2.44.1:
MICROBIAL PLUGGING**

September 30, 1992

555 Booth Street
Ottawa, Ontario

Attendance:	K. Wheeland, Noranda D. Gould, CANMET M. Skaff, CANMET	A. Oliver, CANMET L. Lortie, CANMET G. Tremblay, MEND
Absent:	G. Hope, Lac Minerals P. Dauphin, CANMET	T. Hynes, Inco R. McCready, C.A.R.E.

A meeting to discuss the results to date on the microbial plugging of tailings was held at CANMET on Wednesday, September 30, 1992. A draft report was tabled and Ms. L. Lortie, Project Leader, presented the results. The conclusions of the study were that:

- (1) It was not demonstrated that a microbial cap could significantly reduce the permeability of the tailings over long periods of time;
- (2) A decrease of the oxygen penetration was observed for the columns that received nutrients;
- (3) An increase in pH was observed for all of the permeates from the columns that received nutrients; and
- (4) Addition of ultramicrobacteria's (UMB's) to the tailings is not necessary because native strains are present in the tailings.

It was noted that although a reduction in permeability was not observed in the experiments, it is possible that in fine textural tailings a significant reduction in water permeability would be expected.

Ms. Lortie then presented the work needed to complete this study. It includes:

- (1) Microbial counts;
- (2) Identification of strains to verify the presence of UMB's (*Klebsiella oxytoca*);
- (3) Measure uronic acid concentrations; and

(4) Determining the economics of the process.

The samples needed to complete the first three tasks are scheduled to be taken at the end of the study because the coring procedure used will prevent any further laboratory experimental work. G. Tremblay, who visited R. McCready of C.A.R.E. in Calgary in early September reported that C.A.R.E. had agreed to produce a one-page summary on the economics of the process to be included in the final report.

The recommendations made by CANMET at this stage of the project followed and included:

- (1) No further studies be undertaken by CANMET (other than complete the present study and issue a final report);
- (2) Continue project and complete additional test work:
 - a) Using humidity cells;
 - b) Trying different acid generating tailings;
 - c) Running the experiments for a longer period of time;
 - d) Adding nutrients more often; and
 - e) Analyzing for SO_4 .

It was agreed by those present that CANMET will have completed their portion of the experimental work when the final report is submitted and accepted by MEND (i.e. recommendation (1)). Although only one of the two objectives was accomplished, it appears that the technique of microbial plugging has promise and further evaluation should be continued by other organizations.

Gilles Tremblay
MEND - Prevention and Control

APPENDIX - B

ECONOMICS OF BACTERIAL CAPPING OF TAILINGS

The current study has indicated that the tailings need not be inoculated with vegetative or ultramicrobial cells in order to produce an oxygen scavenging layer in the upper surface of the tailings. There are sufficient indigenous heterotrophs present in the surface layers of the tailings. When stimulated with rich carbohydrate medium buffered to pH 7.0, the indigenous organisms multiply and produce sufficient exopolysaccharides to reduce the percolation of precipitation and to remove the dissolved oxygen from the interstitial solution within the tailings resulting in a decrease in the Eh, and a concurrent increase in the pH of the effluents which have permeated through the tailings.

The use of a food industry waste product as the microbial nutrient, which will have to be applied at monthly intervals has resulted in a substantial reduction in the cost of the technology. Considering the cost of the waste product, the shipping costs and the application costs over a three to four month period, the cost is estimated to be approximately \$25,000 per hectare. Once the cap has been established the surface of the tailings would be re-vegetated. By judiciously selecting a revegetation cover which exudes root exudates, the microbial oxygen scavenging layer should become self-sustaining. The revegetation aspect requires further research to prove that the concept is correct and economically feasible.

In some tailings, where an indigenous population of aciduric heterotrophs has not become established, microbial inoculation may be required. In such cases, this would result in a substantial increase in the cost of the technology, increasing the estimated cost to approximately \$50,000 per hectare.

Dr. Ron McCready
C.A.R.E.

APPENDIX - C

APPENDIX - D

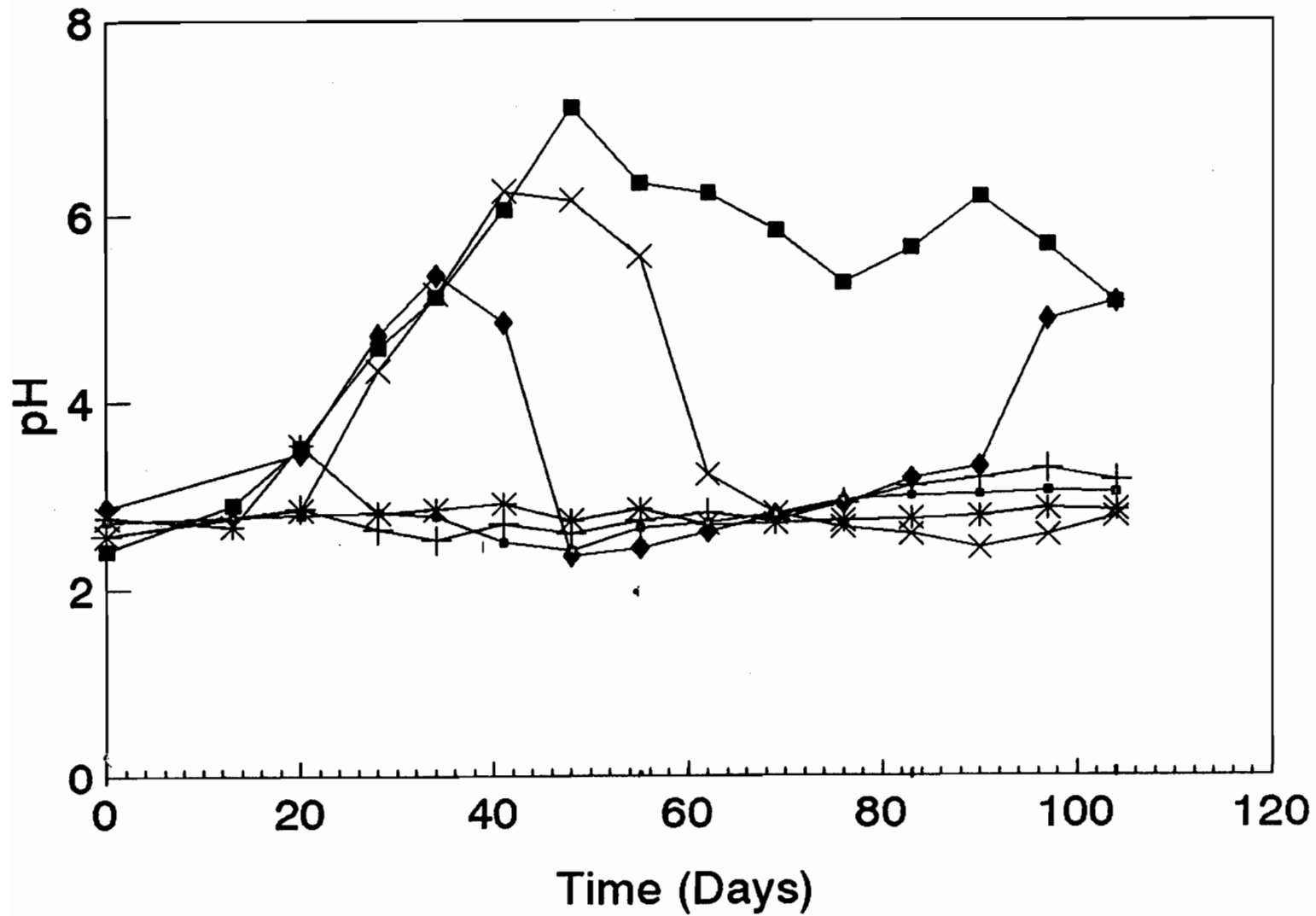


Figure 21. Controls pH compared to columns 4, 5, 6

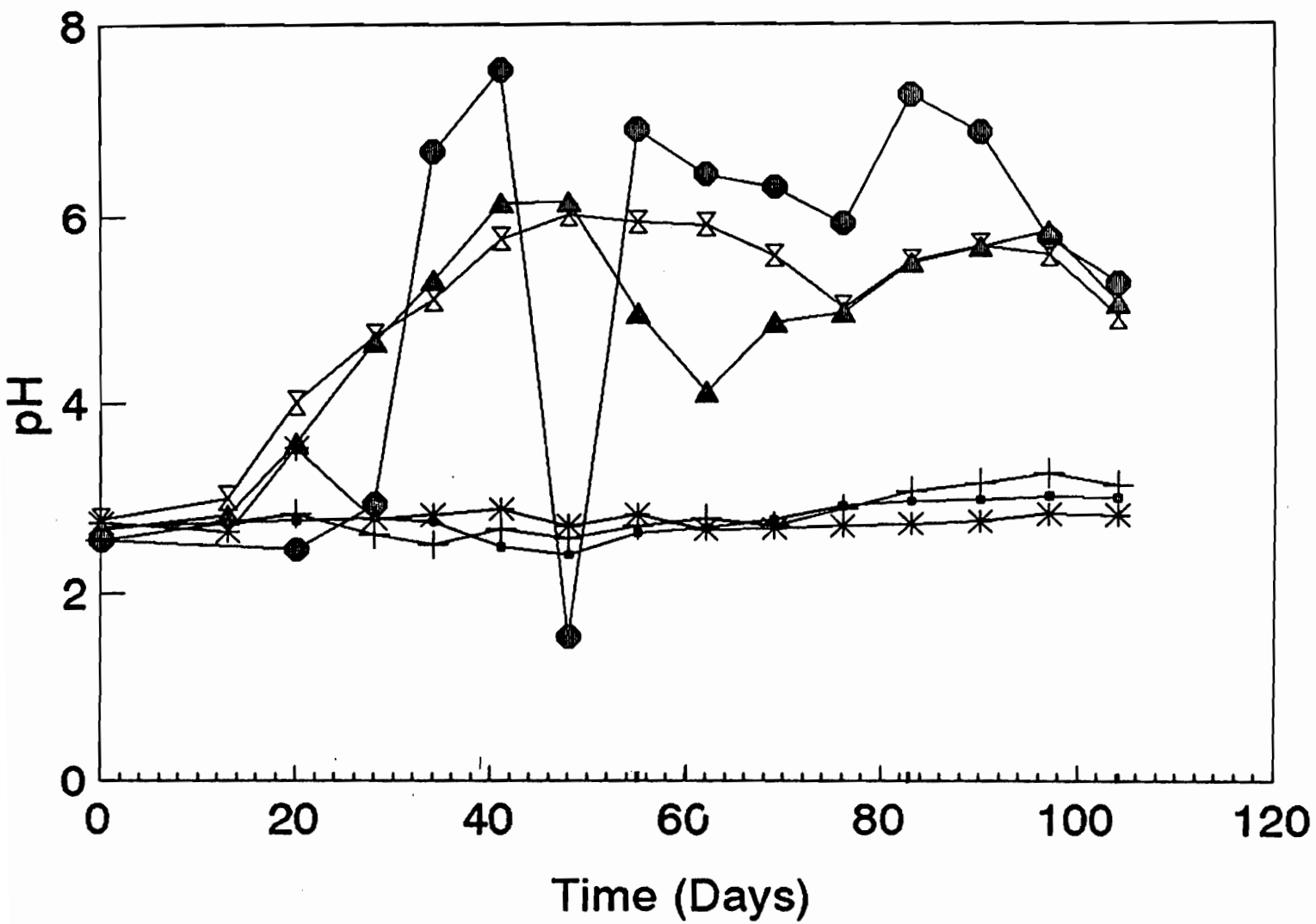


Figure 22. Controls pH compared to columns 7, 8, 9

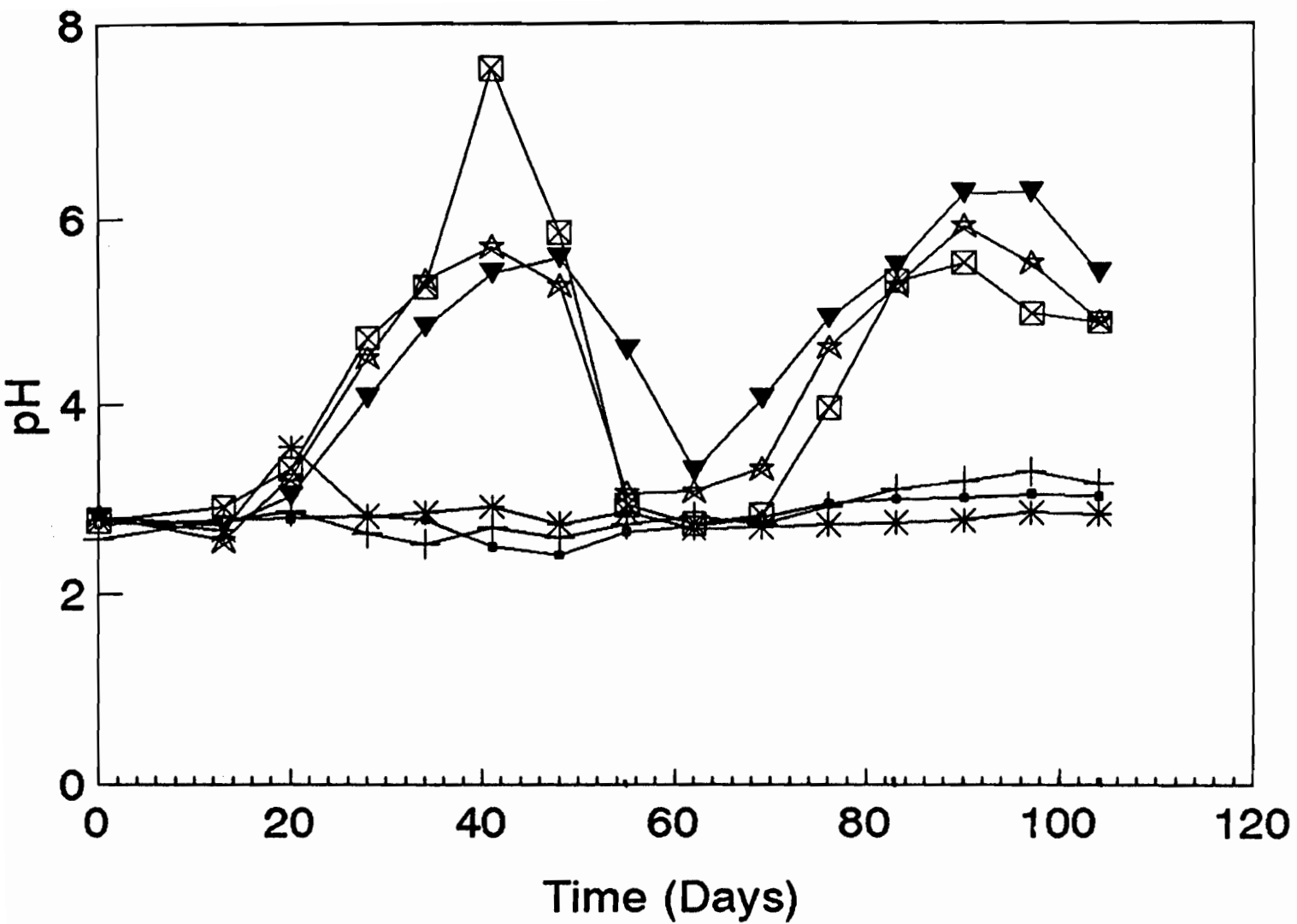


Figure 23. Controls pH compared to columns 10, 11, 12

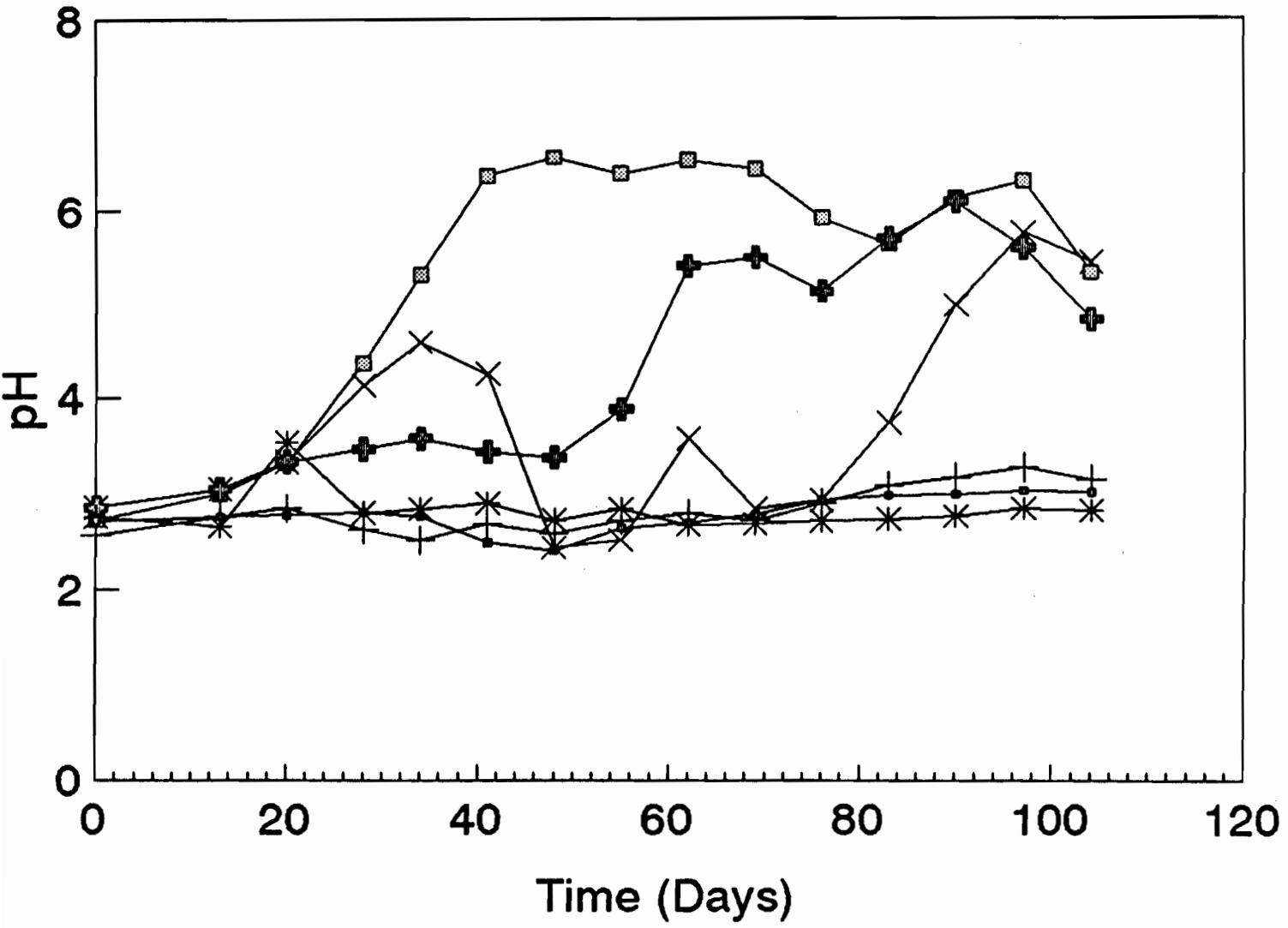


Figure 24. Controls pH compared to columns 13, 14, 15

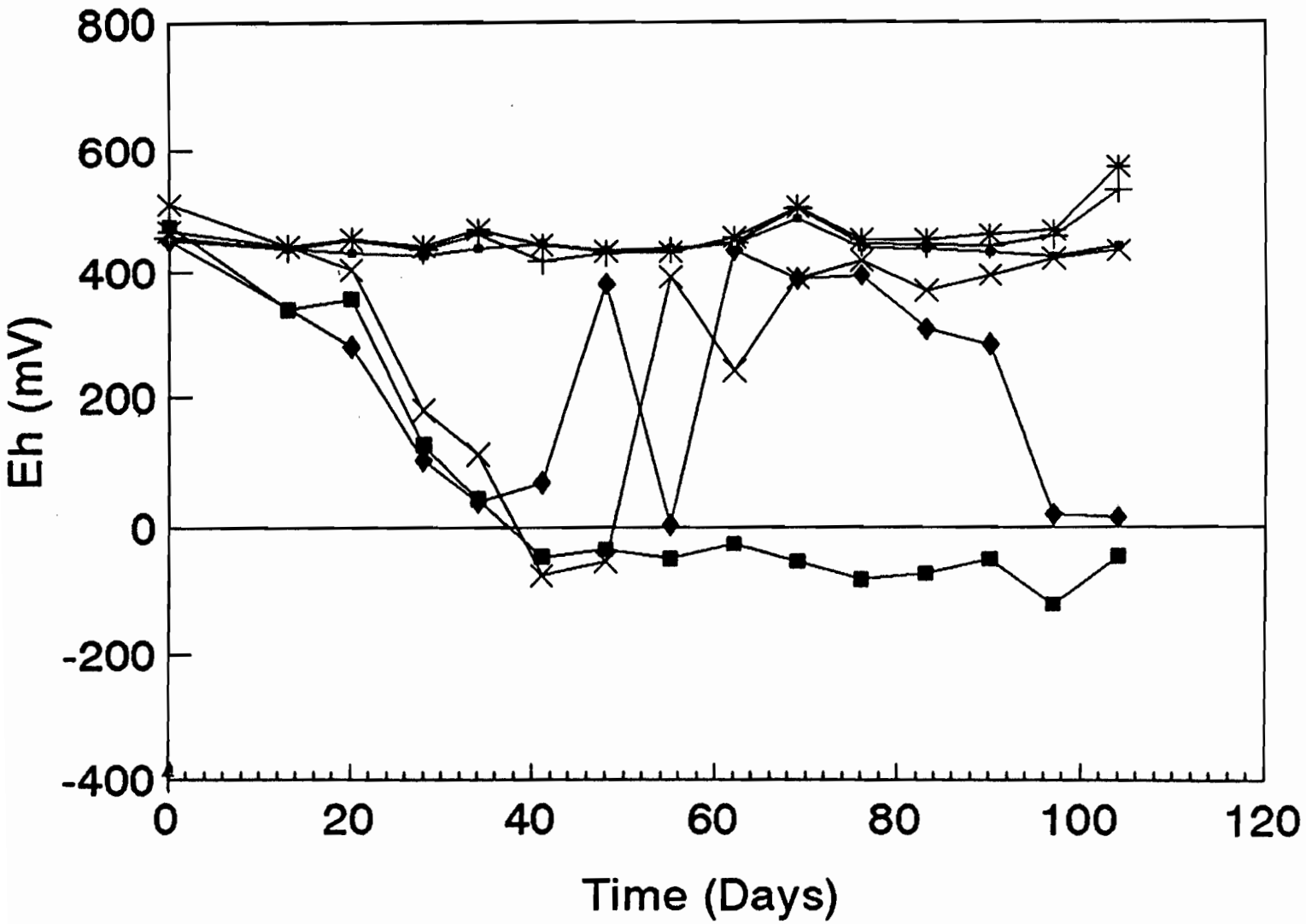


Figure 25. Controls Eh compared to columns 4, 5, 6

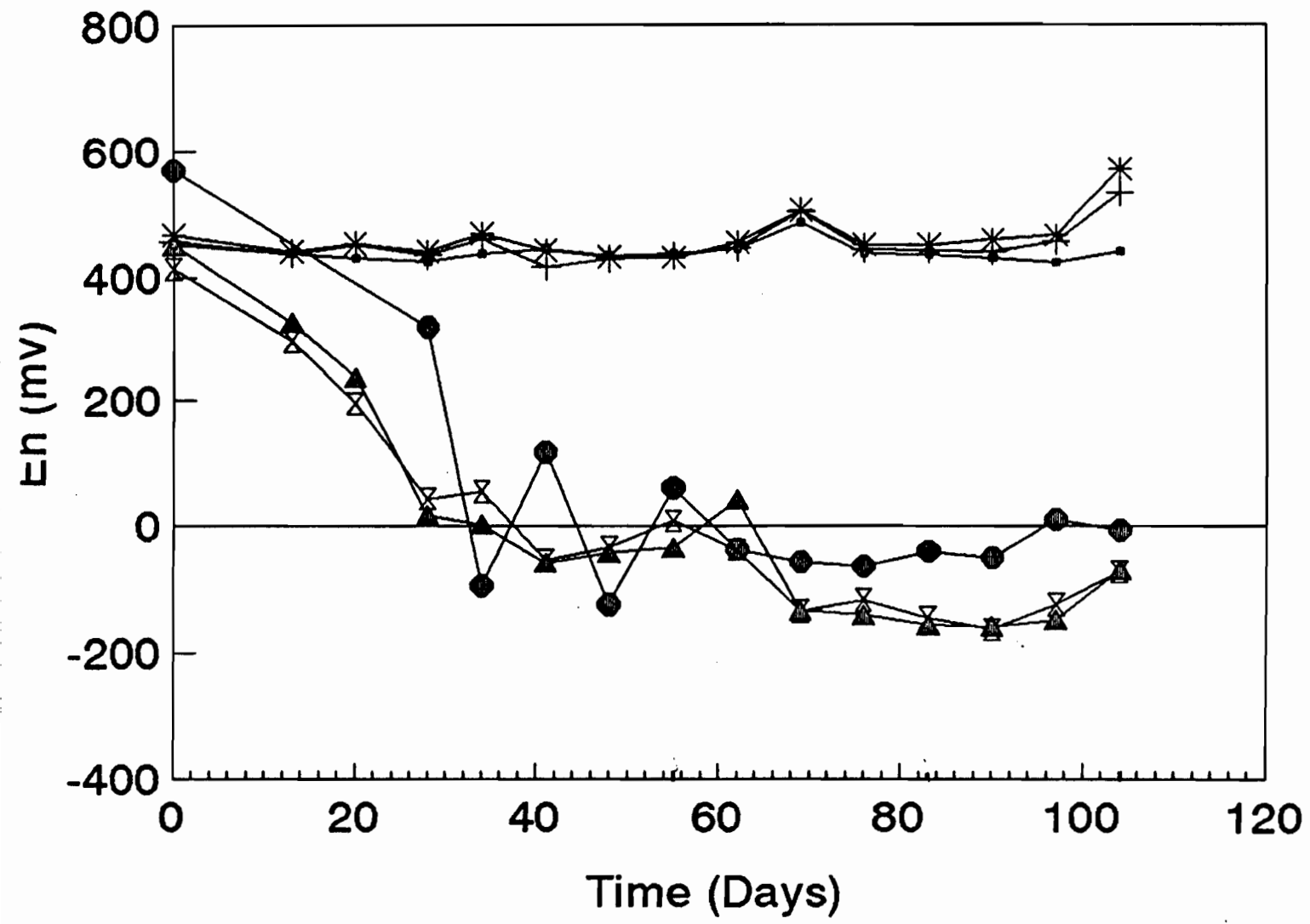


Figure 26. Controls Eh compared to columns 7, 8, 9

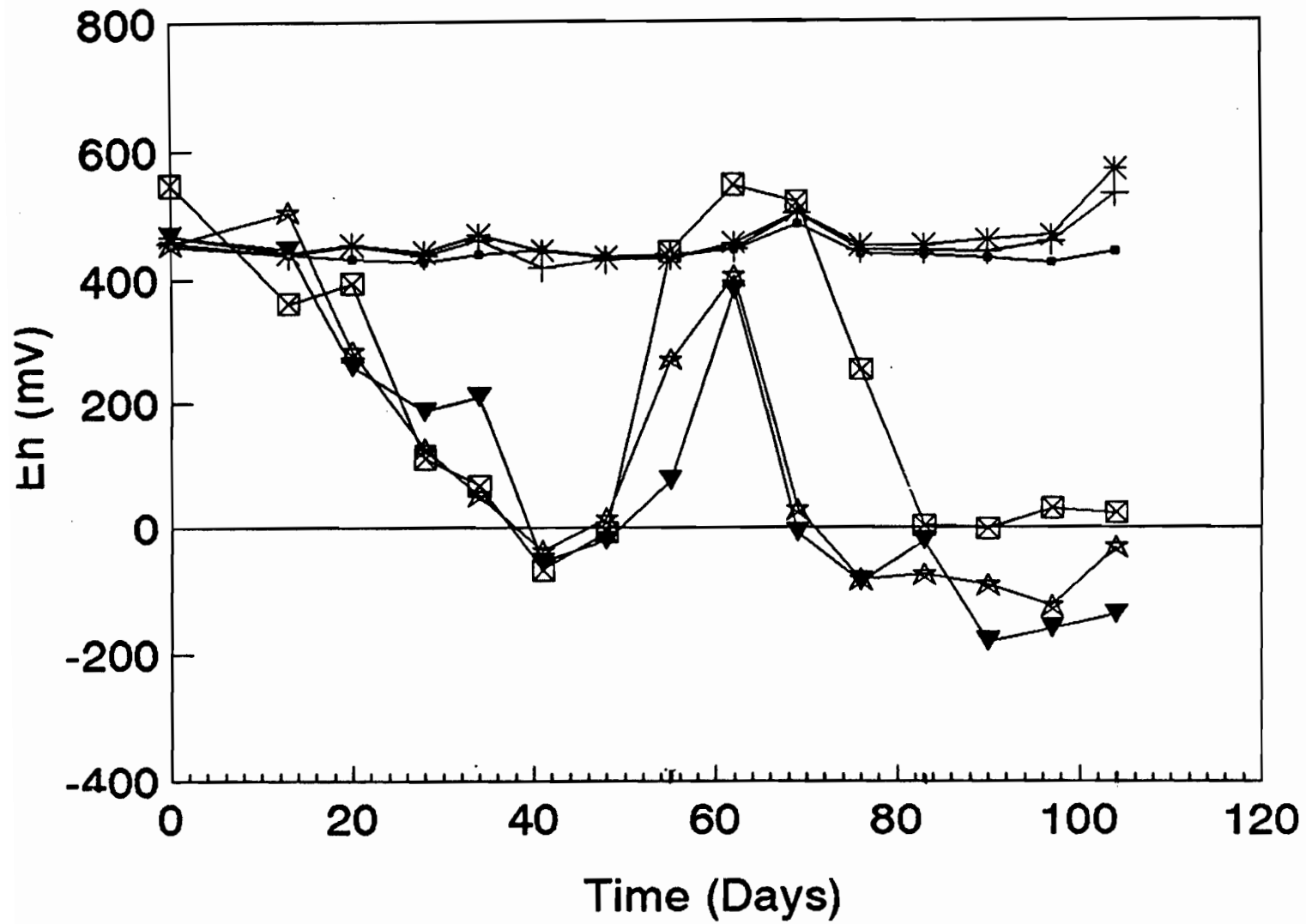


Figure 27. Controls Eh compared to columns 10, 11, 12

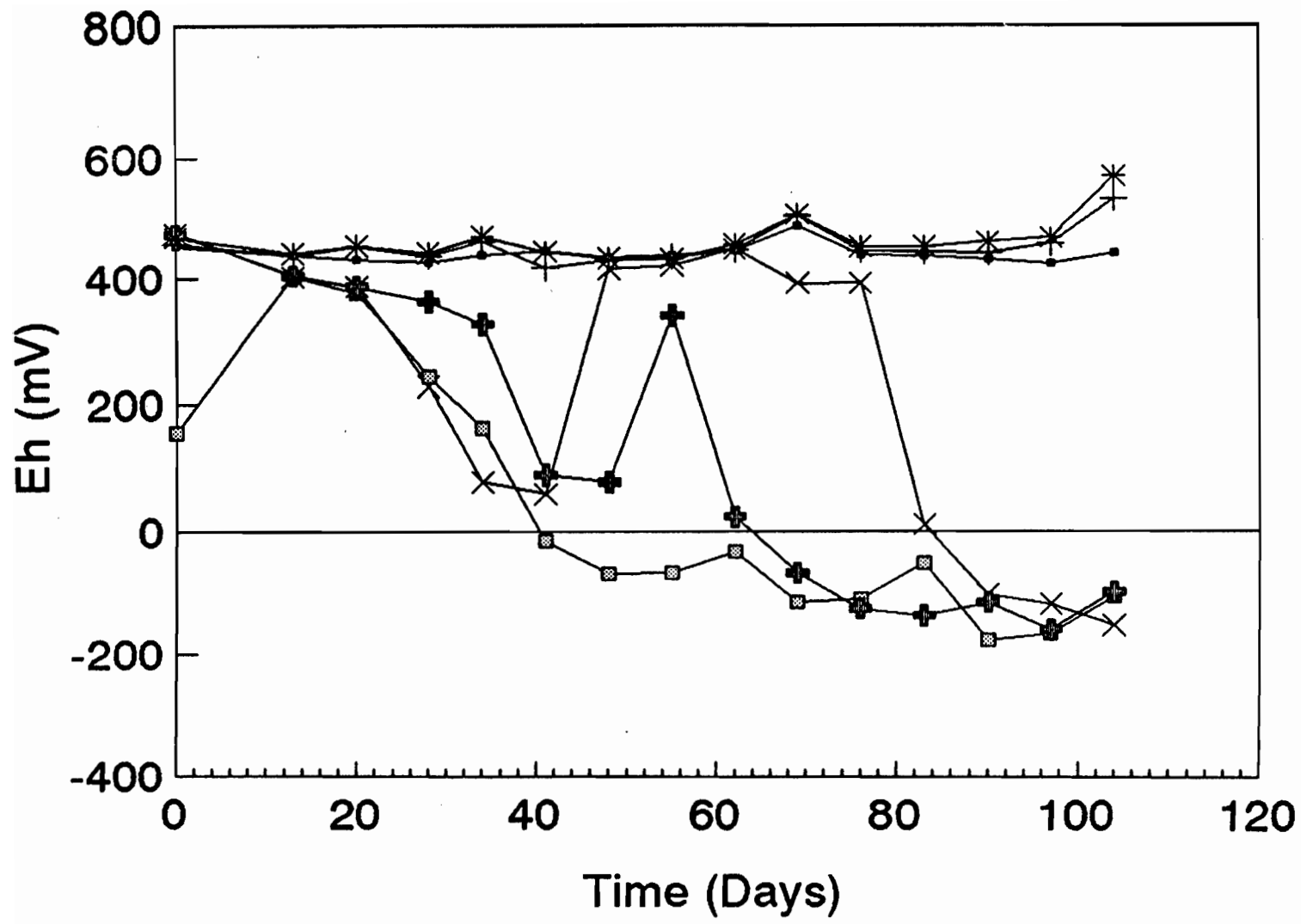


Figure 28. Controls Eh compared to columns 13, 14, 15

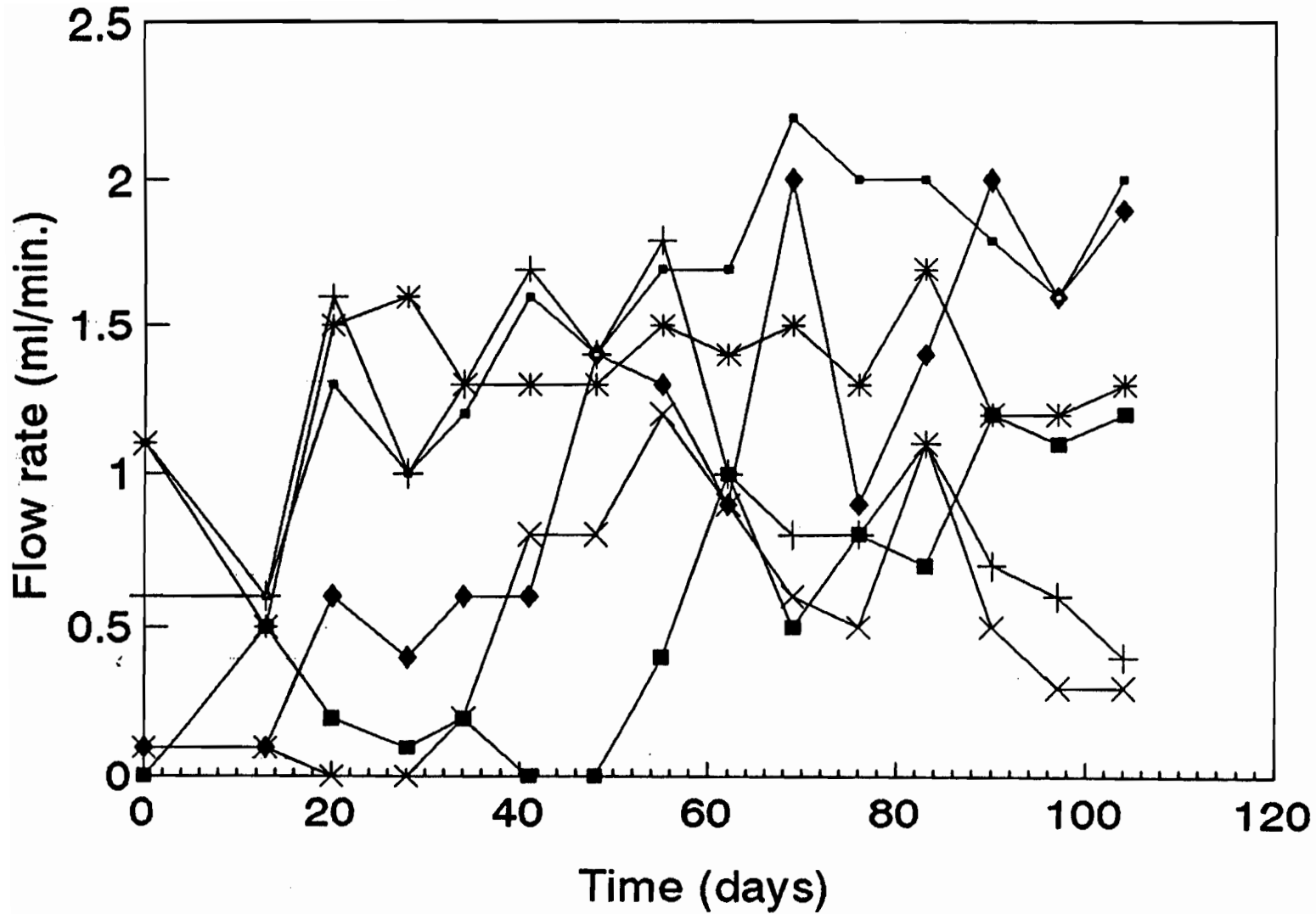


Figure 29. Controls flow rate compared to columns 4, 5, 6

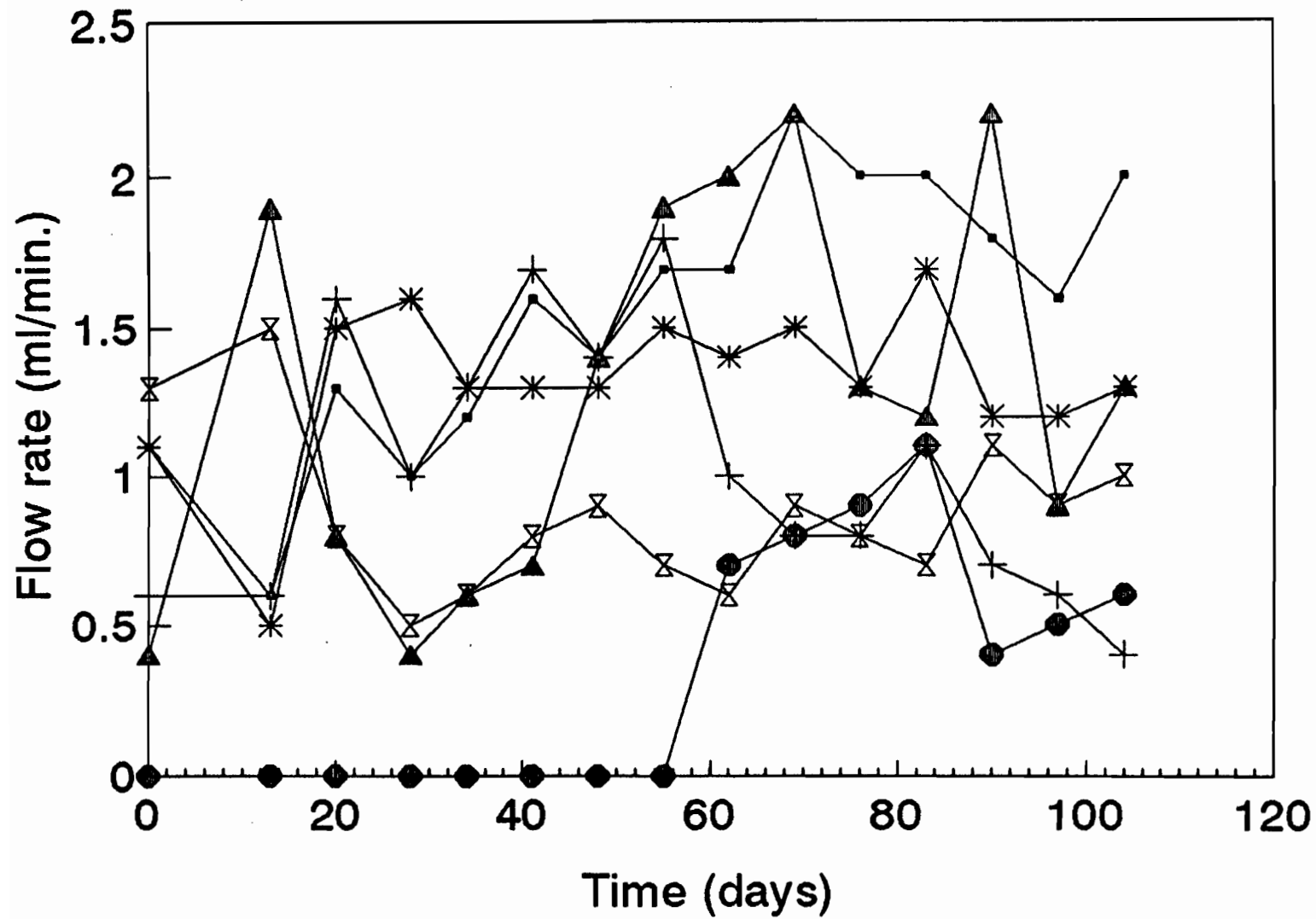


Figure 30. Controls flow rate compared to columns 7, 8, 9

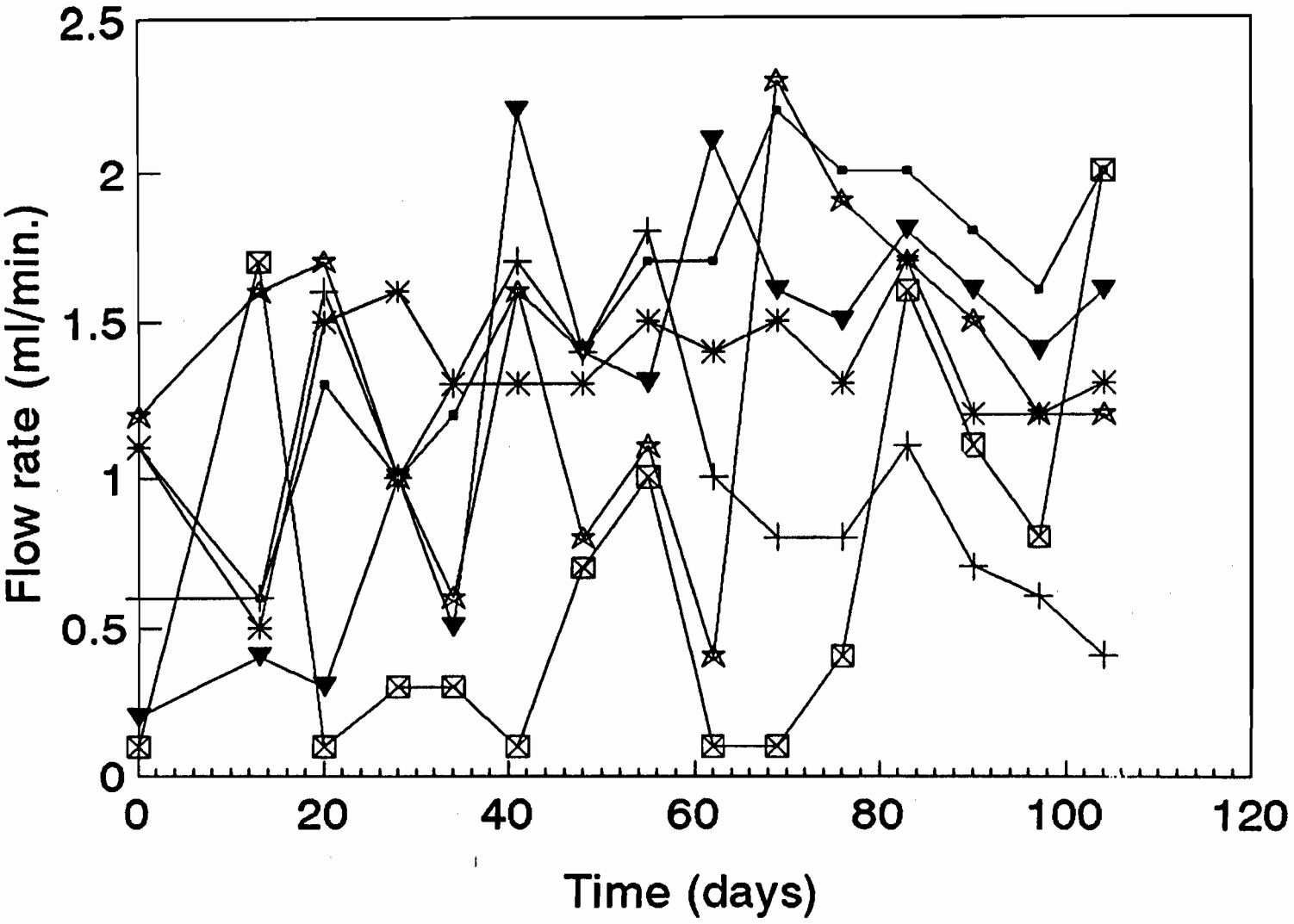


Figure 31. Controls flow rate compared to columns 10, 11, 12

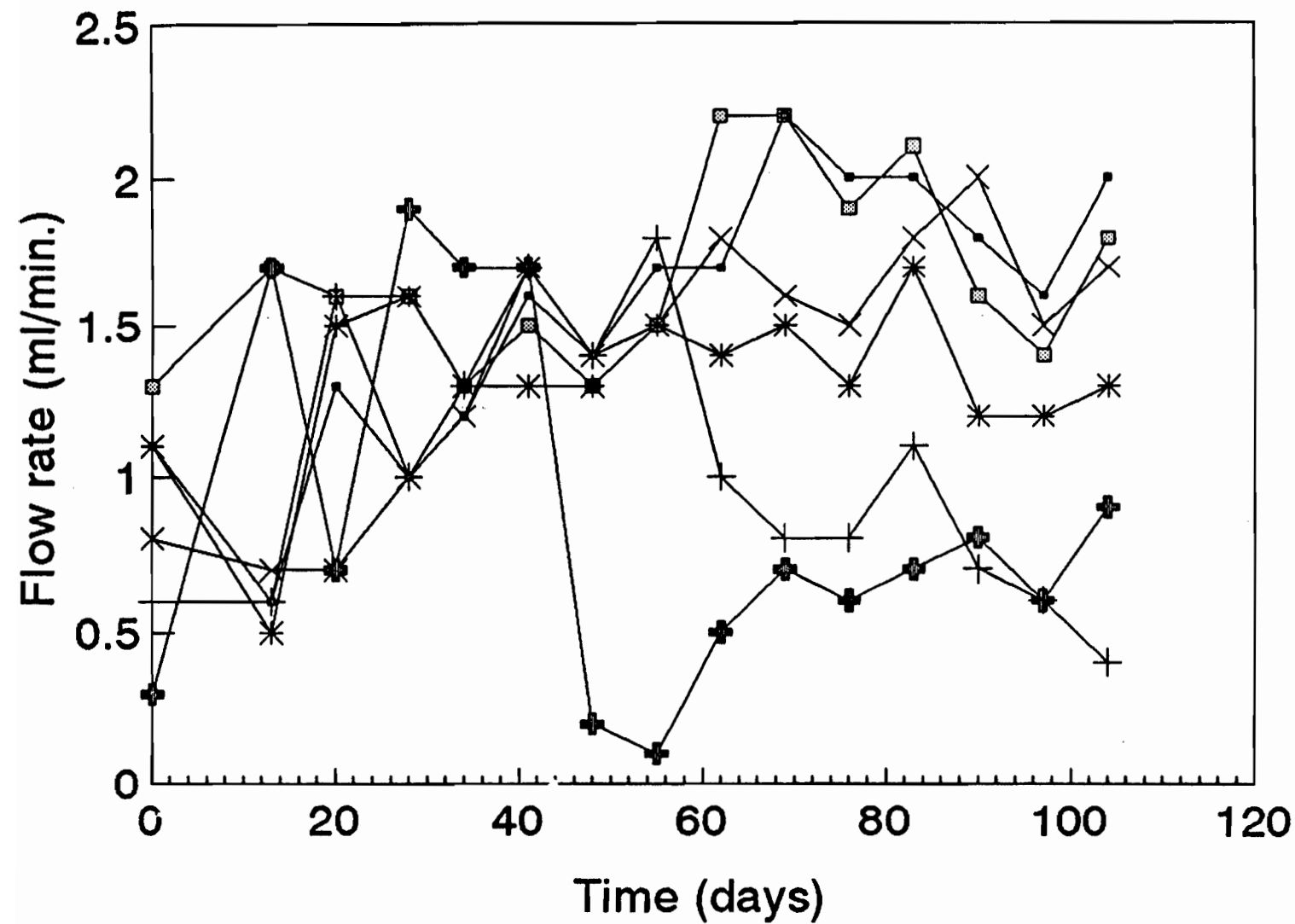


Figure 32. Controls flow rate compared to columns 13, 14, 15

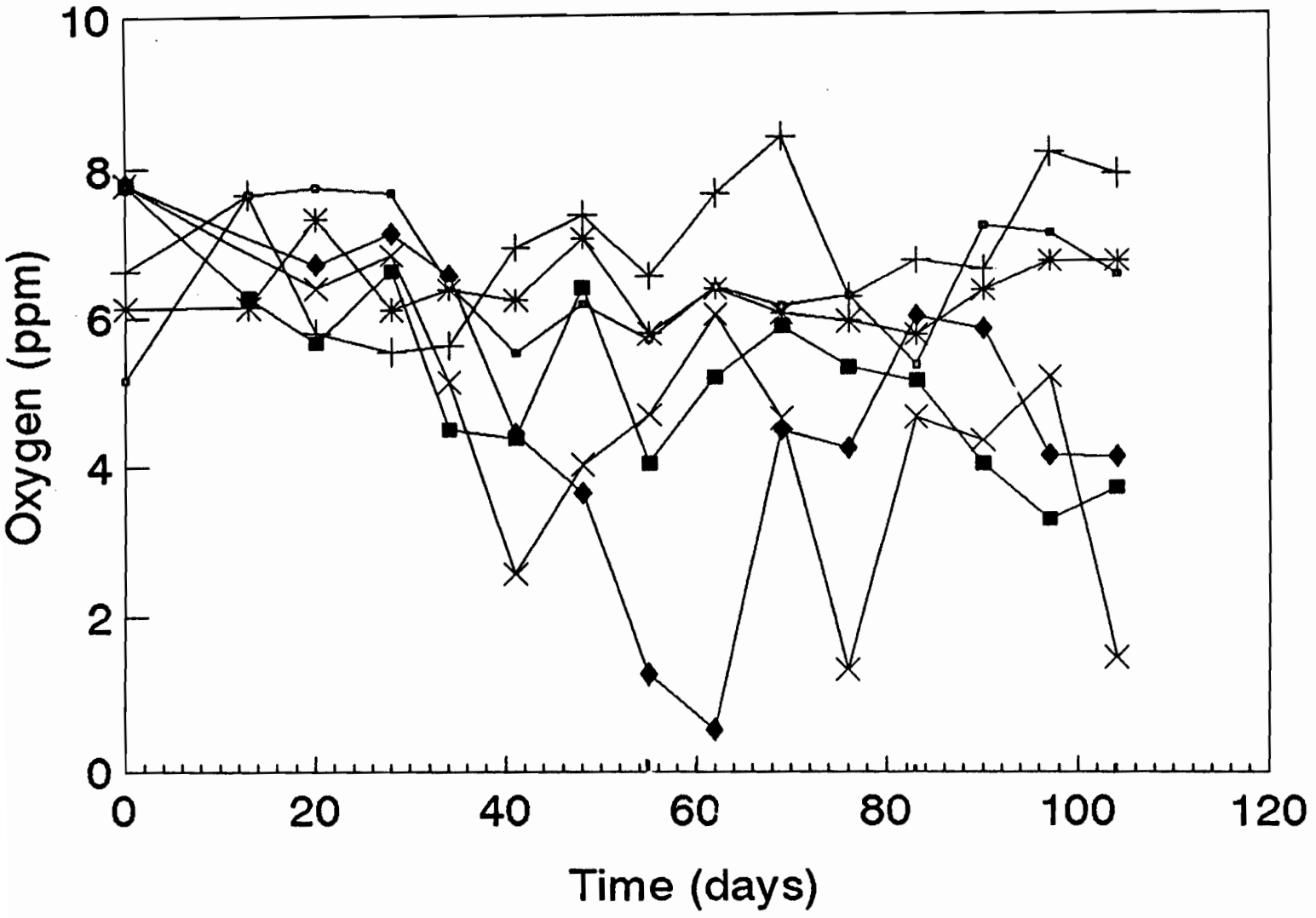


Figure 33. Controls dissolved oxygen compared to columns 4, 5, 6

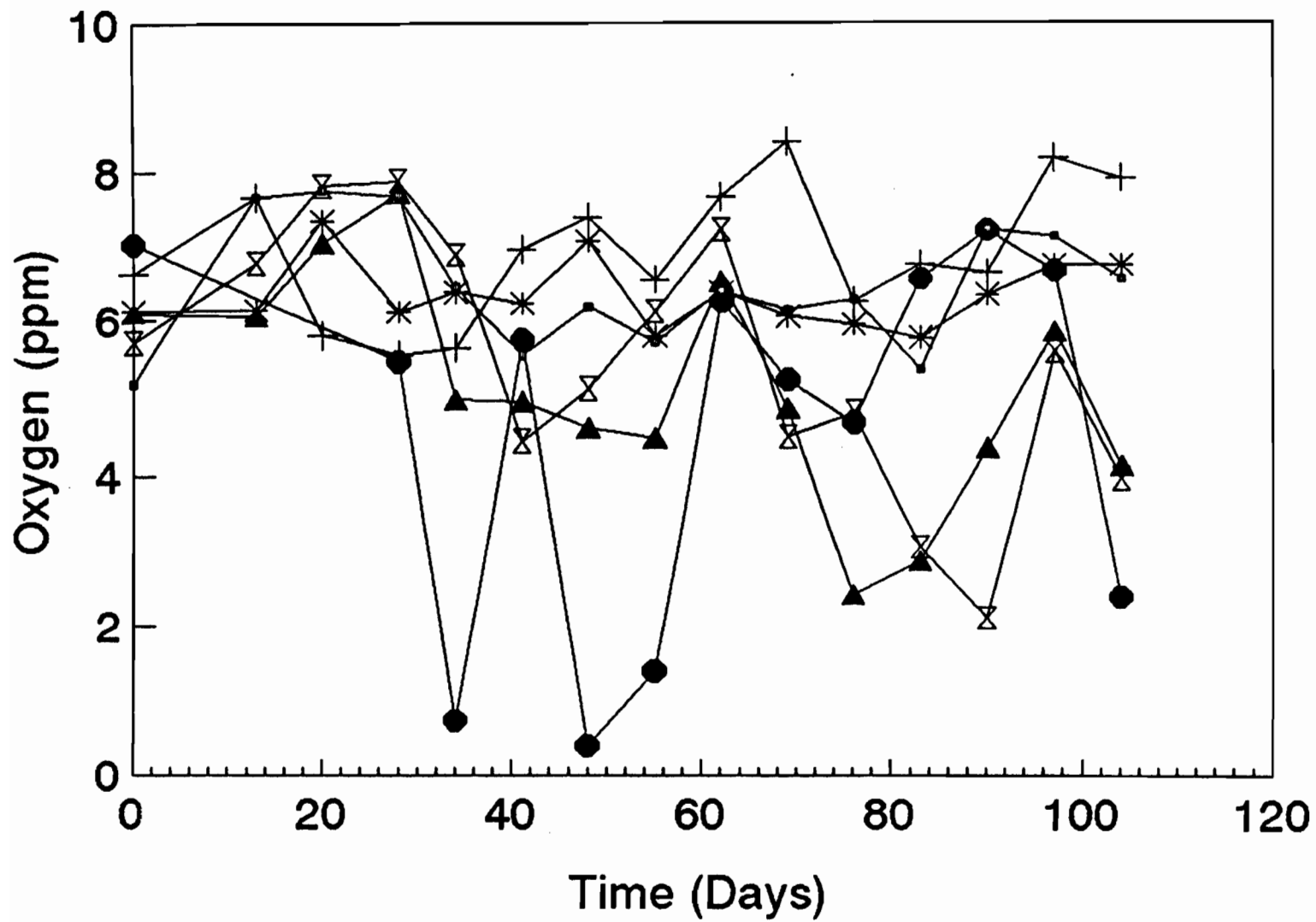


Figure 34. Controls dissolved oxygen compared to columns 7, 8, 9

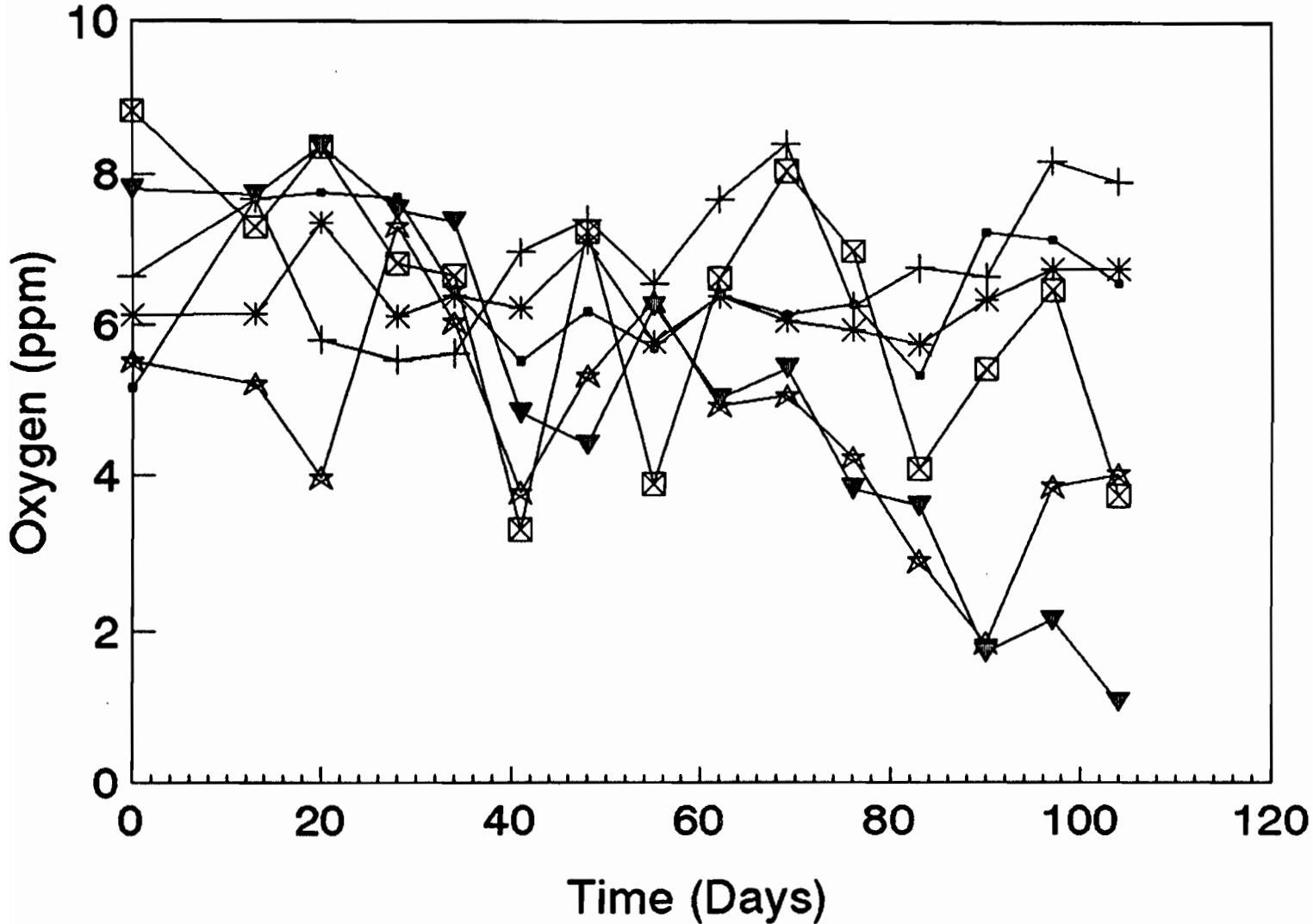


Figure 35. Controls dissolved oxygen compared to columns 10, 11, 12

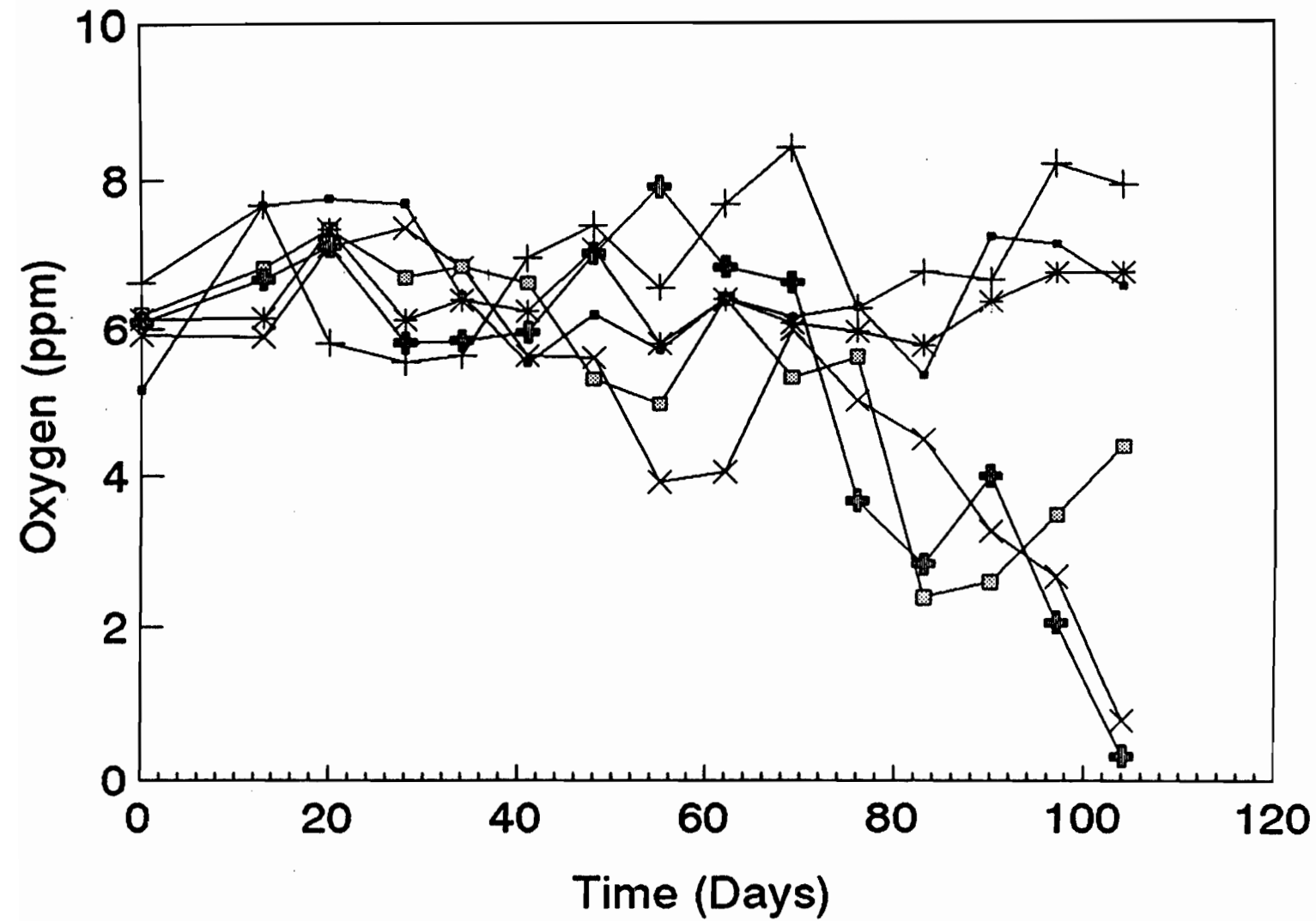


Figure 36. Controls dissolved oxygen compared to columns 13, 14, 15

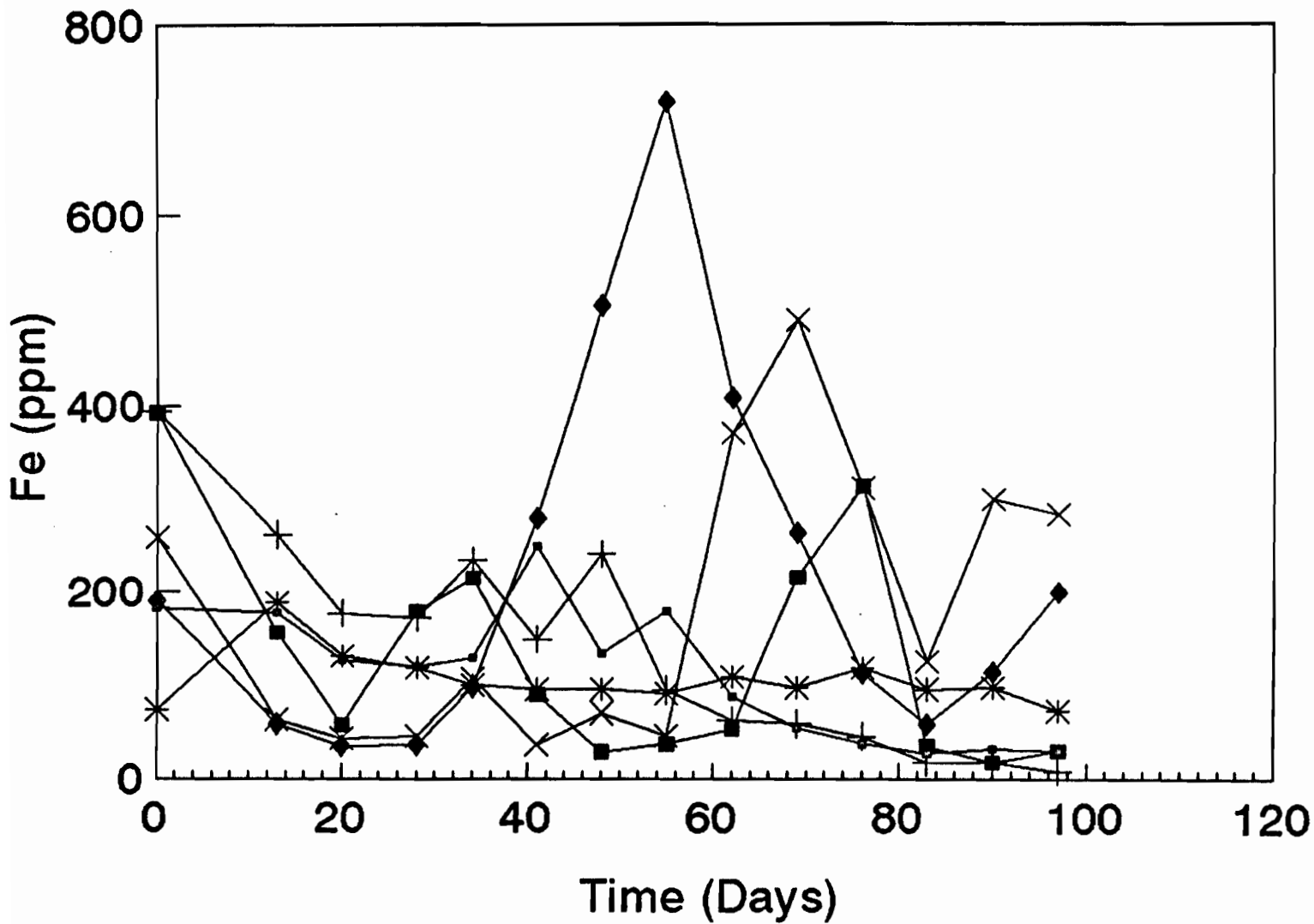


Figure 37. Controls total iron compared to columns 4, 5, 6

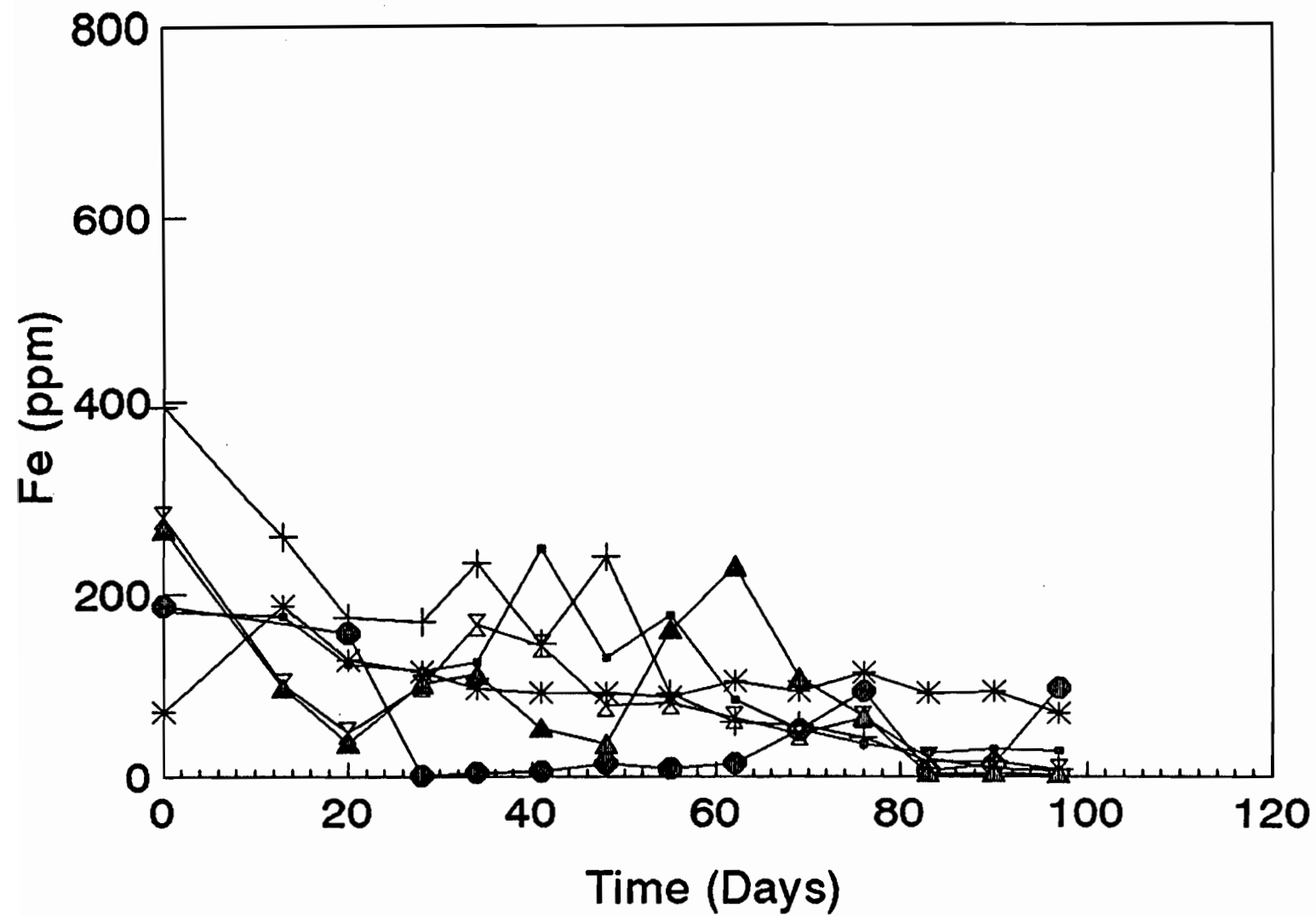


Figure 38. Controls total iron compared to columns 7, 8, 9

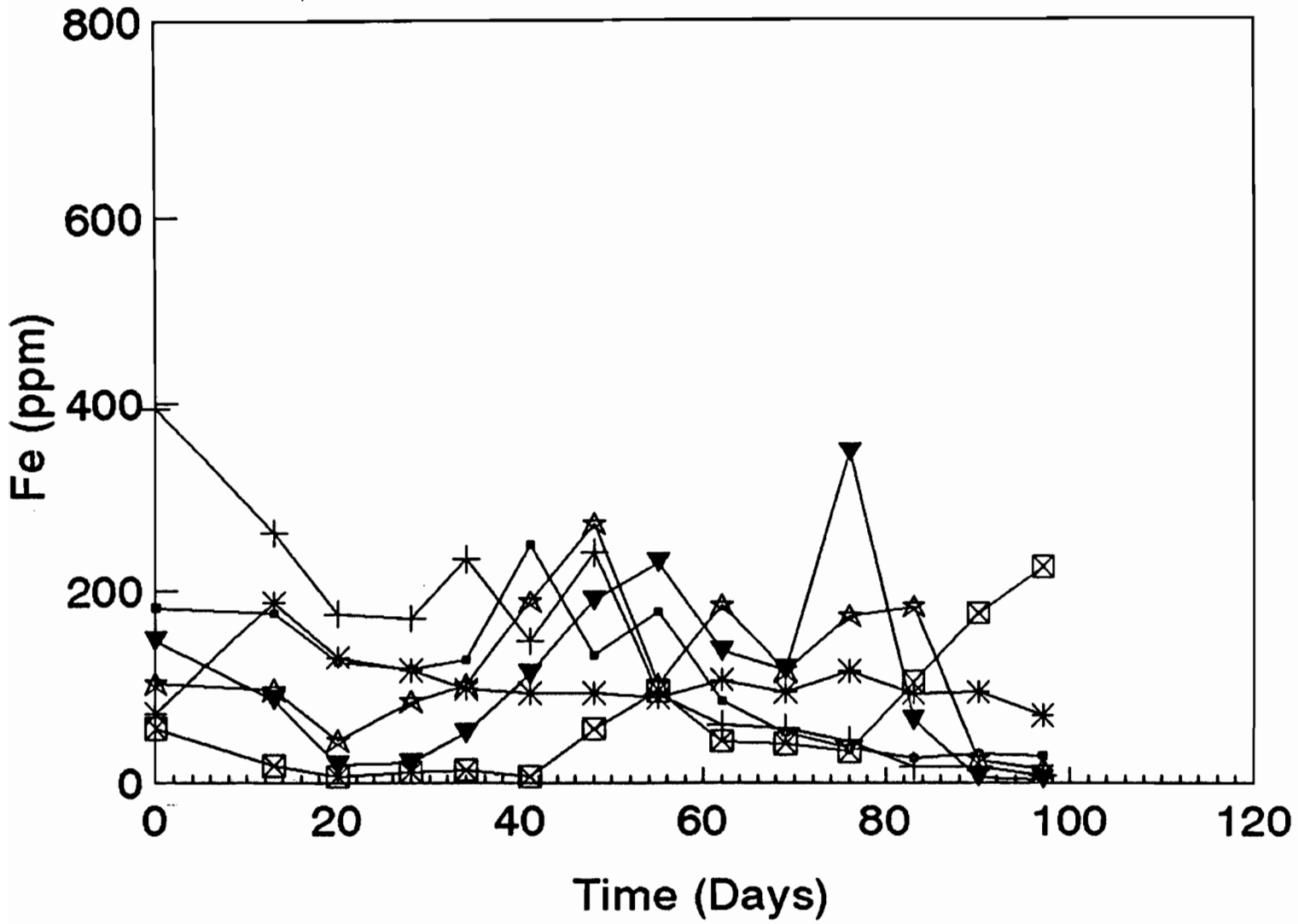


Figure 39. Controls total iron compared to columns 10, 11, 12

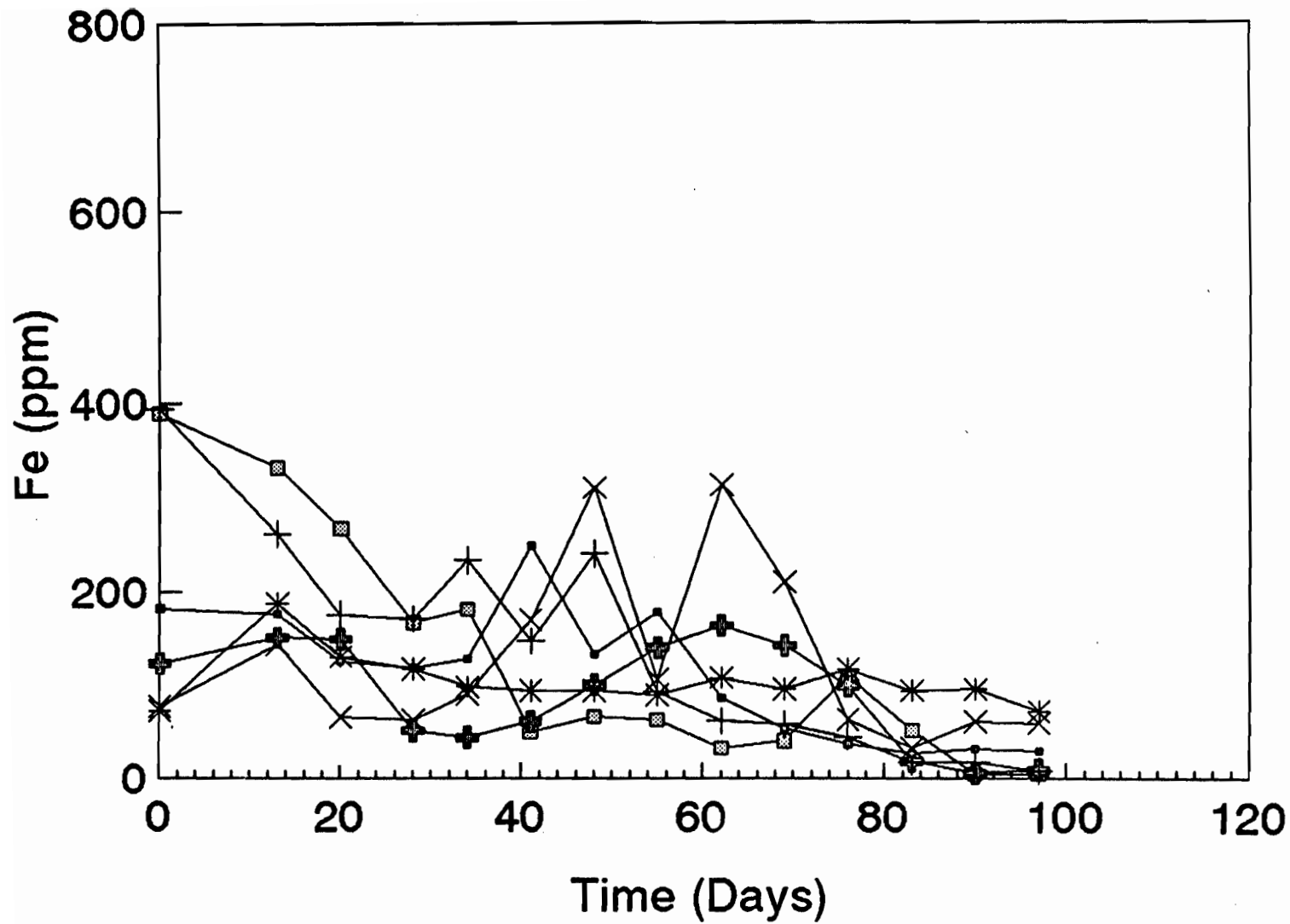


Figure 40. Controls total iron compared to columns 13, 14, 15

APPENDIX - E

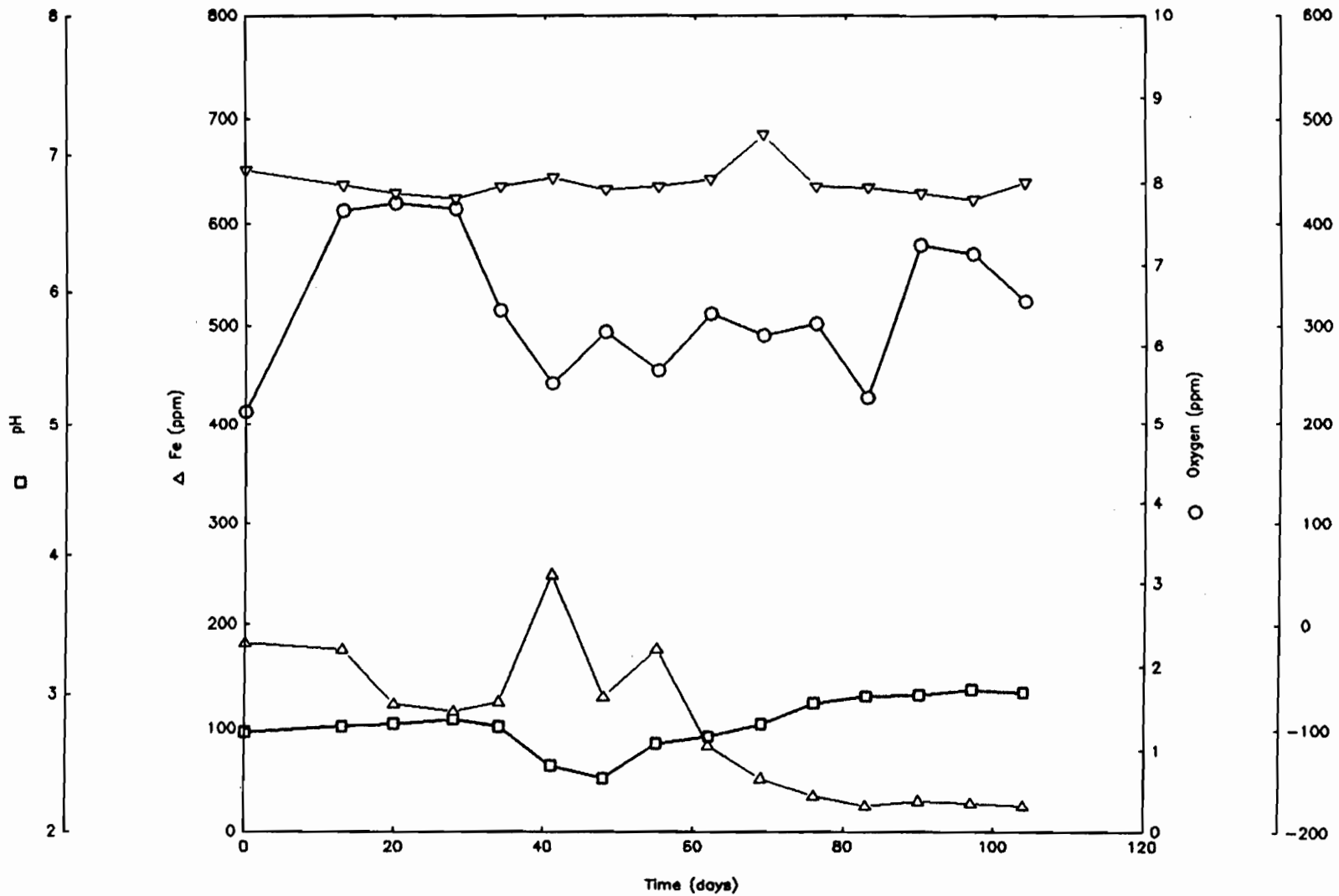


Figure 41. Column 1 pH, total iron, dissolved oxygen and Eh.

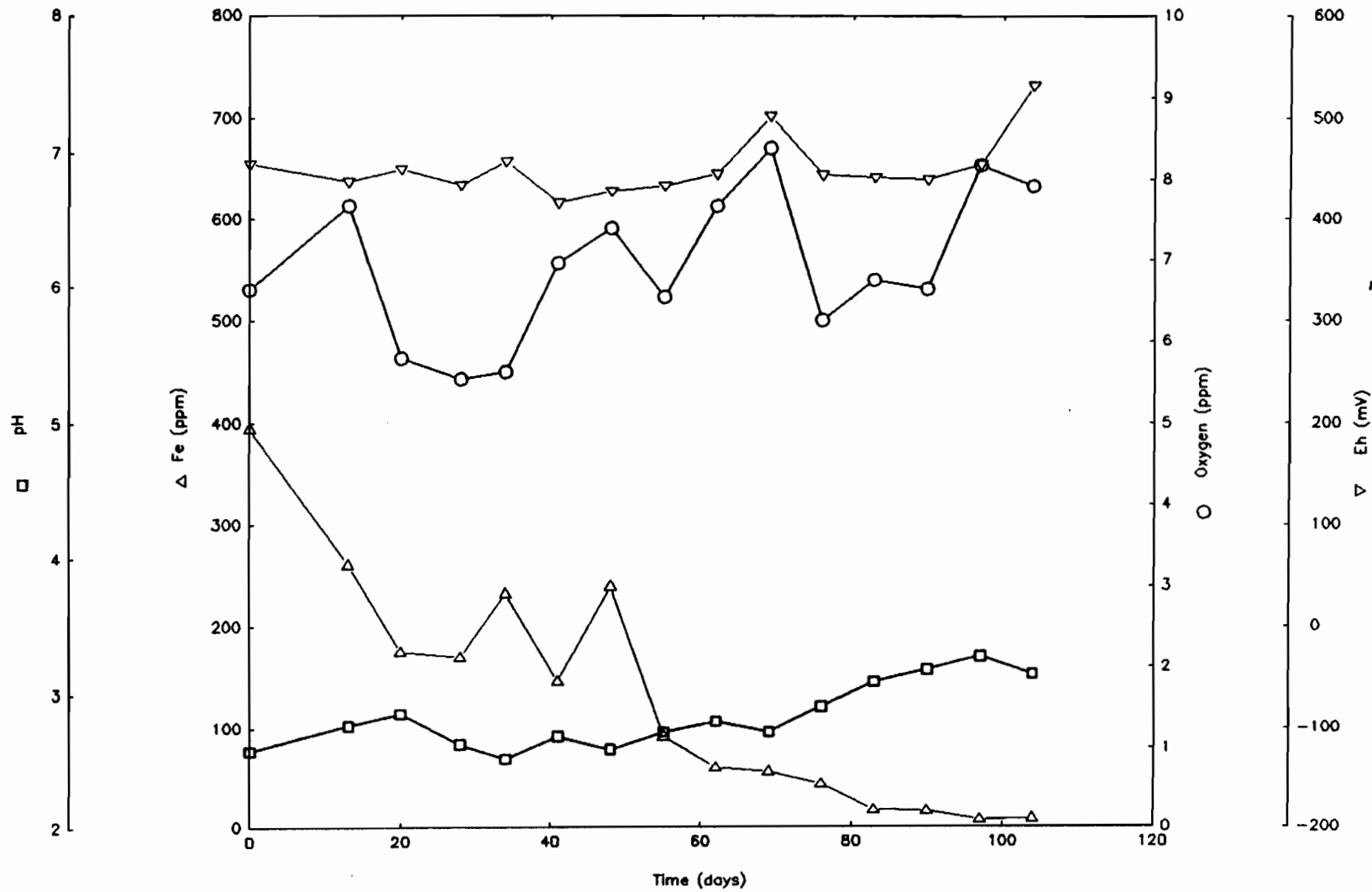


Figure 42. Column 2 pH, total iron, dissolved oxygen and Eh.

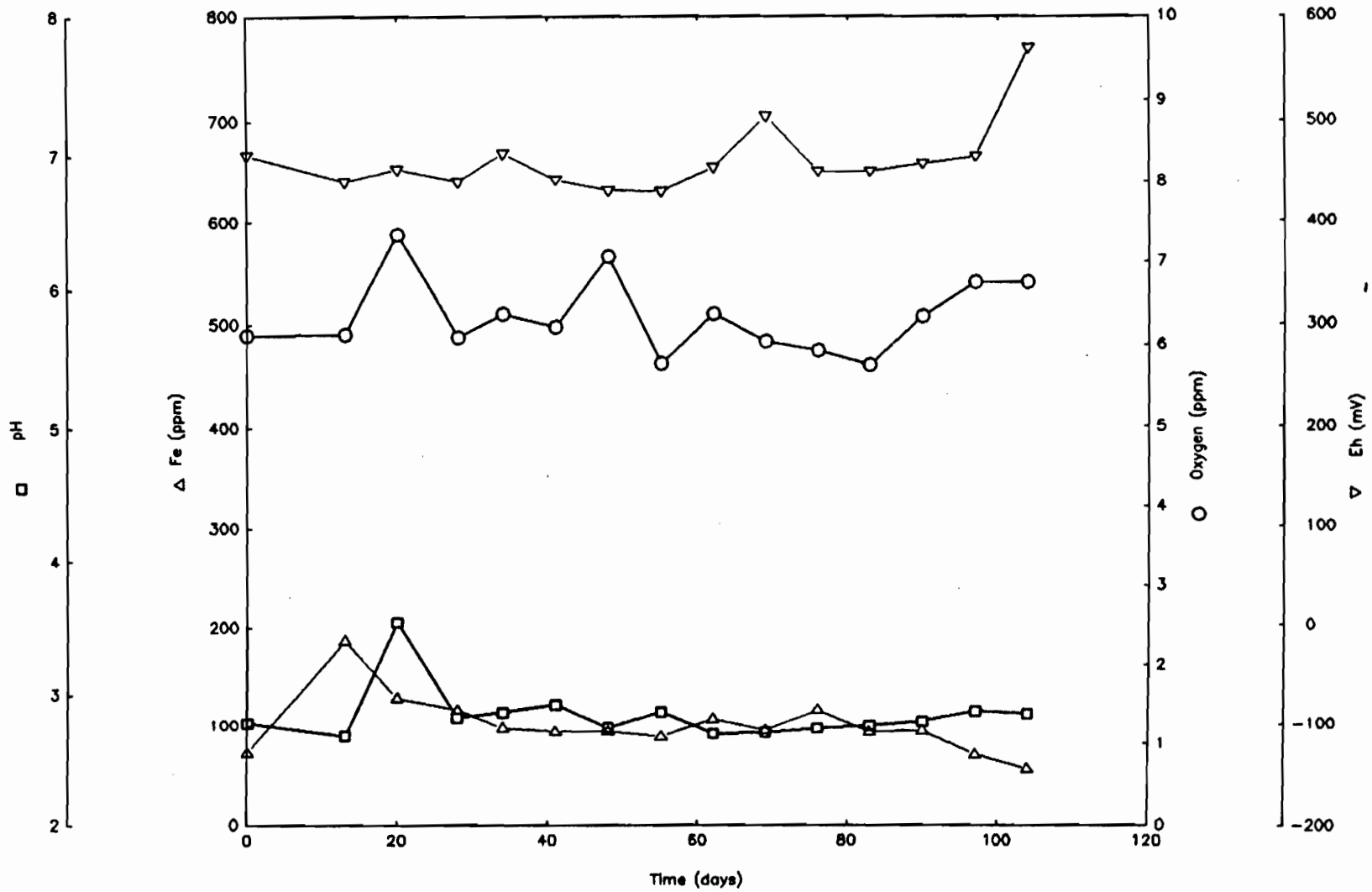


Figure 43. Column 3 pH, total iron, dissolved oxygen and Eh.

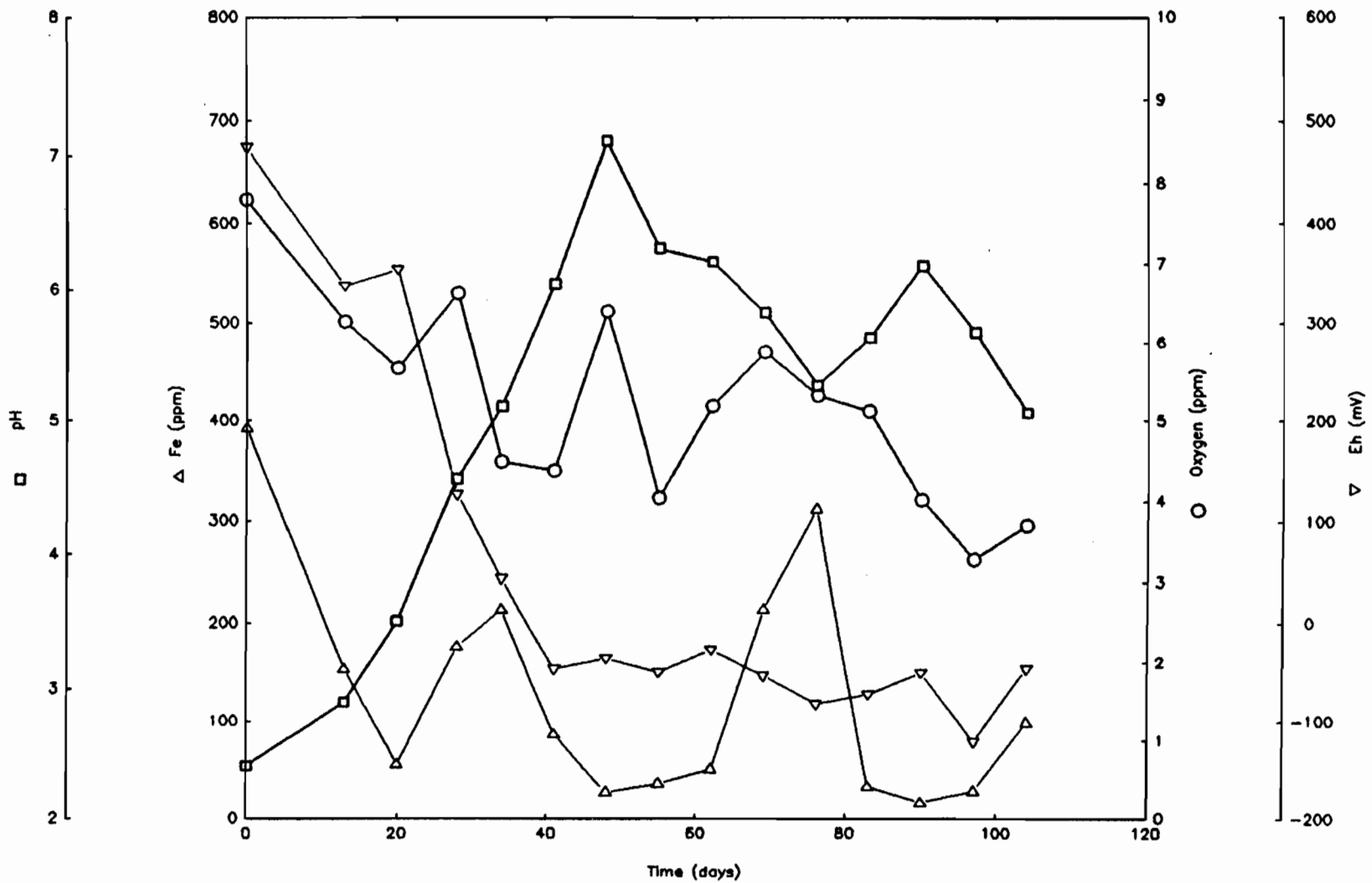


Figure 44. Column 4 pH, total iron, dissolved oxygen and Eh.

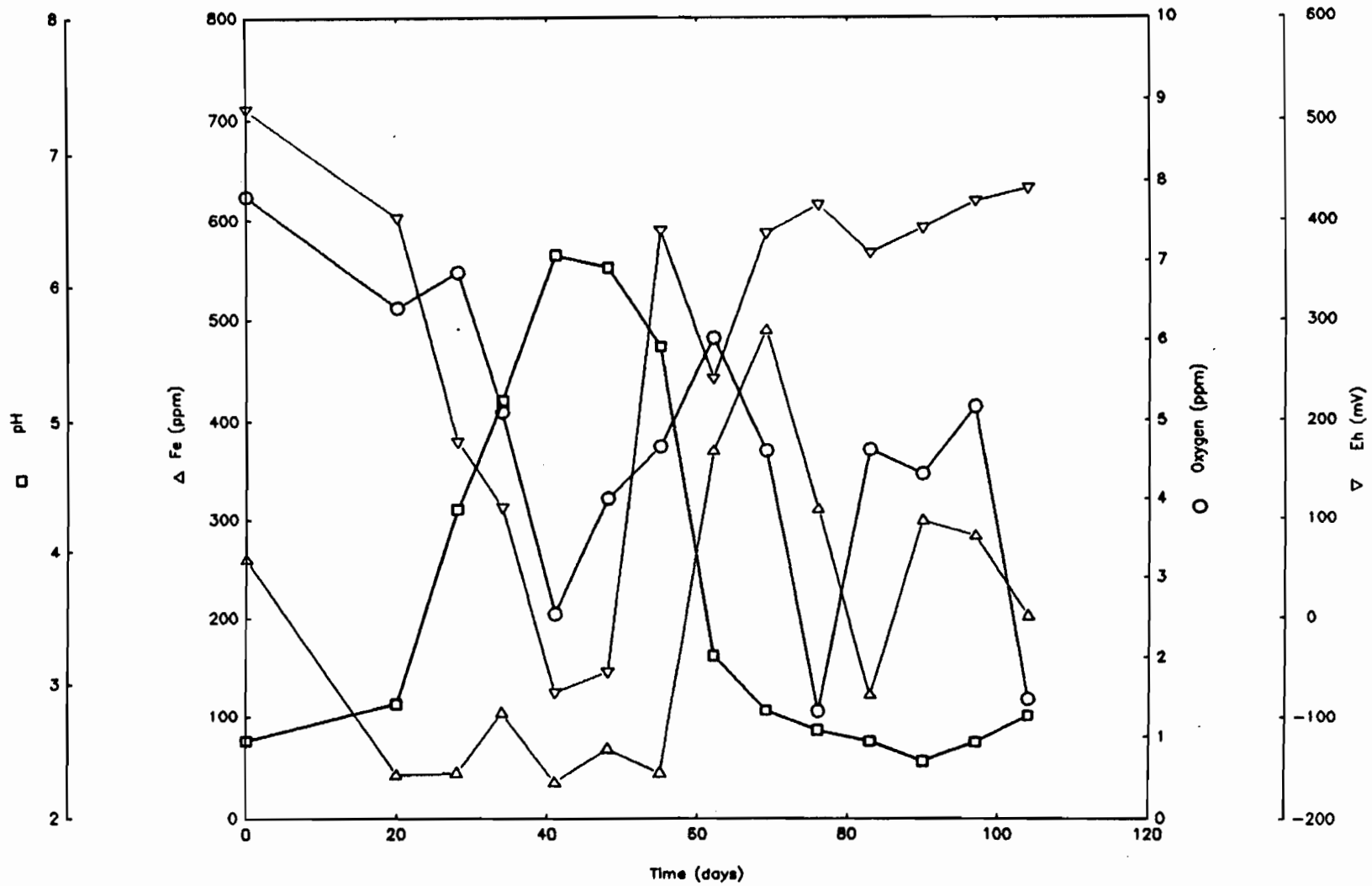


Figure 45. Column 5 pH, total iron, dissolved oxygen and Eh.

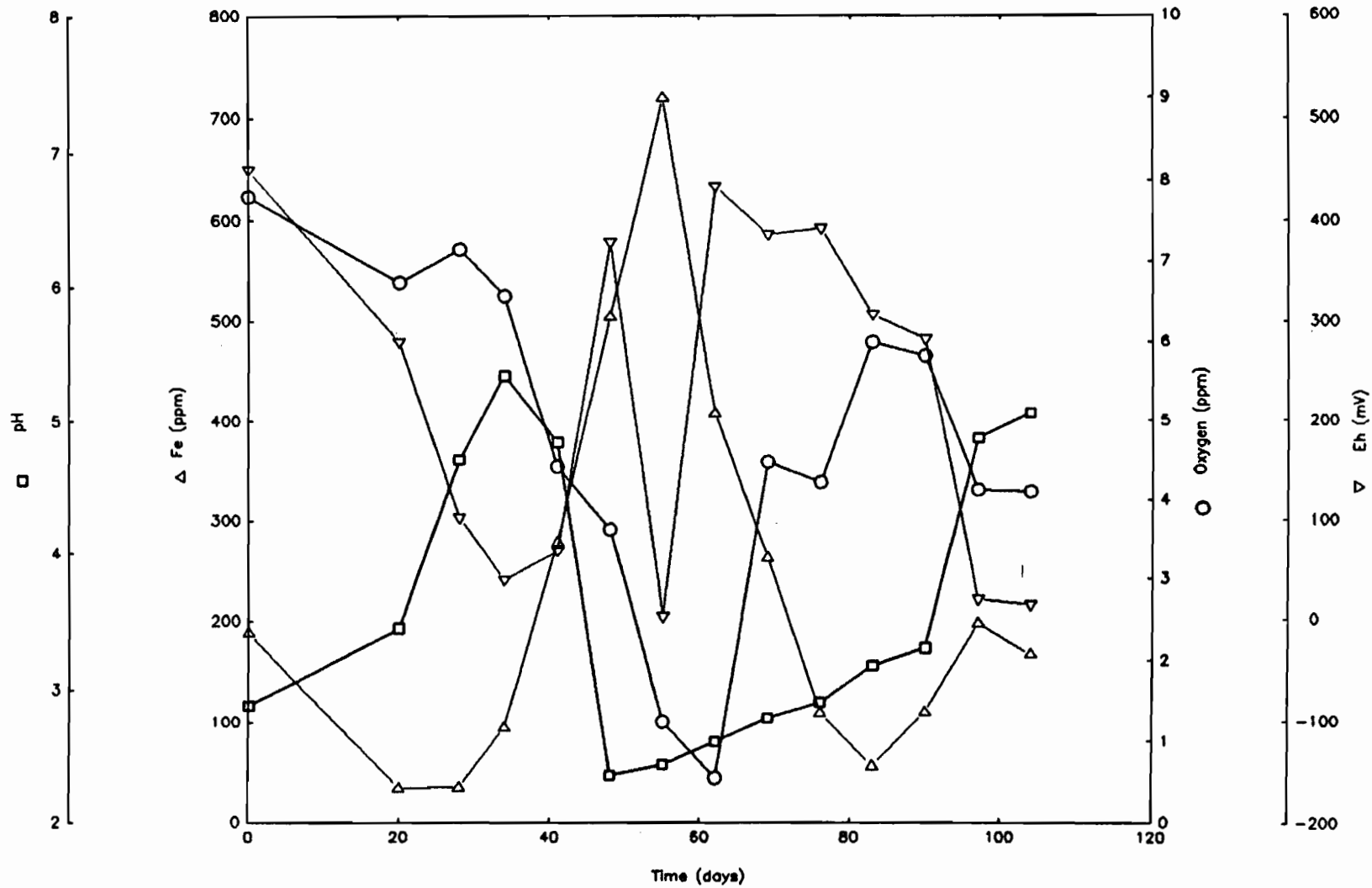


Figure 46. Column 6 pH, total iron, dissolved oxygen and Eh.

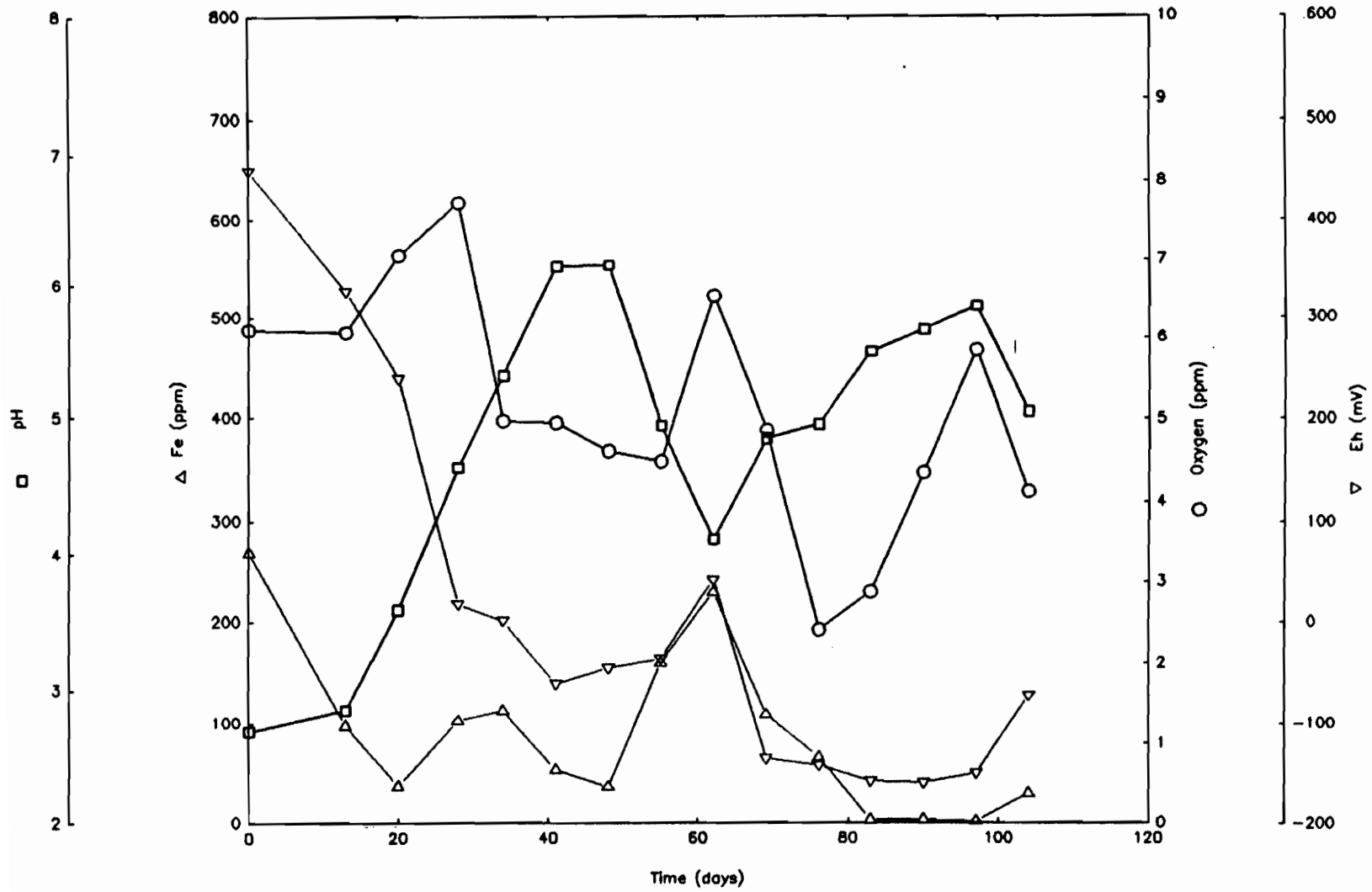


Figure 47. Column 7pH, total iron, dissolved oxygen and Eh.

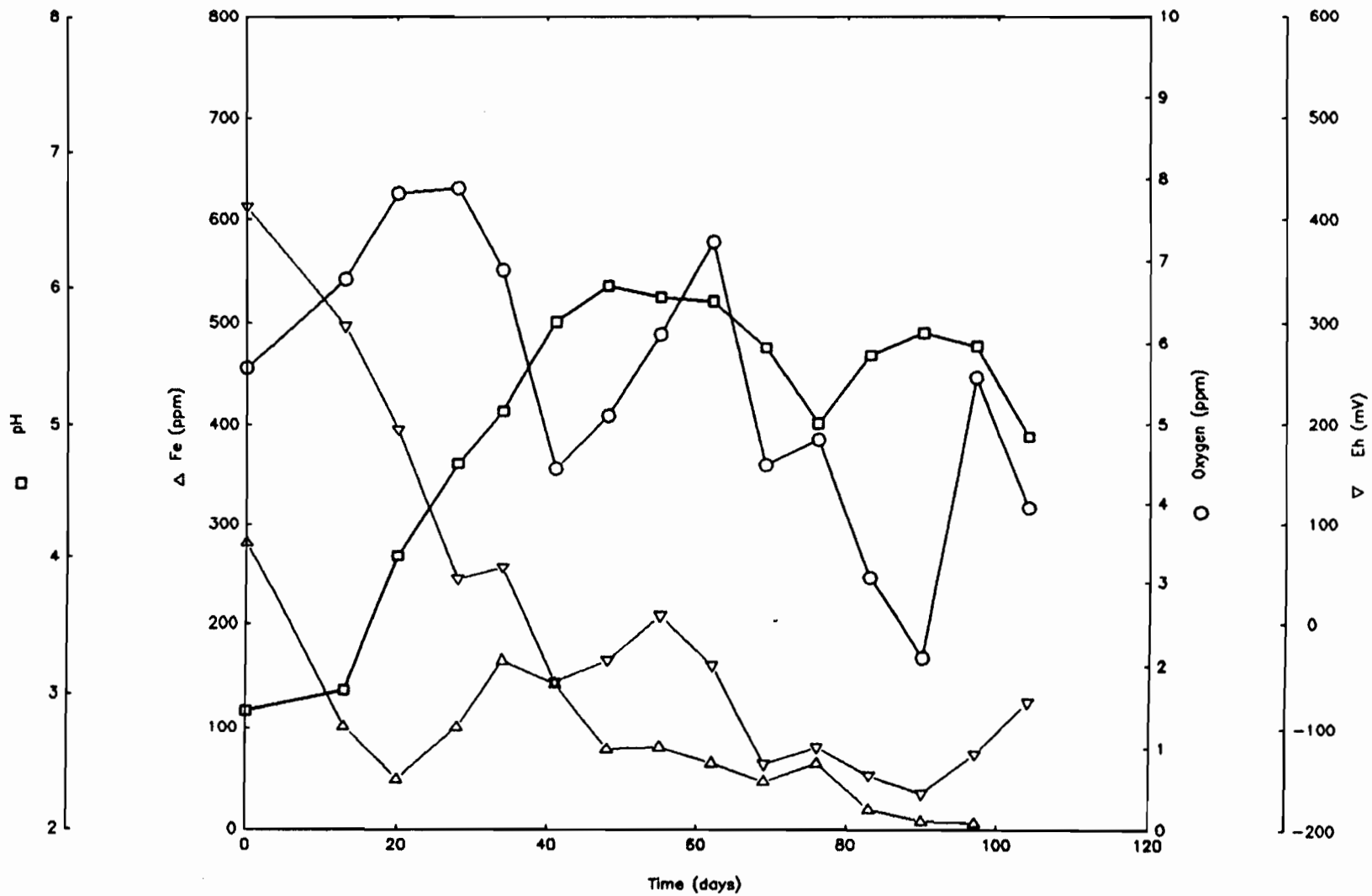


Figure 48. Column 8 pH, total iron, dissolved oxygen and Eh.

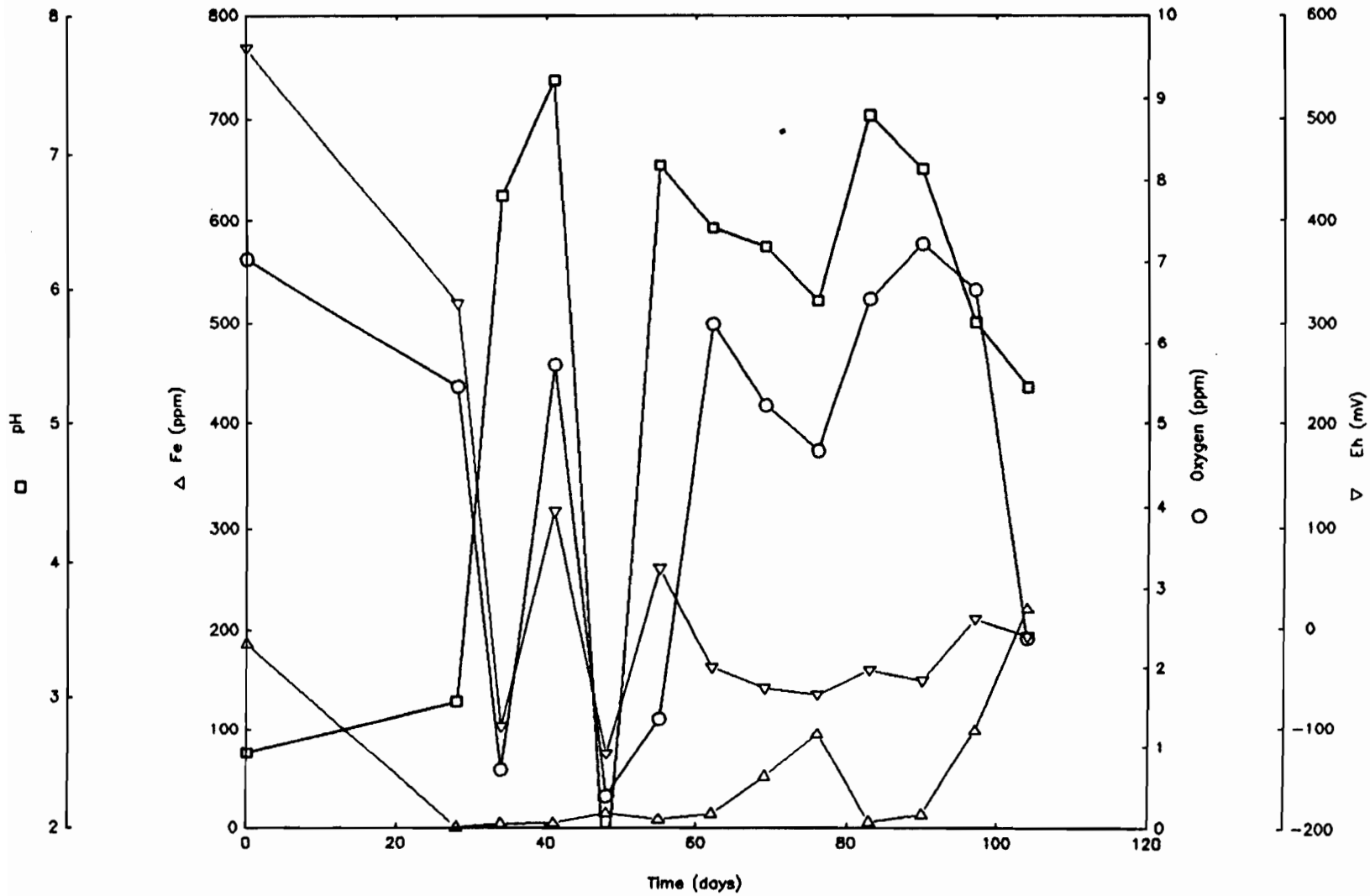


Figure 49. Column 9 pH, total iron, dissolved oxygen and Eh.

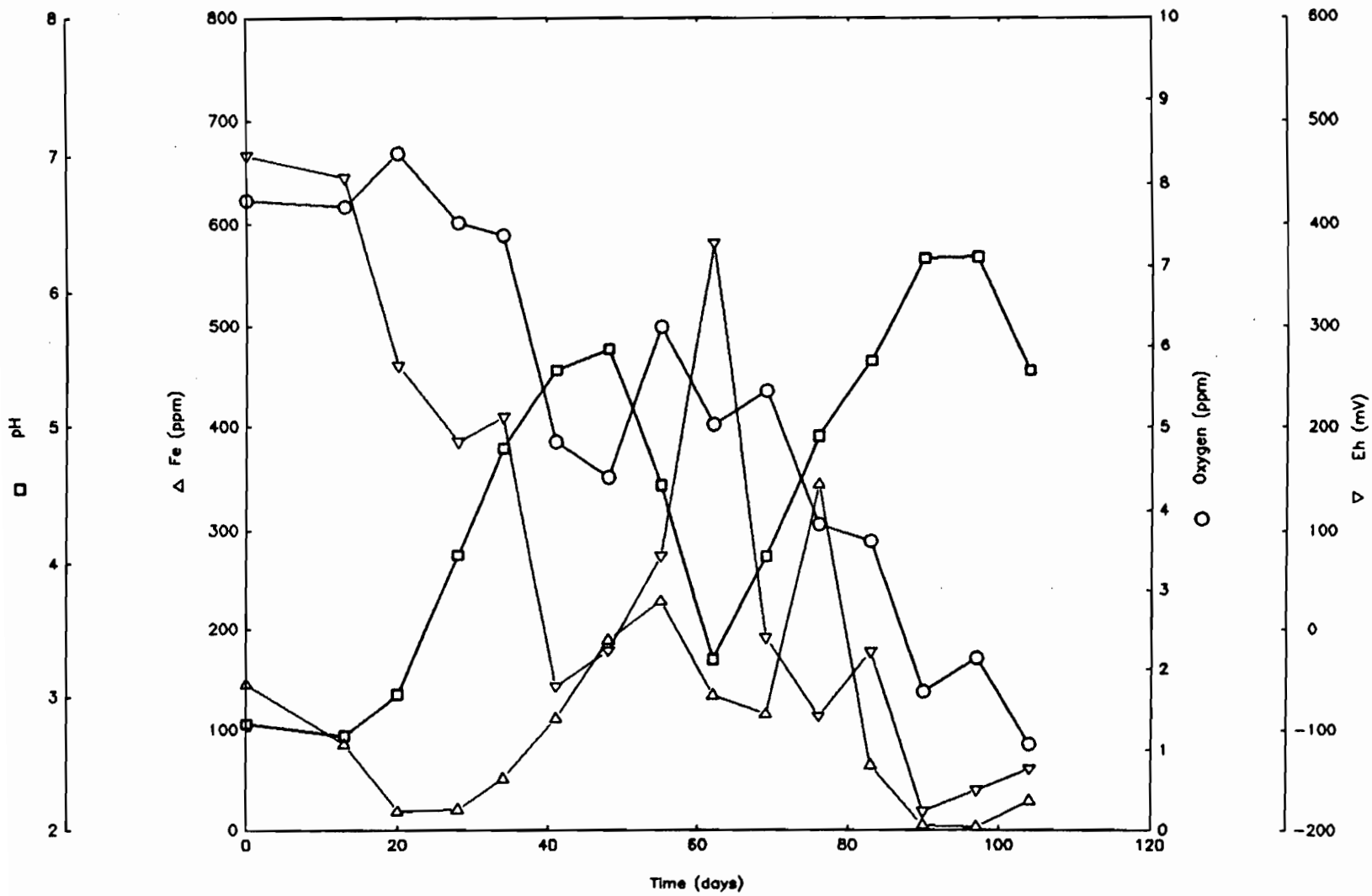


Figure 50. Column 10 pH, total iron, dissolved oxygen and Eh.

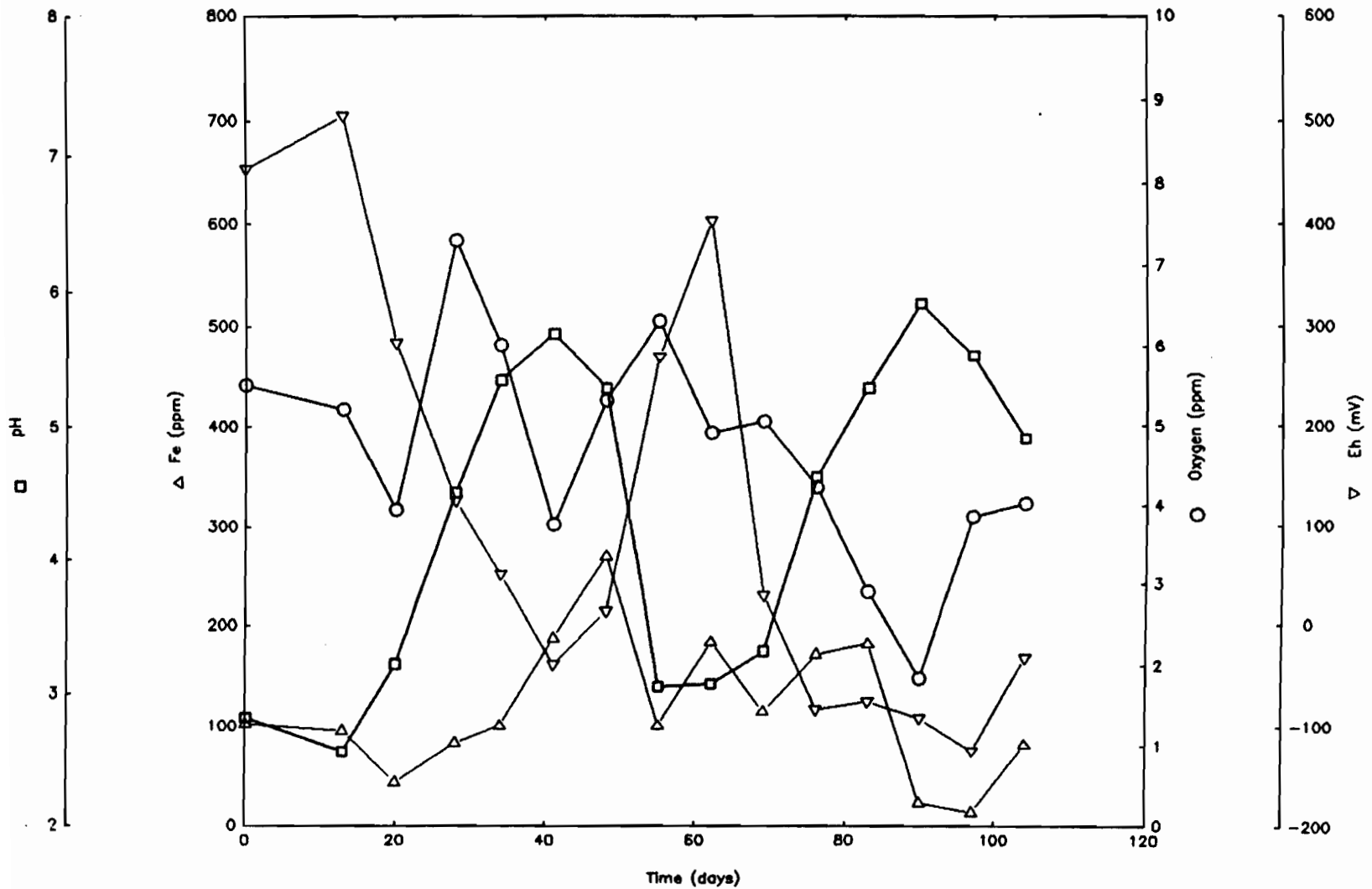


Figure 51. Column 11 pH, total iron, dissolved oxygen and Eh.

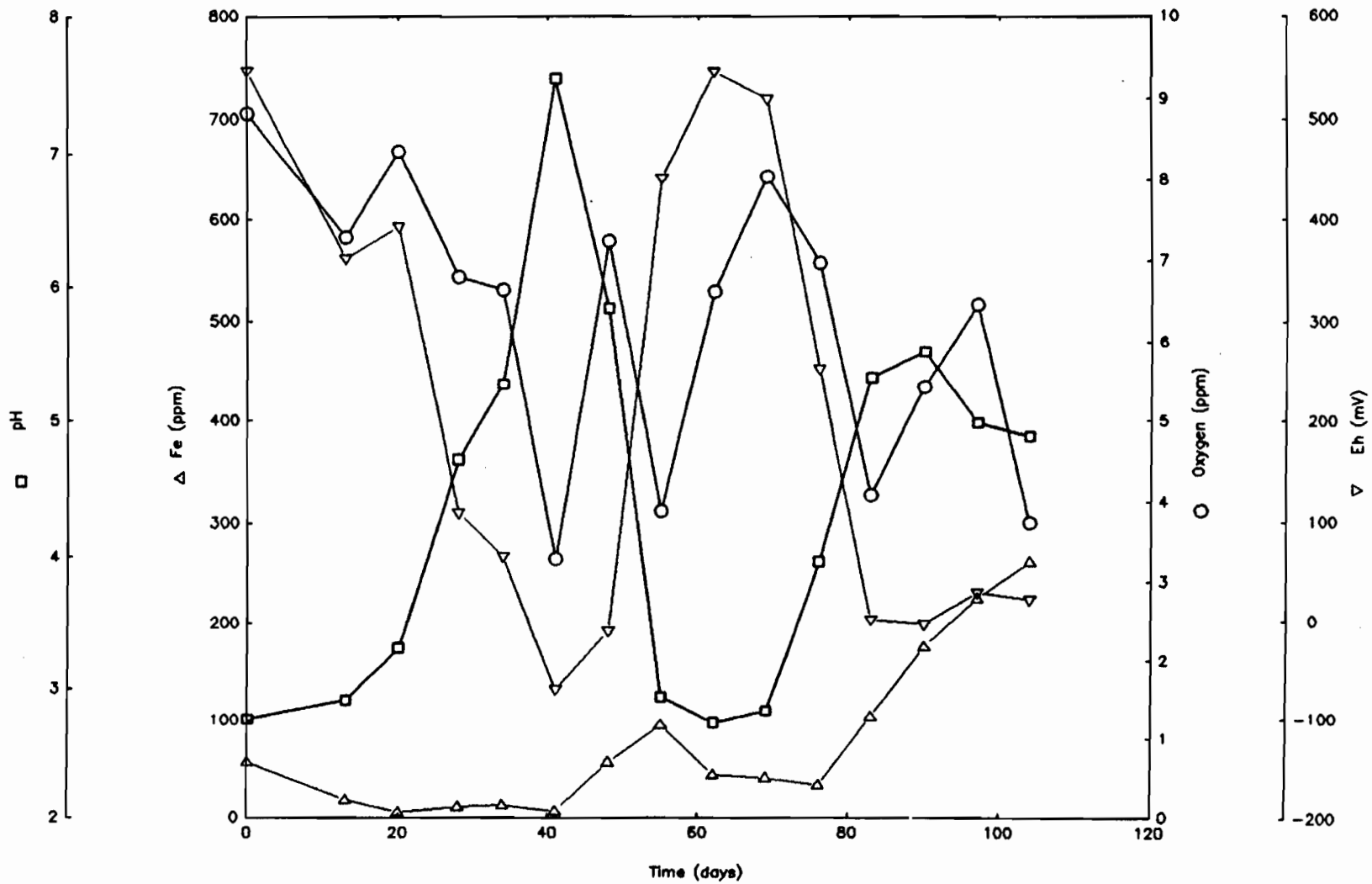


Figure 52. Column 12 pH, total iron, dissolved oxygen and Eh.

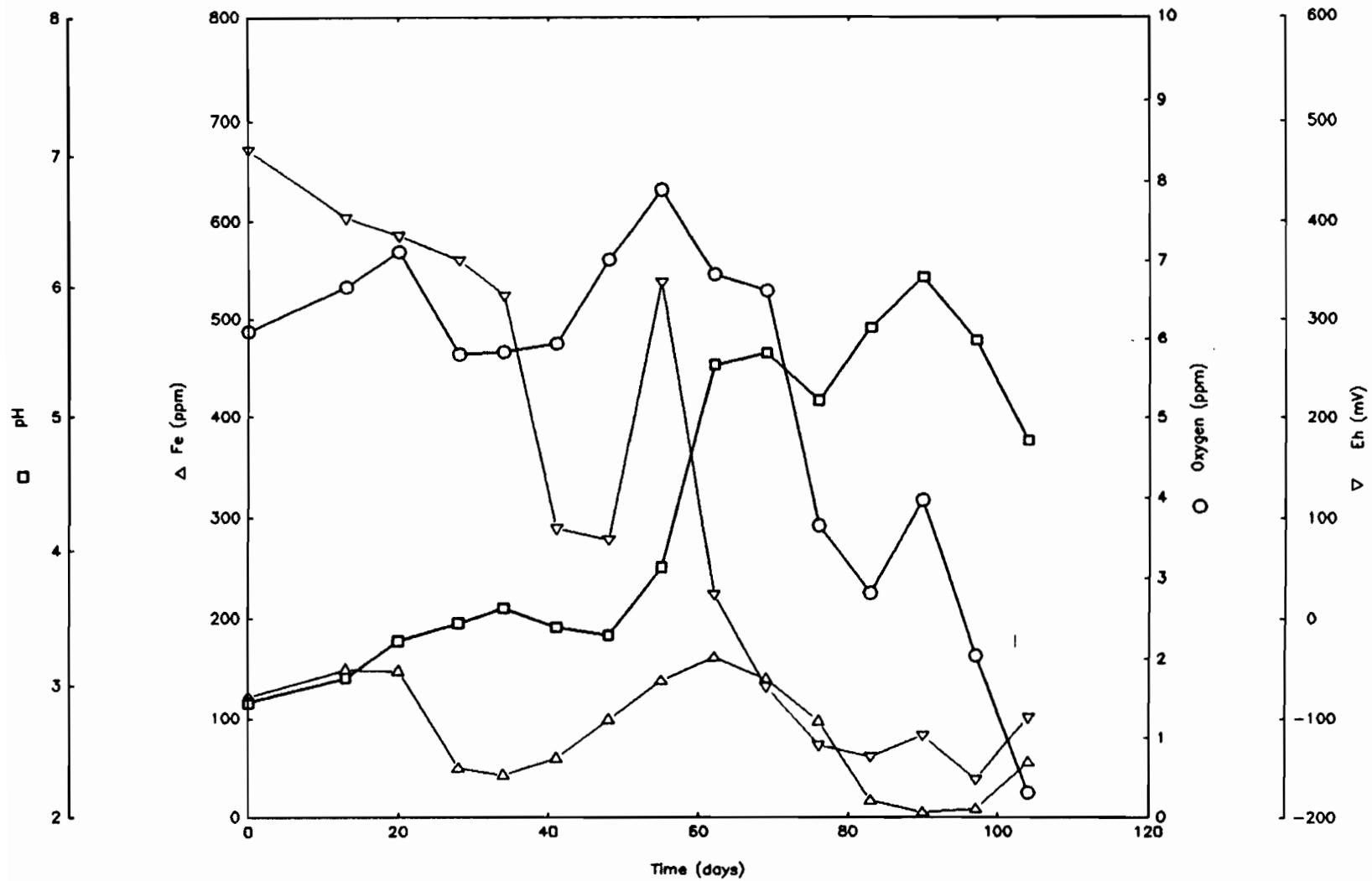


Figure 53. Column 13 pH, total iron, dissolved oxygen and Eh.

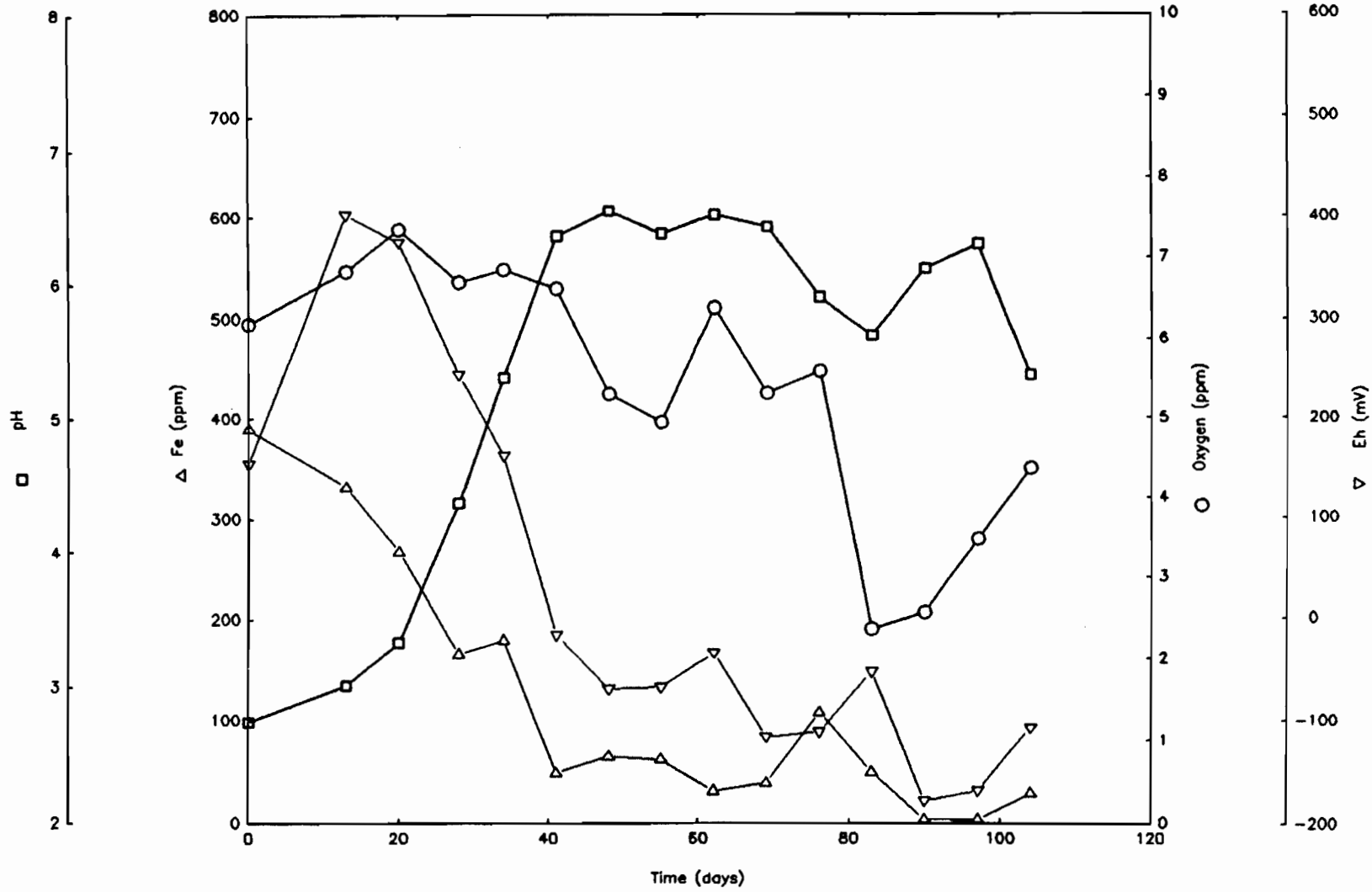


Figure 54. Column 14 pH, total iron, dissolved oxygen and Eh.

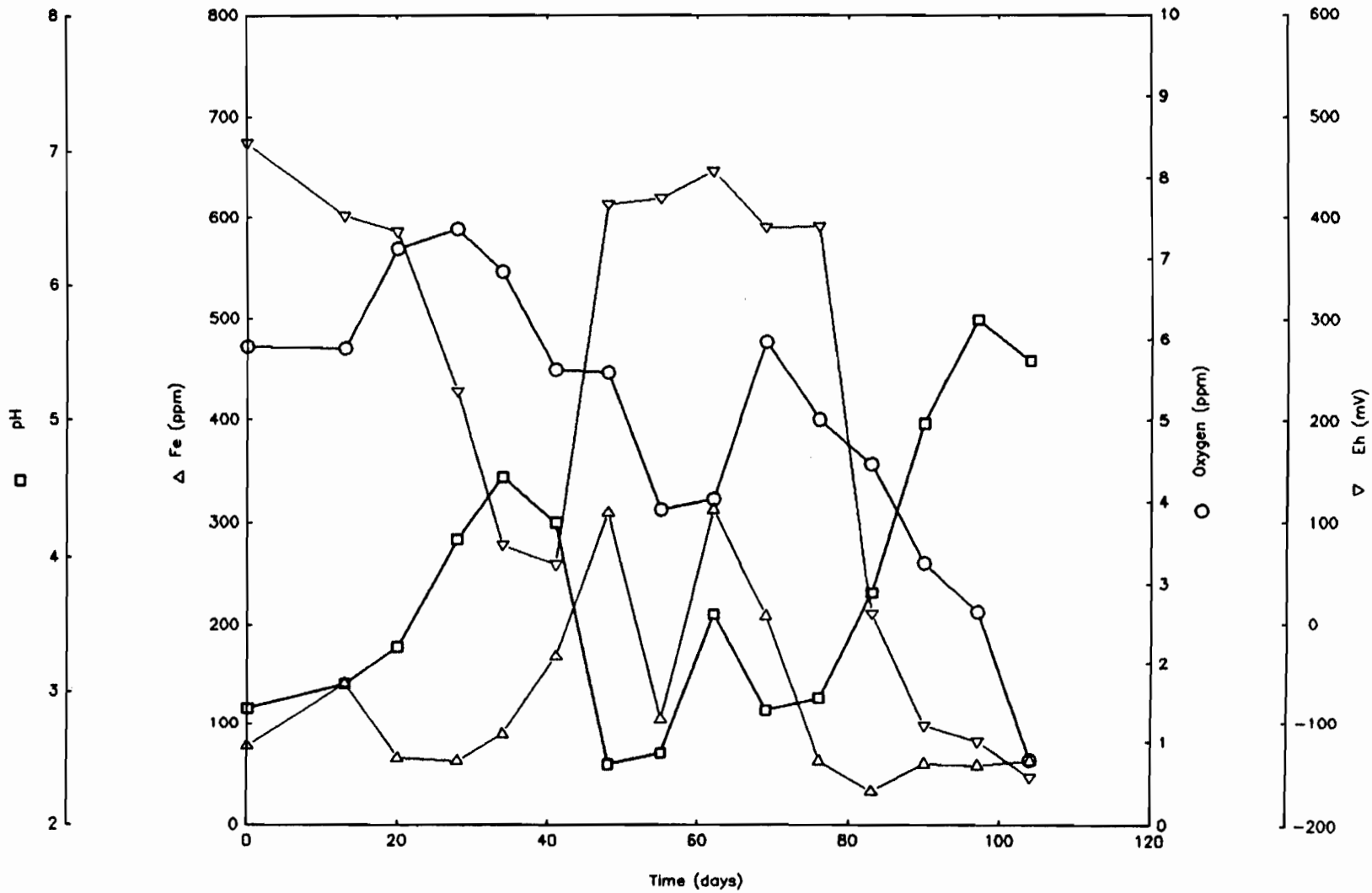


Figure 55. Column 15 pH, total iron, dissolved oxygen and Eh.